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# Kizuna is a novel mitotic substrate for CDC25B phosphatase

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**Keywords:** CDC25B, centrosome, Kizuna, mitosis

**Abbreviations:** Kiz, Kizuna protein; PCM, pericentriolar material; DMSO, dimethyl-sulfoxide; DSP, Dithiobis [succinimidyl propionate].

CDC25 dual-specificity phosphatases play a central role in cell cycle control through the activation of Cyclin-Dependent Kinases (CDKs). Expression during mitosis of a stabilized CDC25B mutant (CDC25B-DDA), which cannot interact with the F-box protein  $\beta$ TrCP for proteasome-dependent degradation, causes mitotic defects and chromosome segregation errors in mammalian cells. We found, using the same CDC25B mutant, that stabilization and failure to degrade CDC25B during mitosis lead to the appearance of multipolar spindle cells resulting from a fragmentation of pericentriolar material (PCM) and abolish mitotic Plk1-dependent phosphorylation of Kizuna (Kiz), which is essential for the function of Kiz in maintaining spindle pole integrity. Thus, in mitosis Kiz is a new substrate of CDC25B whose dephosphorylation following CDC25B stabilization leads to the formation of multipolar spindles. Furthermore, endogenous Kiz and CDC25B interact only in mitosis, suggesting that Kiz phosphorylation depends on a balance between CDC25B and Plk1 activities. Our data identify a novel mitotic substrate of CDC25B phosphatase that plays a key role in mitosis control.

## Introduction

CDC25 dual-specificity phosphatases govern cell cycle progression notably through the spatially and temporally regulated activation of Cyclin-Dependent Kinase (CDK)/Cyclin complexes.<sup>1,2</sup> In higher Metazoans, each CDC25 isoform (CDC25A, B and C) is generally considered to play a specific role at different phases of the cell cycle.<sup>3</sup> Although there is still some debate over the possible redundancy of the 3 isoforms, particularly based on the observations made in CDC25 knockout mice,<sup>4-6</sup> CDC25B is considered to play a major role in initiating entry into mitosis and is often called the starter of mitosis.<sup>7-9</sup> At the G2/M transition, a pool of CDC25B is phosphorylated and activated by Aurora-A kinase at centrosomes,<sup>10</sup> where the initial activation of CDK1/Cyclin B complexes takes place,<sup>11</sup> suggesting that CDC25B might locally participate in the control of the onset of mitosis. Furthermore, CDC25B also participates in the control of  $\gamma$ -Tubulin localization to the centrosomes, is involved in the centrosome duplication cycle,<sup>12</sup> and regulates proteasome-mediated degradation of centrin 2.<sup>13</sup>

Centrosomes are the major microtubule organizing centers. They are composed of a pair of centrioles surrounded by a network of scaffold proteins called pericentriolar material (PCM). They play a central role during mitosis in guiding spindle formation and therefore in maintaining genomic integrity.<sup>14</sup> Indeed, loss of the correct coordination of centrosome duplication and defects in centrosome function result in the formation of multipolar spindles that can lead to abnormal cell divisions.<sup>15-17</sup> The centrosome cycle (duplication, separation and maturation) is under the control of 2 major types of protein kinases: CDK2 associated with either Cyclin E or Cyclin A<sup>18-20</sup> and the Polo-like kinase family (Plk).<sup>21</sup> Among the Plk family members, Plk4 is considered as a master regulator of centriole biogenesis,<sup>22-24</sup> whereas Plk1 initiates centrosome maturation<sup>25</sup> and participates in the formation of the bipolar mitotic spindle.<sup>26</sup> At the onset of mitosis, centrosome maturation requires a critical reorganization in which the PCM expands through rapid recruitment of additional proteins and is stabilized to form stable spindle poles.<sup>27</sup> Loss of PCM factors, such as Kiz,<sup>28</sup> Astrin,<sup>29</sup> and Tastin,<sup>30</sup> leads to centriole disengagement and

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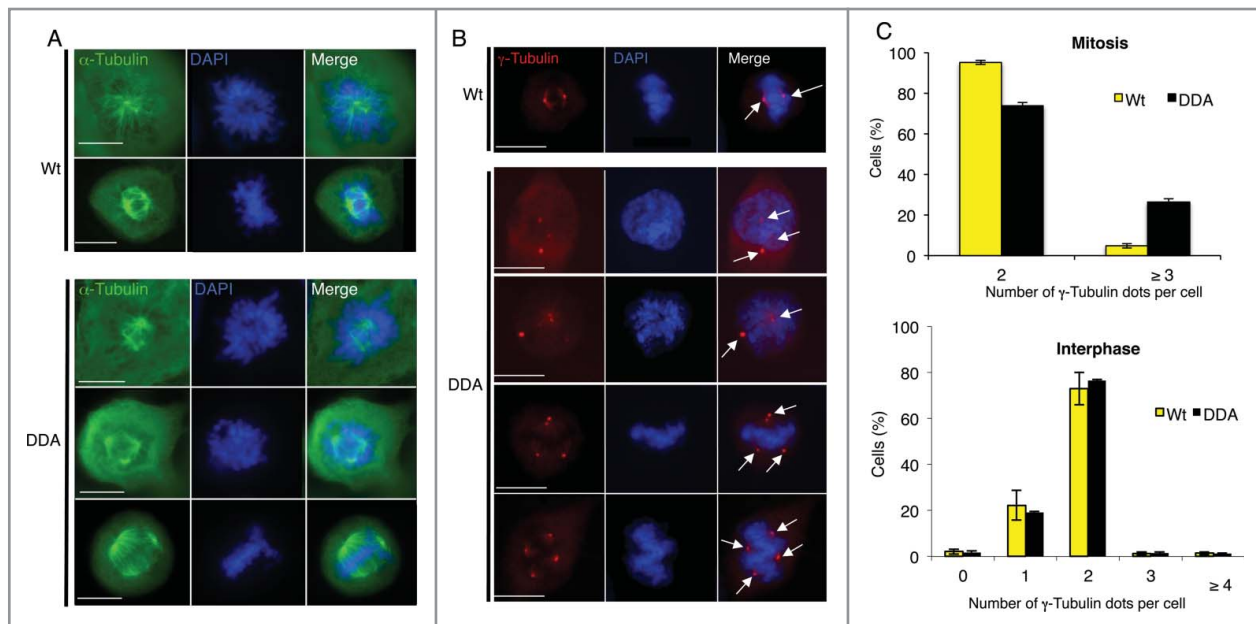
PCM fragmentation. Among these proteins, Kiz plays a critical role in PCM stabilization via its association with Pericentrin. The scaffold Kiz function at spindle poles to prevent PCM fragmentation and the appearance of multipolar spindle depends on its Plk1-mediated phosphorylation on residue Thr379.<sup>28,31</sup>

We previously showed that the expression of a CDC25B mutant (CDC25B-DDA), which cannot interact with the F-box protein  $\beta$ TrCP (the substrate-recognition component of the SCF <sup>$\beta$ TrCP</sup> ubiquitin-ligase complex) and thus is not targeted for proteasome-dependent degradation, stabilizes CDC25B in mitosis. This mutant induces important mitotic defects, as indicated by the appearance of lagging and misaligned chromosomes and the presence of cells with multipolar spindles, without affecting the activity of CDK1/Cyclin B complexes.<sup>32</sup> In this work, we show that the presence of cells presenting multipolar spindles, during mitotic phases, is not due to centrosome amplification but to PCM fragmentation as a consequence of untimed CDC25B-dependent Kiz dephosphorylation. This, in turn, demonstrates that Kiz is a novel substrate for CDC25B.

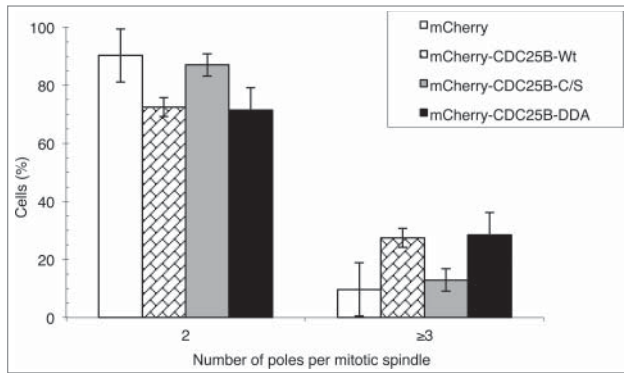
## Results and Discussion

In agreement with our previous results,<sup>32</sup> immunofluorescence analysis with anti- $\alpha$ -Tubulin antibodies showed that the fraction of cells with multipolar spindles, present at different

stages of mitosis, was significantly higher in U2OS Tet-off cells following induced expression of a stabilized CDC25B mutant (CDC25B-DDA) (24%) than a wild-type CDC25B (U2OS-CDC25B-Wt) (3%) (Fig. 1A). As this phenotype is frequently due to amplification of centrosomes,<sup>33</sup> we quantified the number of centrosomes in U2OS-CDC25B-Wt and U2OS-CDC25B-DDA cells by labeling them with anti- $\gamma$ -Tubulin antibodies, a centrosome marker (Fig. 1B-C). More than 95 % of mitotic U2OS-CDC25B-Wt cells (n = 200) had 2  $\gamma$ -Tubulin dots (i.e., bipolar spindles), while 26.3 % of mitotic U2OS-CDC25B-DDA cells (n = 180) displayed more than 2  $\gamma$ -Tubulin dots (i.e., multipolar spindles) (Fig. 1B and C, upper panel). Conversely, during interphase, the number of centrosomes was comparable in U2OS-CDC25B-DDA, U2OS-CDC25B-Wt (Fig. 1C, lower panel) and parental U2OS cells (data not shown). Indeed, only 2.75 % of U2OS-CDC25B-DDA cells in interphase contained more than 2  $\gamma$ -Tubulin dots (i.e., cells with centrosome amplification), a percentage similar to that found in U2OS-CDC25B-Wt cells (2.65 %) (Fig. 1C, lower panel). This finding contradicts a previous report showing that forced expression of CDC25B induces centrosome over-duplication in S phase.<sup>12</sup> This discrepancy could be linked to different levels of ectopic CDC25B expression relative to the endogenous protein. As shown previously,<sup>32</sup> the expression levels of HA-tagged CDC25B-Wt or -DDA in asynchronous stable inducible U2OS cells were similar. However, during mitosis, the level of HA-tagged CDC25B-DDA is higher than that of HA-tagged



**Figure 1.** Stabilization of CDC25B causes centrosome aberration only in mitosis. **(A)** Twenty-four hours after induction of CDC25B-Wt (Wt) or CDC25B-DDA (DDA) expression in U2OS Tet-off cell lines stably transfected with the respective plasmids, asynchronous cells were fixed, and incubated with an anti- $\alpha$ -Tubulin antibody (green). DNA was stained with DAPI. Images are from fluorescence microscopy of prophase and metaphase cells. Scale bar: 10  $\mu$ m. **(B)** Experiments were performed as in A, except that cells were stained with an anti- $\gamma$ -Tubulin antibody (red). Positive  $\gamma$ -Tubulin dots are highlighted by arrows. Images are from fluorescence microscopy of representative prophase and metaphase cells (3 independent experiments). Scale bar: 10  $\mu$ m. **(C)** Quantification of the number of  $\gamma$ -Tubulin-positive dots per cell in mitotic (upper panel) or interphase (lower panel) U2OS cells that express either CDC25B-Wt or CDC25B-DDA. Error bars indicate  $\pm$  SD of 3 independent experiments.

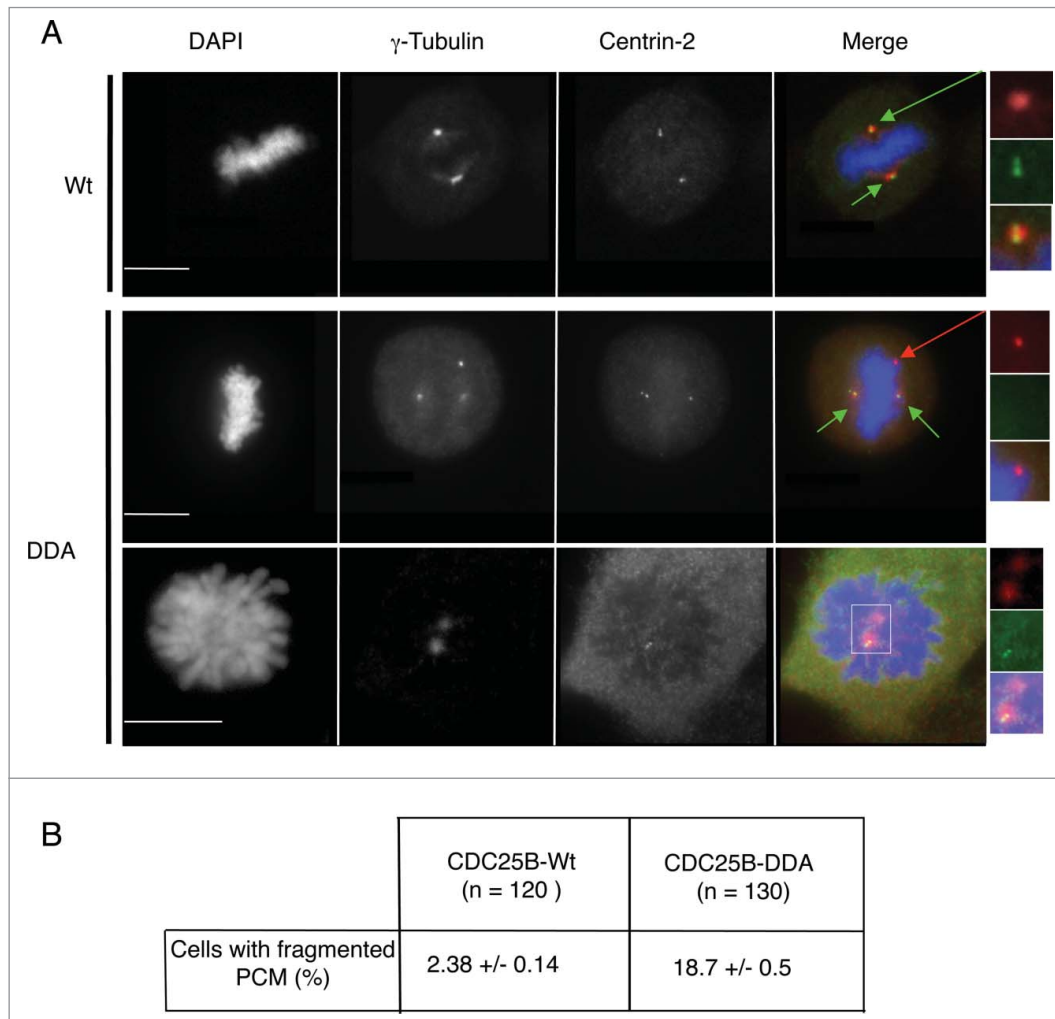


**Figure 2.** CDC25B phosphatase activity is required to induce multipolar spindles in cells. HeLa cells were transiently transfected with pmCherry-CDC25B (-Wt, -C/S or -DDA) or pmCherry expressing vectors. Fourteen hours post-transfection, cells expressing mCherry-tagged CDC25B-Wt, CDC25B-C/S, CDC25B-DDA, or mCherry alone were imaged by time-lapse microscopy for 24 hours. The number of poles per mitotic spindle was counted. Data represent the percentage of cells with 2 or more than 2 poles and values are the mean  $\pm$  SD of 3 independent experiments. Total number of counted cells: mCherry, n = 161; mCherry-CDC25B-Wt, n = 106; mCherry-CDC25B-C/S, n = 126; mCherry-CDC25B-DDA, n = 144.

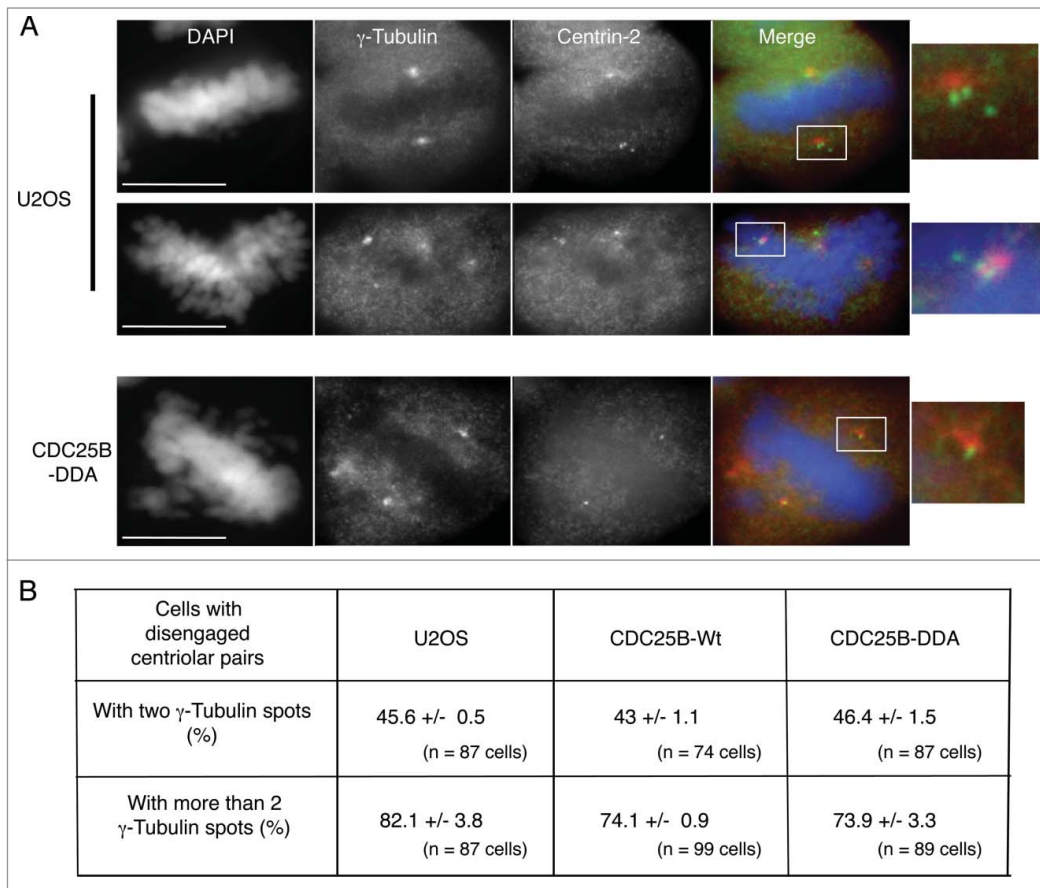
CDC25B-Wt (Fig. 5A). Importantly, while ectopic expression of mCherry-CDC25B-Wt or -DDA (under a strong promoter) induced a similar multipolar spindle phenotype in 27.5 % and 28.5 % of mitotic cells, respectively (Fig. 2), overexpression of mCherry-CDC25B-C/S, a catalytic inactivated mutant of CDC25B, did it at a lower percentage (13%) comparable to mCherry alone (13%). These results indicate that the phosphatase activity of CDC25B is absolutely required to induce multipolar spindle.

Since the abnormal number of  $\gamma$ -Tubulin dots was only observed in mitosis, with spindle poles that fragment, a consequence resulting from aberrant splitting and fragmentation of centrioles,<sup>34-35</sup> we co-stained asynchronous U2OS-CDC25B-Wt and U2OS-CDC25B-DDA cells with antibodies against  $\gamma$ -Tubulin and Centrin-2, a centriole marker, to investigate spindle pole composition (Fig. 3). In mitotic U2OS-CDC25B-Wt cells (n = 120),  $\gamma$ -Tubulin and Centrin-2 co-localized (Fig. 3A) in all, but 2.38 % of cells (Fig. 3B). Conversely, in mitotic U2OS-CDC25B-DDA cells (from prophase to metaphase) (n = 130),  $\gamma$ -Tubulin and Centrin-2 dots did not co-localize in 18.7 % of cells (Fig. 3A-B). In some U2OS-CDC25B-DDA cells with more than 2  $\gamma$ -Tubulin dots, the distance between centrin-2

spots was significantly greater than expected for a normal diplosome, a phenotype corresponding to a disengagement of centriolar pairs (Fig. 3A).



**Figure 3.** Multipolar spindles in U2OS-CDC25B-DDA cells are the result of PCM fragmentation. (A). Representative fluorescent microscopy images of asynchronously growing U2OS cells in metaphase or prophase that express CDC25B-Wt or CDC25B-DDA fixed and stained with anti- $\gamma$ -Tubulin and anti-Centrin-2 antibodies. DNA was stained with DAPI. Centrosomes are highlighted by green arrows, and PCM fragmentation by a red arrow (magnified view in insets). In the merged images, yellow shows co-localization of Centrin-2 and  $\gamma$ -Tubulin. Scale bar: 10  $\mu$ m. (B). The number of cells with fragmented PCM (i.e., cells with an excess of  $\gamma$ -Tubulin dots) was quantified in U2OS-CDC25B-Wt and U2OS-CDC25B-DDA cells. Values are the mean  $\pm$  SD of 3 independent experiments.



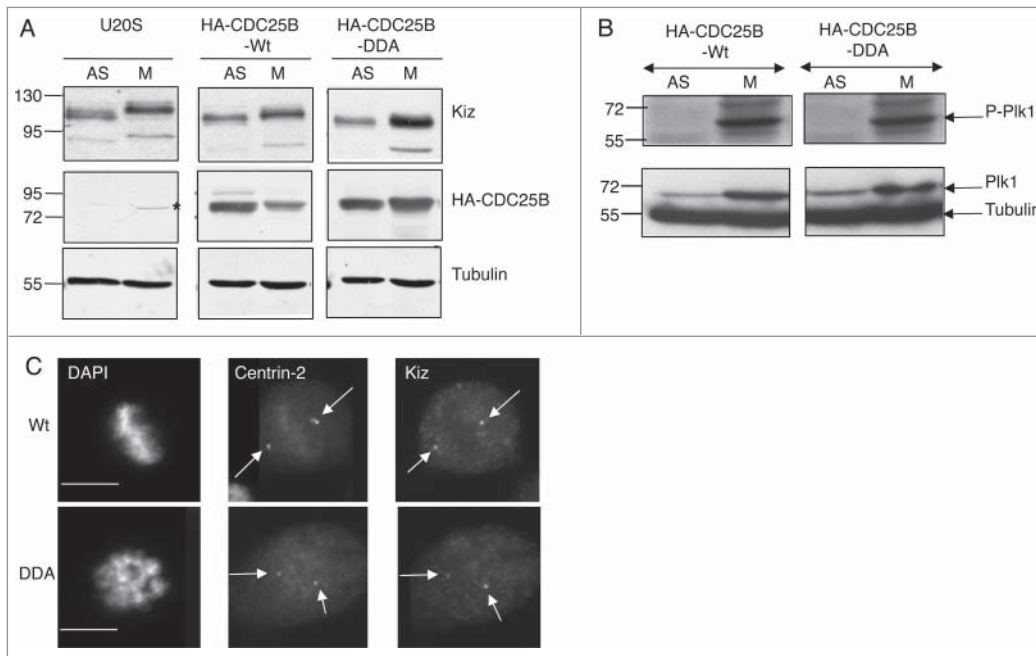
**Figure 4.** Stabilization of CDC25B does not induce a disengagement of centriolar pairs. **(A)** Representative fluorescent microscopy images of mitotic cells from an asynchronous population of parental or CDC25B-DDA U2OS cells fixed and stained with anti- $\gamma$ -Tubulin and anti-Centrin-2 antibodies. DNA was stained with DAPI. In the merged images, yellow shows co-localization of Centrin-2 and  $\gamma$ -Tubulin. Scale bar: 10  $\mu$ m. **(B)** The number of cells with disengaged centriolar pairs (i.e., cells in which the distance between centriols is greater than expected for a diplosome) in cells presenting either 2 or more than 2  $\gamma$ -Tubulin dots was quantified in U2OS, U2OS-CDC25B-Wt and U2OS-CDC25B-DDA cells. Values are the mean  $\pm$  SD of 3 independent experiments.

Moreover only one centrin-2 spot was present on the other pole. Quantification of the proportion of cells with disengaged centriolar pairs showed that it was not an artifact of CDC25B-DDA expression as a similar percentage of cells with this phenotype (Fig. 3A and 4A) was also observed in U2OS-CDC25B-Wt and parental U2OS cells, either in cells containing 2 or more than 2  $\gamma$ -Tubulin dots (Fig. 4B). Similarly, the absence of Centrin-2 spot on the opposite spindle pole was also observed in the same proportion in the 3 cell lines. Taken together, these results demonstrate that the presence of extra  $\gamma$ -Tubulin dots in U2OS-CDC25B-DDA cells (Fig. 3A) is neither a consequence of centrosome over-duplication nor a disengagement of centriolar pairs, but rather the result of PCM fragmentation. The results in Figures 1 and 3 show that high level CDC25B expression (due to its stabilization) during mitosis weakens PCM cohesion, leading to its fragmentation and consequently to the formation of multipolar spindles.

During mitosis, the activity of CDK1/Cyclin B complexes is not affected in U2OS-CDC25B-DDA cells,<sup>32</sup> suggesting that

multipolar phenotype is directly linked to CDC25B. We know that inhibition of Plk1-mediated Kiz protein phosphorylation at Thr379, a phosphorylation that positively controls Kiz function in the stabilization of PCM during mitosis, causes PCM fragmentation.<sup>28</sup> In addition we show that the expression of non-phosphorylatable Kiz mutant (Kiz-T379A) in cells depleted for Kiz, did not rescue PCM fragmentation induced by the depletion of Kiz,<sup>28</sup> but rather increased it at a level comparable to that observed in U2OS-CDC25B-DDA cells (Fig. S1). Thus, we investigated whether CDC25B-DDA could affect Kiz phosphorylation. Kiz electrophoretic mobility was reduced in mitotic parental U2OS and U2OS-CDC25B-Wt cells in comparison to asynchronously growing cells (Fig. 5A), showing that Kiz was phosphorylated in mitosis as previously described.<sup>28</sup> Conversely, in nocodazole treated U2OS-CDC25B-DDA cells, Kiz electrophoretic mobility was not reduced (Fig. 5A), indicating that CDC25B stabilization affects Kiz phosphorylation. CDC25B stabilization could impair Kiz phosphorylation indirectly by affecting either the level and/or activity of Plk1 kinase, or Kiz centrosomal localization, which depends on Kiz interaction with Cep72.<sup>36</sup> To explore these possibilities, the expression level and activity of Plk1 kinase were assessed by immunoblotting. We found that neither Plk1 expression level nor its activation, visualized by its phosphorylation on Thr120 were affected in mitotic U2OS-CDC25B-DDA cells in comparison to mitotic U2OS-CDC25B-Wt cells, in which Kiz phosphorylation was not impaired (Fig. 5B). Also, immunofluorescence analysis showed that Kiz localized normally at centrosomes in mitotic U2OS-CDC25B-DDA cells (Fig. 5C).

These findings suggest a potential enzyme-substrate relationship between CDC25B and Kiz at centrosomes that could affect Plk1-mediated phosphorylation of Kiz. To confirm this possibility, we performed an in vitro kinase and phosphatase assay with recombinant GST-Kizuna (GST-Kiz), His<sub>6</sub>-Plk1 and MBP-CDC25B (Fig. 6A, left panel). As a specificity control, we used CDC25A, which is closely related to CDC25B and shares CDK/



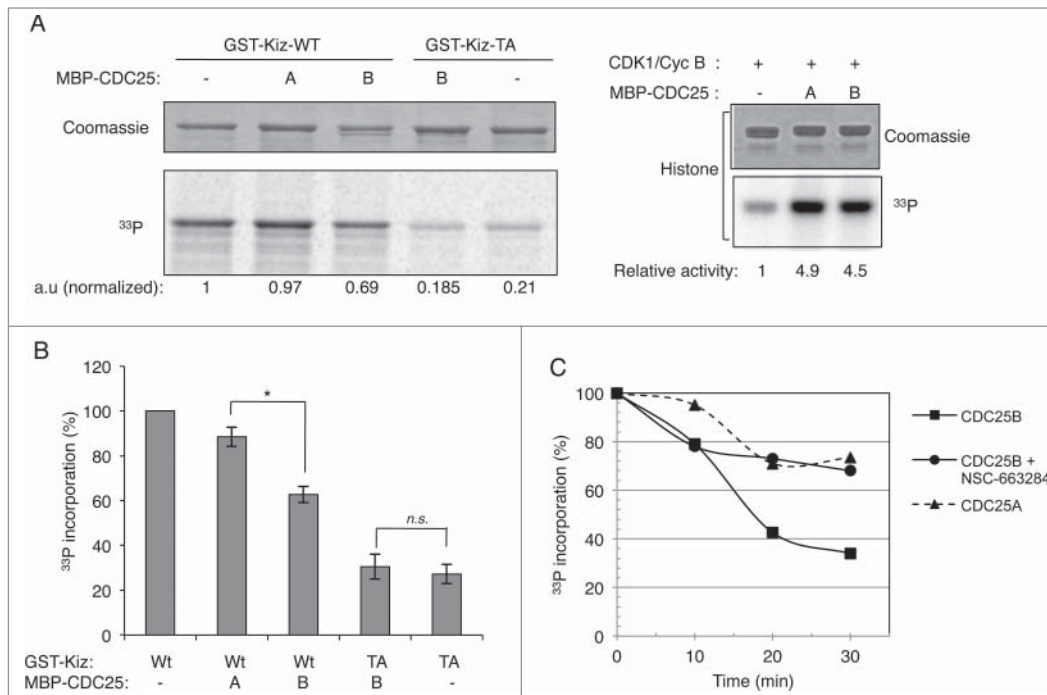
**Figure 5.** Stabilization of CDC25B during mitosis modifies Kiz phosphorylation without affecting Plk1 activity and centrosomal localization of Kiz. **(A).** Asynchronously growing (AS) and nocodazole treated (M) parental U2OS cells or that express CDC25B-Wt or CDC25B-DDA were lysed and immunoprobed as indicated. \*: non-specific band. **(B).** Experiments were performed as in A, except that cell extracts were immunoprobed with anti-Plk1 and anti-Phospho-Plk1 (Thr120). **(C).** Twenty-four hours after the induction of CDC25B-Wt or CDC25B-DDA expression in U2OS Tet-off cells, cells were fixed and incubated with anti-Centrin-2 and -Kiz antibodies. DNA was stained with DAPI. Images of mitotic cells are shown. Scale bar, 10  $\mu$ m.

Cyclin substrates with CDC25B. We first verified that recombinant CDC25A and B had a similar phosphatase activity, i.e., were able to activate to a similar extent the initially inhibited (phosphorylated) CDK1/Cyclin B complexes (Fig. 6A, right panel). When GST-Kiz, previously phosphorylated by Plk1, was incubated with MBP-CDC25B for 20 min,<sup>33</sup> P incorporation in Kiz was significantly reduced to 30 to 35 % of the initial phosphorylation level, showing that Plk1-phosphorylated Kiz can be a CDC25B substrate (Fig. 6A-B). Conversely, under the same conditions, MBP-CDC25A did not dephosphorylate Kiz as efficiently as CDC25B (3 to 11 % reduction). Analysis of a time-course dephosphorylation of Plk1-phosphorylated Kiz by both phosphatases showed that Kiz-dephosphorylation was more efficient with MBP-CDC25B than with MBP-CDC25A (Fig. 6C), and when MBP-CDC25B was inactivated by a specific CDC25-inhibitor (NSC-663284), the level of Kiz dephosphorylation was similar to that obtained with MBP-CDC25A alone (20 and 30 min) (Fig. 6C). The 35% remaining Kiz-phosphorylation observed in the case of MBP-CDC25B treatment could indicate that either residual phosphorylation corresponds to off-target phosphorylation (see below) or in vitro Plk1 phosphorylates also Kiz, on sites not targeted by CDC25B but maybe by other phosphatases, including CDC25C. Surprisingly, we never obtained a total inhibition of Kiz dephosphorylation in presence of CDC25 inhibitor possibly due to the presence of contaminating phosphatases. As the 3 CDC25 phosphatases bear a catalytic domain

Kiz dephosphorylation by this phosphatase. To support these in vitro experiments, we expressed a phosphomimetic mutant of Kiz (Kiz-T379E) to counteract the appearance of multipolar spindles in U2OS-CDC25B-DDA cells. Unfortunately, all our attempts resulted in cell death (Fig. S2, upper panel) or the total absence of mitotic cells (Fig. S2, lower panel), depending of expression vector used.

To determine if CDC25B could directly regulate the phosphorylation level of Kiz, we investigated whether Kiz-CDC25B complexes exist in cells. Immunoprecipitation experiments (Fig. 7A) showed a weak interaction between co-expressed HA-CDC25B and Myc-Kiz in asynchronously growing HeLa cells. The weak signal probably reflects a transient enzyme-substrate interaction that occurs in a precise cell compartment and cell cycle phase. Indeed, while endogenous Kiz is localized at the centrosome throughout the cell cycle,<sup>28</sup> only a subset of endogenous CDC25B is located there.<sup>10,37</sup> We next assessed a potential differential interaction between endogenous Kiz and CDC25B in mitosis relative to interphase. CDC25B and Kiz were immunoprecipitated from asynchronous or mitotic HeLa cells, and the immunoprecipitates analyzed by immunoblotting. While no interaction could be detected between CDC25B and Kiz in extracts from asynchronous cells (Fig. 7B, left panel), hypophosphorylated forms of Kiz were co-immunoprecipitated with CDC25B from mitotic cell extracts (Fig. 7B, middle panel). These

highly conserved but greatly differ in their N-terminal regulatory domain, our results strongly support the idea that CDC25B effect on Kiz is highly specific and is not the result of a promiscuous in vitro interaction. Since Kiz Thr379 is the only phosphorylation site for Plk1 in cellulo,<sup>28</sup> we also used in the same assay the Kiz-T379A mutant (GST-Kiz-TA) as a control of in vitro off-target phosphorylation. As with GST-Kiz-Wt, CDC25B did not modify the residual phosphorylation level of GST-Kiz-TA (Fig. 6A-B). Altogether, these results demonstrate that CDC25B, but not CDC25A, can specifically reverse Plk1-dependent Kiz phosphorylation. Thus, occurrence of multipolar spindles caused by PCM fragmentation in cells where CDC25B is stabilized could be the consequence of



**Figure 6.** Kiz is dephosphorylated by CDC25B, not CDC25A. **(A).** Recombinant GST-Kiz-Wt or GST-Kiz-TA (T379A) immobilized on Glutathione beads was used in a phosphorylation/dephosphorylation assay in the presence of recombinant His<sub>6</sub>-Plk1 and MBP-CDC25A or MBP-CDC25B, as described in "Materials and methods." The phosphorylation level of GST-Kiz was then determined by autoradiography. Incorporation of <sup>33</sup>P into GST-Kiz, was normalized to the protein amount of each band based on Coomassie staining (left panel). The relative activity of recombinant MBP-CDC25 phosphatases was indirectly determined by measuring the activation of inactive CDK1 associated with Cyclin B (i.e., CDK1 phosphorylation on Thr14 and Tyr15), in a kinase assay using histone H1 as substrate. For this 50 ng of recombinant MBP-CDC25A or MBP-CDC25-B were incubated with immunoprecipitated inactive CDK1/Cyclin B complexes. Activity of CDK1/Cyclin B complexes was then tested in an in vitro histone H1 kinase assay, as described in.<sup>51</sup> Equal loading was shown by Coomassie staining. The incorporation of <sup>33</sup>P into histone H1 reflects the activity of CDK1/Cyclin B complexes (right panel). **(B).** Quantification of <sup>33</sup>P incorporation in recombinant GST-Kiz obtained in A. Values from 3 independent experiments were normalized to those of GST-Kiz-Wt incubated only with His<sub>6</sub>-Plk1. Data are the mean ± SEM of 3 independent experiments (\*, P = 0.0318). **(C).** Quantification of <sup>33</sup>P incorporation in recombinant GST-Kiz-Wt during a kinetic of dephosphorylation by recombinant MBP-CDC25B, in presence or not of NSC-663284 (500 nM), or MBP-CDC25A. Values were normalized to those of GST-Kiz-Wt incubated only with His<sub>6</sub>-Plk1.

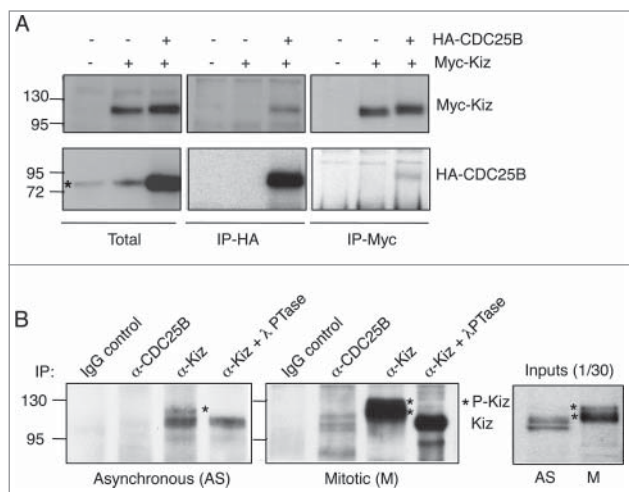
forms co-migrate with the hypo-phosphorylated Kiz bands following  $\lambda$  phosphatase treatment of the immunoprecipitates obtained with anti-Kiz antibodies. These results demonstrate that CDC25B can interact with Kiz and dephosphorylates it in mitosis.

To dissect the mechanism governing Kiz regulation by CDC25B during mitosis, we investigated in cellulo the precise timing of Kiz phosphorylation and the effects of Plk1 and CDC25B inhibitors. Immunoblotting analyses of extract from asynchronous, G<sub>2</sub>/M transition block or pro-metaphase arrested cells indicated that Kiz was neither phosphorylated in interphase nor at the G<sub>2</sub>/M transition, but fully phosphorylated in pro-metaphase resulting in a mobility shift in SDS-PAGE (Fig. 8A). We then examined Kiz mobility in mitotic cells during a time-course after release from G<sub>2</sub>/M block. Probing for Kiz revealed that the change in electrophoretic mobility of Kiz occurred already 12 min after G<sub>2</sub>/M-block release (when cells enter mitosis), but no reduction of the shift was observed in the presence of Plk1 inhibitor (Fig. S3). These results suggest that: i) Kiz is

phosphorylated as soon as cells enter mitosis and ii) under our condition the mitosis duration (40-50 min) is too short to see in early phases of mitosis the effect of Plk1 inhibitor or alternatively Kiz is phosphorylated by other kinases in addition to Plk1. Entrance into mitosis is induced by the activation of CDK1/Cyclin B complexes, which phosphorylate many substrates, phosphorylation that can prime for Plk1 phosphorylation in some cases. Performing an in vitro kinase assay with CDK1/Cyclin B, we could show that Kiz is a substrate for CDK1/Cyclin B in vitro (Fig. 8B). Next we used a pro-metaphase cell extract in an in vitro kinase assay with recombinant GST-Kiz-Wt in presence or not of Plk1,<sup>38</sup> CDK1 and CDC25<sup>39</sup> inhibitors (Fig. 8C). As shown in Figure 8C, the pro-metaphase cell extract was able to phosphorylate GST-Kiz-Wt in vitro, and the pre-incu-

bation of the same extract with either Plk1 or CDK1 inhibitors decreased GST-Kiz phosphorylation of 30% and 40% after 30 min, respectively. Moreover, a pre-incubation of the extract with both inhibitors had an additive inhibition of Gst-Kiz phosphorylation. These results are in agreement with the hypothesis that CDK1 is involved in the priming phosphorylation of Kiz. Noteworthy, phosphorylation of GST-Kiz was inhibited by 50% when CDC25 inhibitor was added to the pro-metaphasic cell extract. Knowing that CDK1 contributes to the phosphorylation of Kiz and CDC25B positively controls CDK1 kinase activity (Fig. 8D) our result appears consistent. Altogether these results support the involvement of CDC25B in the complex regulation of Kiz activity in PCM stabilization through the control of its phosphorylation.

In summary, our experiments show that: i) in mitosis, expression of a CDC25B mutant (CDC25B-DDA) whose proteasomal degradation is impaired<sup>32</sup> leads to Kiz dephosphorylation and PCM fragmentation, ii) in vitro, CDC25B, but not its close relative CDC25A, can reverse Plk1-dependent phosphorylation of



**Figure 7.** CDC25B interacts with Kiz during mitosis. **(A).** HeLa cells were transfected or not with HA-CDC25B-Wt or Myc-Kiz (pRK5) and lysed 24 h after transfection. Cell extracts were incubated with control IgG or anti-Myc/anti-HA antibodies, and immunocomplexes probed for the indicated proteins. \*: non-specific band. **(B).** Asynchronous (left panel) or mitotic (right panel) HeLa cells were lysed in the presence of a cross-linking agent (DSP), immunoprecipitated with anti-CDC25B or anti-Kiz antibodies and immunoprobed with anti-Kiz antibodies. In indicated cases, immunoprecipitates obtained with anti-Kiz antibodies were treated with lambda phosphatase ( $\lambda$ PTase, 200 units) 15 min at 30°C before SDS-PAGE ( $\alpha$ -Kiz +  $\lambda$ PTase).

Kiz, a modification that is critical for PCM stability,<sup>28</sup> iii) in cellulo, CDC25B interacts with Kiz during mitosis, when its phosphorylation is required for PCM stability. Based on these results, we propose a model in which, during mitosis (from prophase to the end of metaphase), the phosphorylation level of Kiz at centrosomes is controlled by a balance between Plk1 kinase and CDC25B phosphatase activities (Fig. 9). When cells enter mitosis, unphosphorylated Kiz would be first phosphorylated by CDK1/Cyclin B (on site(s) that remain to be characterized), which would prime it for Plk1 phosphorylation. Until metaphase, a precise balance between Plk1 kinase and CDC25B (and potentially CDC25C) phosphatase activities could allow at the same time expansion and stabilization of the PCM necessary for a proper establishment of the mitotic spindle. Under normal conditions, the abrupt, proteasome-dependent degradation of CDC25B at the metaphase-anaphase transition, tilts the balance in favor of Plk1. Thus ensuring the complete phosphorylation of Kiz that is required for its function to stabilize and maintain a robust PCM architecture (Fig. 9A). When CDC25B is stabilized, i.e., U2OS-CDC25B-DDA cells, this results in its high expression level throughout mitosis, which exceeds the CDC25B threshold required to maintain a proper balance between Kiz-phosphorylation and -dephosphorylation. As a consequence, Kiz is maintained under an hypo-phosphorylated status, and PCM is structurally weakened during all mitotic phases (Fig. 9B).

Furthermore, these data identify Kiz as a new substrate of the CDC25B phosphatase other than CDK1/Cyclin B during mitosis. Thus, CDC25B participates not only in the control of

centrosome duplication<sup>12</sup> through the regulation of CDK/Cyclin complexes and centrin 2 stability,<sup>13</sup> but also in the control of the stability of the PCM around centrosomes during mitosis by dephosphorylating Kiz. This finding allows us to extend the list of substrates for the members of CDC25 phosphatase family. Some of them require the catalytic activity of CDC25, such as the transcription factor Cut,<sup>40</sup> Raf kinase<sup>41</sup> that in turn controls CDC25A activity,<sup>42</sup> the epidermal growth factor receptor (EGFR),<sup>43</sup> or the extracellular signal-regulated kinase (ERK)<sup>44</sup> for CDC25A. Others, such as steroid receptors for CDC25B, are independent of the phosphatase activity,<sup>45</sup> and CDC25B is considered as a co-activator. Our result suggests that, during the cell cycle, a defect in the degradation of CDC25 phosphatases will affect differently their various substrates, ultimately leading to genomic instability. It will be interesting to determine whether such defects exist in cancers cells overexpressing CDC25B, as it was shown for CDC25A,<sup>46</sup> and to identify other new CDC25 substrates that might participate in chromosomal instability during carcinogenesis.

## Materials and Methods

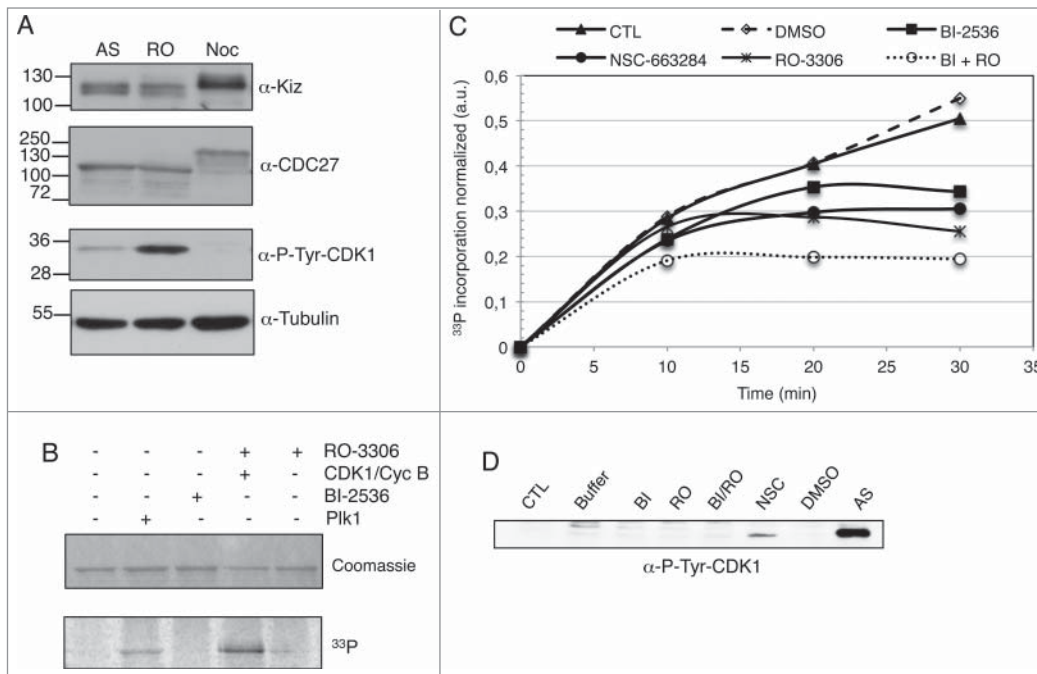
### Plasmids

Human Kizuna cDNA was PCR-amplified from pDONR223 (provided by the Montpellier Genomic Collections facility, Institut de Génétique Moléculaire de Montpellier, Montpellier, France) and cloned into pRK5-Myc (provided by G. Bompard, Institut de Génétique Humaine, Montpellier, France), and pGEX-2T (GE Healthcare). PCR-based site-directed mutagenesis (Thr379-Ala) was performed using the pGEX-Kiz plasmid as a template. pcDNA-6Myc-Kizr (Wt, T379E and T379A) were provided by M. Ohsugi and T. Yamamoto (Institute of Medical Science, university of Tokyo, Tokyo, Japan). CDC25B1-Wt and CDC25B1-DDA subcloned into pcDNA3-HA and pMAL-C2 were described elsewhere.<sup>47,48</sup> Both CDC25B-Wt and -DDA open reading frames were cloned into pmCherry-C1. Point mutation in CDC25B catalytic domain (Cyst473Ser) was generated by PCR mutagenesis on the pmCherry-CDC25B.

### Antibodies and compounds

The following antibodies were used for immunoprecipitation or immunoblotting: rabbit polyclonal anti-CDC25B (sc-326, Santa Cruz Biotechnology, Inc.), anti-Kizuna (A300-946A, Bethyl Laboratories, Inc.), anti-Phospho-Plk1 (Thr210) (# 5472, Cell Signaling Technology), anti-P-Tyr-CDK1 (# 9111, Cell Signaling Technology), anti-CDC27 (a gift of T. Lorca (Center de Recherche de Biochimie Macromoléculaire, Montpellier, France), monoclonal anti- $\alpha$ -Tubulin (clone DM1A, Sigma-Aldrich), anti-Plk1 (F8, sc-17783, TEBU), anti-HA (clone 12CA5, Roche Diagnostic), anti-Myc (clone 9E10, Abcam), and GFP-Trap agarose beads (ChromoTek). Secondary HRP-conjugated antibodies were purchased from Bio-Rad SA.

The following antibodies were used for indirect immunofluorescence: rabbit polyclonal anti- $\gamma$ -Tubulin and anti-Centrin-2 were a gift of B. Raynaud-Messina (Center de Biologie du



**Figure 8.** CDK1/Cyclin B, Plk1 and CDC25B regulate Kiz-phosphorylation level during mitosis. **(A).** Total cell extract (30  $\mu$ g) of asynchronous (AS), RO-3306 (RO) or nocodazole treated (Noc) HeLa cells were analyzed on SDS-PAGE and immunoprobed as indicated. Anti-CDC27 and anti-P-Tyr-CDK1 were used to check mitosis progression. Anti-Tubulin served as a loading control. **(B).** Recombinant GST-Kiz-Wt immobilized on Glutathione beads was used in an in vitro kinase assay in the presence of recombinant His<sub>6</sub>-Plk1 or GST-CDK1/GST-Cyclin B kinases, pre-incubated or not with BI-2536 (250 nM) and RO-3306 (1  $\mu$ M), respectively. The phosphorylation level of GST-Kiz was then determined by autoradiography. **(C).** Recombinant GST-Kiz-Wt immobilized on Glutathione beads was used in a phosphorylation assay in the presence of pro-metaphase HeLa cell extract, as described in "Materials and methods," in presence or not of BI-2536 (250 nM), RO-3306 (1  $\mu$ M), NSC-663284 (1  $\mu$ M) or DMSO. The phosphorylation level of GST-Kiz was then determined by autoradiography. Incorporation of <sup>33</sup>P into GST-Kiz, was normalized to the protein amount of each band based on Coomassie staining. Values from 3 independent experiments were normalized to those of GST-Kiz-Wt incubated only with pro-metaphase extract. Data are the mean of 3 independent experiments. **(D).** Pro-metaphase HeLa cell extract was incubated for 30 min at 30°C with kinase buffer in presence of the indicated kinases or phosphatase inhibitors, as in C, analyzed on SDS-PAGE and immunoprobed with anti-P-Tyr-CDK1. Extract from asynchronous HeLa cells was used as control for CDK1 phosphorylation on Tyr-15.

Développement, Toulouse, France) and J. Piette (Center de Recherche de Biochimie Macromoléculaire, Montpellier, France), respectively. Secondary antibodies were Alexa Fluor 488 or 594 conjugates (Interchim).

BI-2536 (Plk1 inhibitor, S1109, Selleckchem), RO-3306 (CDK1 inhibitor, SML0569, Sigma-Aldrich), NSC-663284 (CDC25 inhibitor, N7537, Sigma-Aldrich) and MG132 (Calbiochem) were dissolved in DMSO and used at the indicated concentrations.

#### Cell culture and transfections

HeLa cells, U2OS cells and U2OS Tet-off cells were grown in DMEM (Lonza) containing 4.5 g/liter glucose, 10% heat inactivated fetal bovine serum (Biowest), 2mM glutamine, 100 U/ml penicillin and 10  $\mu$ g/ml streptomycin (Lonza). The U2OS Tet-off cell lines that stably express HA-CDC25B1-Wt or HA-CDC25B1-DDA were previously described;<sup>47,32</sup> culture medium for U2OS Tet-off cell was supplemented with 250  $\mu$ g/ml G418 (Sigma-Aldrich), 200  $\mu$ g/ml hygromycin B (Calbiochem) and 2  $\mu$ g/ml tetracycline (Sigma-Aldrich). For induction

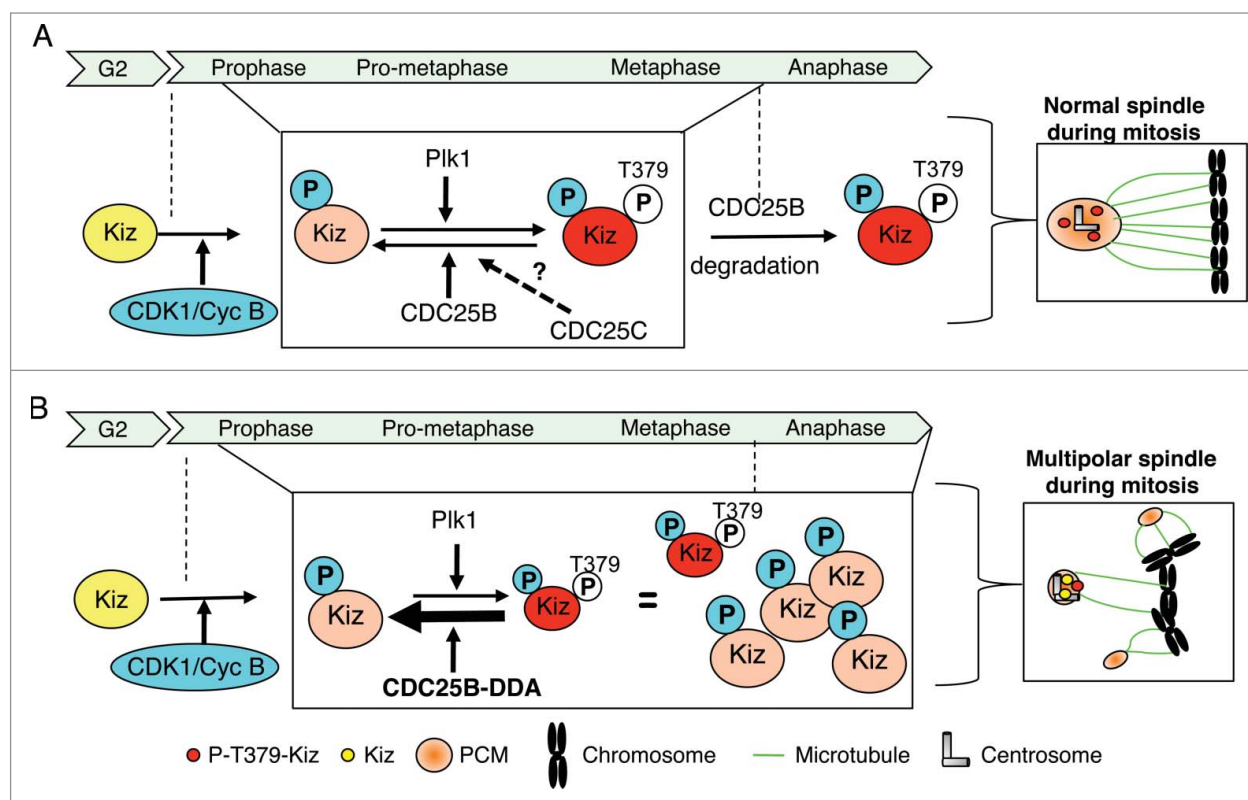
of HA-CDC25B1 expression, tetracycline was removed from the medium and cells were analyzed 24 hours after induction. HeLa and U2OS cells were arrested in pro-metaphase by adding 200 ng/ml or 40 ng/ml nocodazole (Sigma-Aldrich), respectively, for 14 h. Mitotic cells were recovered by shake-off. To obtain U2OS cells arrested at G2/M transition, cells were first blocked at G1/S transition by a treatment with 2.5 mM Thymidine (Sigma-Aldrich) for 24 h, then released for 6 h before adding 9  $\mu$ M RO-3306 (Sigma-Aldrich) for 11 h. To induce entrance and progression into mitosis, cells were extensively washed with fresh media. Transient transfections with expression vectors (Myc-Kiz, HA-CDC25B or mCherry-CDC25B) in asynchronous HeLa or U2OS cells were performed using JetPEI™ (Ozyme), according to the manufacturer's instructions.

#### Indirect immunofluorescence and time-lapse

#### analysis

Cells grown on coverslips were either fixed with 3.7% paraformaldehyde and then permeabilized with 0.25% Triton X100 and cold methanol, or only fixed/permeabilized with cold methanol at -20°C. Cells were washed with PBS and blocked with 1% FCS/PBS for 15 min. Incubation with primary antibodies was carried out at 37°C for 1 h and with secondary antibodies at room temperature for 40 min. DNA was stained with DAPI (D9542, Sigma-Aldrich). Microscopic examinations were performed with 63X/1.4 NA or 100X/1.4 NA oil immersion objective lenses using a DM 6000 microscope (Leica). Microphotographs were taken with a 12-bit Coolsnap HQ2 1 camera. Images were acquired as TIF files using the MetaMorph imaging software (Molecular Devices).

HeLa cells, transfected with various pmCherry expression vectors, were imaged 14 hours after transfection, using a DMIRE2 microscope (Leica) with a 40X N PLAN L 0.55 LMC objective. Images were collected every 10 minutes using a Micromax YHS 1300 camera (Roper Scientific Princeton Instruments) monitored by MetaMorph software.



**Figure 9.** Proposed model for the regulation of Kiz phosphorylation during mitosis. **(A).** During mitosis, phosphorylation level of Kiz results from CDK1/Cyc B kinase activity and a balance between Plk1 kinase and CDC25B phosphatase activities. At the entrance into mitosis Kiz is phosphorylated by CDK1/Cyc B, a priming phosphorylation that facilitates its phosphorylation on Thr379 by Plk1. Until metaphase-anaphase transition, phosphorylation state of Thr379 results from a balance between activities of Plk1 and CDC25B (and maybe CDC25C) to finely control the equilibrium between expansion and stabilization of the PCM around the centrosomes. At the metaphase-anaphase transition, when CDC25B is degraded by the proteasome, Kiz is fully phosphorylated on Thr379 by Plk1. **(B).** During mitosis, if a defect in proteasome-dependent degradation of CDC25B occurs, such as when the CDC25B-DDA mutant is expressed, the high level of CDC25B maintains Kiz in an hypo-phosphorylated form that hinders its ability to stabilize PCM around the centrosomes during all phases of mitosis, resulting into the formation of multipolar spindles.

#### Expression and purification of recombinant proteins

Recombinant MBP-CDC25A and MBP-CDC25B1 were expressed in the *E. coli* JM109 strain in the presence of 0.2% glucose, induced with 100  $\mu$ M isopropyl  $\beta$ -D-thiogalactoside (IPTG) (EU0008, Euromedex) at 25°C for 3 hours, and affinity purified with amylose beads (New England Biolabs, Inc.) as described previously.<sup>47</sup> Recombinant GST-Kizuna-Wt (GST-Kiz-Wt) and GST-Kizuna-T379A (GST-Kiz-TA) were expressed in the *E. coli* TOP10 strain and induced with 1 mM IPTG at 37°C for 3 hours. After lysis in a buffer containing 20 mM Tris-HCl, pH 7.5, 150 mM NaCl, 5 mM EDTA, 0.5% NP40 and 1X complete protease inhibitor cocktail (11873580001, Roche Diagnostic), recombinant proteins were purified on Glutathione-Sepharose beads (17-0756-01, GE Healthcare). Recombinant His<sub>6</sub>-Plk1, a gift of Dr. S. Rouquier (Institut de Génétique Humaine, Montpellier, France), was purified under native conditions as described previously.<sup>49</sup> Recombinant CDK1-Cyclin B (# 7518) was purchased from Cell Signaling Technology.

#### Co-immunoprecipitation

For co-immunoprecipitation of ectopically expressed proteins, HeLa cells were co-transfected with the corresponding constructs. Cells were homogenized in lysis buffer (50 mM Tris-HCl, pH 7.5, 250 mM NaCl, 0.1 % Triton X100, 1 mM DTT, 5 mM EDTA, 50 mM NaF and 0.1 mM Na<sub>3</sub>VO<sub>4</sub>) in the presence of 1X complete protease inhibitor cocktail and 25  $\mu$ M MG132 (BML-PI102-0005, ENZO life) at 4°C for 30 min. After centrifugation at 13,200 rpm for 10 min, 2  $\mu$ g anti-HA or anti-Myc antibodies were added to the pre-cleared supernatants. Proteins were immunoprecipitated at 4°C for 2 hours and collected by addition of 10  $\mu$ l protein-A or -G Sepharose. After extensive washes, beads were boiled in 2X loading sample buffer and analyzed by SDS-PAGE and immunoblotting.

For co-immunoprecipitation of endogenous proteins, cell extracts were obtained as described in MacKay et al.<sup>50</sup> Briefly, HeLa cells were homogenized in lysis buffer (40 mM Hepes-KOH, pH 7.5, 120 mM NaCl, 1% Triton X100, 1 mM EDTA), complete protease inhibitor cocktail and 2 mg/ml of Dithiobis [succinimidyl propionate] (DSP) (22585, Pierce) at

4°C for 30 min. DSP was neutralized by addition of 1M Tris-HCl, pH 7.5, to a final concentration of 200 mM, 25 µM MG132 was then added and the mix incubated at 4°C for another 30 min. After centrifugation at 13,200 rpm for 10 min, 3 µg anti-CDC25B or anti-Kiz antibodies were added to the pre-cleared supernatants and proteins were immunoprecipitated as above. As control, random rabbit IgGs were used.

### Kinase and phosphatase assays

1 µg of GST-Kiz bound to Glutathione-Sepharose beads was incubated in kinase buffer (20 mM Tris-HCl, pH 7.5, 10 mM MgCl<sub>2</sub>, 1 mM EGTA, 5 mM DTT, 5 mM NaF, 50 µM ATP) supplemented with 5 µCi of γ-<sup>33</sup>P-ATP (Perkin Elmer) and 150 ng of recombinant His<sub>6</sub>-Plk1 were added to the mix that was incubated at 30°C for 15 min. After 2 washes with TNE buffer (20 mM Tris-HCl, pH 7.5, 125 mM NaCl, 2 mM EGTA, 0.5 % NP40) and one wash with phosphatase buffer (50 mM Tris-HCl, pH 8, 50 mM NaCl, 1 mM DTT, 1 mM EDTA), GST-Kiz beads were incubated with 72 ng MBP-CDC25A or 78 ng of MBP-CDC25B1 at 30°C for 20 min, washed twice with phosphatase buffer and boiled in 2X loading sample buffer before analysis by SDS-PAGE.

For in vitro phosphorylation assay using cell extract, HeLa cells blocked in pro-metaphase were lysis in NET buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1 mM EDTA, 0.25% IGE-PAL, 1 mM EDTA) containing 10 mM PMSF, 25 µM MG132, complete protease inhibitor cocktail, during 20 min at 4°C. After centrifugation, concentration of clarified cell extract was determined. 1 µg of GST-Kiz bound to Glutathione-Sepharose beads was incubated in kinase buffer (50 mM Tris-HCl, pH 7.5, 5 mM MgCl<sub>2</sub>, 1 mM DTT, 5 µM MG132, 50 µM ATP) supplemented with 5 µCi γ-<sup>33</sup>P-ATP and 30 µg of cell extract was added to the mix that was incubated at 30°C

for indicated time. After 2 washes with TNE buffer, beads were boiled in 2X loading sample buffer before analysis by SDS-PAGE.

### Statistical analysis

Statistical analyses were performed using the non-parametric Mann-Whitney test and the Prism software (GraphPad Software). Statistical significance was defined as  $P \leq 0.05$ .

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

### Acknowledgments

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### Supplemental Material

Supplemental data for this article can be accessed on the publisher's website. <http://www.tandfonline.com/kccy>

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