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# Genome organization via loop extrusion, insights from polymer physics models

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## Abstract

Understanding how genomes fold and organize is one of the main challenges in modern biology. Recent high-throughput techniques like Hi-C, in combination with cutting-edge polymer physics models, [have provided access to](#) precise information on 3D chromosome folding to decipher the mechanisms driving such [multi-scale](#) organization. In particular, Structural Maintenance of Chromosomes (SMC) proteins play an important role in the local structuration of chromatin, putatively via a loop extrusion process. Here, we review the different polymer physics models that investigate the role of SMCs in the formation of topologically-associated domains (TADs) during interphase via the formation of dynamic loops. We describe the main physical ingredients, compare them and discuss their relevance against experimental observations.

## Keywords

Chromosome organization; modeling; polymer; loop extrusion; SMC proteins

## Summary key points

- SMC-mediated chromatin loops may be formed via a loop extrusion process.
- Polymer physics models have been instrumental in studying loop extrusion.
- Loop extrusion may be driven by: directed translocation of active motors, diffusion of slip-links, transcription-induced supercoiling.
- We describe the main physical ingredients and parameters to better understand and compare these three classes of models.

## 1. Introduction

Inside cellular nuclei, genomic DNA is tightly packed into a polymer-like structure called chromatin whose local conformation modulates the accessibility and specificity of regulators to the DNA sequence [1]. Until recently, chromatin was mainly studied as a unidimensional object and little attention was paid to its three-dimensional structure. In the last decade, thanks to the development of chromosome conformation capture (Hi-C) and advanced microscopy techniques, major progress has been realized in our understanding of

chromosome organization during interphase [2,3,4]. The genome is locally partitioned into conserved kbp- to Mbp-size domains with a relatively high number of contacts, known as topologically-associating domains (TADs) that usually encompass promoter-enhancer looping interactions. At larger-scale (1-10 Mbp), loci with similar gene activity or epigenomic content self-organize into spatial compartments [5–8]. Finally, at the nuclear level, chromosomes occupy separate 3D territories that only weakly intermingle [9, 10] and functional chromatin compartments preferentially localize at the periphery for heterochromatic regions or centrally for euchromatic segments [11, 12]. This peculiar multi-scale organization is observed in most of higher eukaryotes, from drosophila to worm and mammals [13], suggesting that 3D organization plays a fundamental role in the regulation of gene expression. Indeed, many studies have illustrated the role of TAD and TAD boundaries in promoting or preventing enhancer-promoter interactions for important genes during development or diseases [14–20], even if several recent works tend to question a global, direct effect of the local 3D organization on transcription [7, 21–23].

While the description of 3D chromatin folding is becoming more and more quantitative, our understanding of the molecular mechanisms that drive the 3D genome organization remains sparse. Several processes have been shown or suggested to play major roles in organizing chromosomes [24]. Formation of epigenomic or other nuclear, membrane-less compartments is putatively mediated by chromatin-binding proteins that have the capacity to self-interact [25–27] and potentially phase-separate with the formation of protein micro-droplets where specific chromatin segments may colocalize [28–30]. TAD formation in mammals has been associated with the translocation along the genome of SMC complexes, like condensin or cohesin rings, and with their association with insulator proteins bound at TAD boundaries, like CTCF [31–34]. In this review, we will focus on this latter family of mechanisms, which are often referred to as loop extrusion processes in the literature.

Application of the loop extrusion process to explain TAD formation originated from the combination of recent high-resolution Hi-C experiments on interphasic mammalian nuclei with older concepts originally developed for the condensation of mitotic chromosomes. In recent years, improvements in Hi-C resolution allowed to get a more precise description of TAD organization and substructures. In particular, in mammals, several groups observed that many TADs possess an enrichment of contact between the two TAD boundaries leading to corner-peaks in Hi-C maps [7] (Fig.1A) or between a boundary and the interior of the TAD leading to prominent stripes [49] (Fig.1B). Careful analyses of these peak signals showed that contacts occur mainly between convergent CTCF-binding sites usually located at domain boundaries [7, 32, 35, 36] and are associated with the presence of cohesin SMC complexes (Fig.1A, B). The preferential sense-antisense orientations between contacting loci cannot be simply explained by specific, 3D interactions [32] (Fig.2A) but are instead consistent with a 1D mechanism of loop formation along the genome (Fig.2B,C). Actually, this type of process was firstly hypothesized by Riggs [110] and Nasmyth [38] to explain the formation of 1D arrays of contiguous loops in mitotic chromosomes (Fig.2B) by condensin SMC complexes, and was clearly formalized by Alipour and Marko [39] in 2012 and coined *loop extrusion*. In 2015, based on these ideas, Lieberman-Aiden's and Mirny's groups concurrently suggested that TAD formation followed the same basic principle: loop extruding factors, likely to be cohesin rings in mammals, are loaded onto chromatin (by Nipbl proteins), sequentially extrude large chromatin loops, and eventually dissociate from DNA (thanks to Wapl proteins) or are halted as they encounter boundary elements (CTCF-bound motifs) with specific orientation (Fig.2C). While the molecular details of the loop extrusion mechanism remain unclear, many direct in

*vitro* observations have confirmed that the ATP-driven activity of SMCs may induce DNA [37,67,111] or chromatin [100] compaction, and that condensin exhibits a loop extrusion-like activity on naked DNA [40,41]. Moreover, *in vivo*, loop extrusion is supported by many indirect experiments in which, for example, the levels of cohesin (or of Nipl or Wapl) and of CTCF were manipulated, or where the positions or orientations of CTCF motifs were modified [31, 42–49] (see [34] for a detailed review on these evidences).

Beyond TAD formation in mammals or mitotic condensation in eukaryotes, loop extrusion mechanism by SMCs has been involved in the regulation of many fundamental biological processes like the meiotic chromosome assembly in yeast [112] or the juxtaposition of chromosome arms in bacteria [106,113]. It is therefore of crucial importance to better characterize this process. In recent years, polymer models have been instrumental in showing that the loop extrusion mechanism was consistent with experimental data. In this article, we review, compare and discuss the different polymer models that have been developed to describe and explain the loop extrusion process. In particular, we aim at describing the physical ingredients, the main parameters, and their typical values for a non-expert reader to better understand the physical grounds of these models and their relative differences.

## 2. Polymer physics models of loop extrusion

One of the great advantages of mechanistic or bottom-up modeling in biology is the possibility to test, based on first principles, if putative mechanisms, processes or hypotheses are compatible with experimental observations, while direct validation might be experimentally difficult due, for example, to technical limitations or because the involved actors have ubiquitous functions. Once validated, these models can then be used to predict novel behaviors or to drive new experiments. Along this line, polymer-physics-based models have been instrumental in 3D genomics to better understand the structures and functions of chromosomes [24, 51] (see also the recent book edited by Giorgetti and Tiana [52] illustrating several existing approaches). In particular, in the context of TAD formation by loop extrusion process, polymer models have played a crucial role to quantitatively relate experiments to mechanisms involved in this complex process [31, 32]. Previously, generic, large-scale features like the formation of chromosome territories or the decay of the average contact frequency  $P(s)$  between two loci as a function of their genomic distance  $s$  ( $P(s) \propto s^{-1}$  [5], Fig.1D) have been well captured by simple topologically-constrained self-avoiding polymers evolving in confined environment, the so-called crumpled or fractal polymer models [51, 53, 54]. Decoration of these models with specific interactions between monomers allowed to describe compartmentalization of chromatin into epigenomic compartments and TADs in higher eukaryotes [55–66]. In particular, the formation of mammalian TADs (with or without corner-peaks, Fig.1 A, C) might be captured by the partial collapse, the so-called theta- or coil-globule transition, of 1D chromatin domains eventually facilitated by point-like interactions between boundaries to model corner-peaks [31, 60]. In these models, corner-peaks result from specific short-range interactions between CTCF-bound sites. However, such a molecular mechanism cannot physically explain why corner-peaks are observed only between sites with particular sense-antisense orientations [7, 32, 35] (Fig.2A). Models of loop extrusion recapitulate this intriguing observation along with the quantitative description of the average intra-TAD organization (Fig.1C) [31, 32]. In the following, we will review the three different types of existing polymer models for loop extrusion, depending on the main physical ingredient invoked to describe the translocation of extruders along chromatin.

## 2.1 Loop extrusion via translocating active motors

Based on in vitro single-cell observations that condensin SMCs are able to processively compact DNA via an unknown ATP-dependent mechanism [37, 67], Alipour and Marko in 2012 [39] were the first to propose a theoretical framework to model the loop extrusion process by SMCs (Fig.2B). They modeled chromosome as a 1D lattice, neglecting the 3D polymeric nature of chromatin. A fixed number of loop extruding factors (LEFs) are initially loaded on the lattice and act as molecular motors by stochastically modifying the linear distance between the two LEF 'legs' leading to growing loops. LEFs can eventually unbind and re-attach at a different location, but cannot overlap or cross each other. Within this framework, Alipour and Marko showed that depending on the concentration of LEFs on the lattice and on the LEF processivity (ie the average genomic distance extruded by an isolated LEF during its lifetime on chromatin), two types of organization are observed: highly ordered stacked configurations for lower processivities and concentrations, and disordered configurations with smaller loops for larger processivities and concentrations. They suggested that mitotic condensation by condensin might result from the former regime with arrays of consecutive stacked loops stabilized by boundary elements (BEs) that halt the progression or favor the dissociation of LEFs.

To quantitatively interpret the current Hi-C data of mammalian genome during interphase [7], Mirny's and Lieberman-Aiden's groups [31, 32], in 2015, developed physics-based models integrating a processive, unidirectional, loop extrusion process inspired by the seminal work of Alipour and Marko. Both approaches modeled chromosomes as 3D bead-and-string polymeric chains (600-1000 bp per monomer) which dynamically evolved under the action of LEFs (cohesin in interphase), modeled as rigid springs. LEFs bind at random position on chromatin, their two legs (i.e. the two edges of the spring) walking processively on chromatin in opposite directions, progressively extruding longer loops until they dissociate or reach a BE (CTCF sites with the correct orientation) where they stop (Fig.2C). Both models interpret phenomenologically the intra-TADs specificities (Fig.1) as the outcomes of this active translocation process: the average partial intra-TAD compaction and the corresponding scaling laws compared to inter-TAD properties (Fig.1C, D) result from the formation of dynamic loops by LEF only between regions belonging to the same TAD. Corner-peaks (Fig.1A) emerge when the processivity of LEFs is large enough for LEFs to reach both BEs at TAD borders before dissociation. The observation that corner-peaks represent contact mainly between convergent CTCF sites arises from the hypothesis that one LEF leg halts at BE only if the direction of translocation of the leg is opposite to the motif orientation, otherwise the leg may pass through the BE and continue its progression. Stripes (Fig.1B) may arise from preferential LEF loading closed to one BE [49]. In addition to BE positions and orientations, concentration and processivity of LEFs, as in Alipour and Marko's work, are the main physical ingredients controlling TAD folding (Fig.2D) with typical concentrations going from 1 LEF every 120 kbp [32] to 1 LEF every ~ 250 kbp [31] and processivities between 120 and 240 kbp in [32] or with higher values (>500 kbp) in [31]. Both models have shown remarkable (semi-) quantitative agreement with further perturbation experiments modifying BE locations or the levels of key components of the loop extrusion process [31, 34]. For example, in the absence of BEs, models predict the disappearance of corner peaks and corresponding TADs but the maintenance of the scaling laws and even their extension beyond the former TAD boundaries, in agreement with CTCF knock-down experiments [43].

It is, however, important to note that these two models have significant differences, making it difficult to directly compare their parameters and predictions. Indeed, both use different

null models for chromatin: Fudenberg et al [32] consider a semi-flexible chain with soft-core potential (weak excluded volume and possibility for polymer strands to cross each other) confined in a box with periodic boundary conditions, hence controlling the volumic density; Sanborn et al [31] adopt an isolated, self-avoiding, flexible chain with short-range, non-specific interactions between all monomers leading to a global collapse of the chain to mimic confinement. Another fundamental difference is the treatment of colliding LEFs and of LEF unbinding: in [32], LEFs follow a simple exclusion rule and may unbind stochastically from chromatin at any time as in [39]; in [31], one of the two interfering LEFs dissociates randomly unless one LEF is already stalled at a BE and thus remains bound, otherwise random dissociation is very rare. Knowing that these two null models may have very different structural behaviors for  $P(s)$  in the absence of LEFs [51, 54, 68], one can easily understand that the optimal parameters for LEF concentrations and processivities are quite different between the two models, in particular, if they employ distinct collision and dissociation rules. For example, these differences may explain why Fudenberg et al correctly predicts average inter-TAD contact frequencies up to 1 Mbp while Sanborn et al does not seem to be quantitative beyond the TAD-scale; or why Fudenberg et al observed generally symmetric stripes with corner-peaks as in Fig.1B, while Sanborn et al mainly predicted corner-peaks as in Fig.1A.

Building on their polymeric loop extrusion process by cohesin SMCs during interphase, Mirny's group also considered the role of condensin SMCs in the organization of prophase and mitotic chromosomes [69, 70]. They showed that a high concentration of LEFs (1 LEF every 30 kbp) coupled to a high processivity ( $\sim 800$  kbp) leads to a dramatic compaction of chromosomes that naturally become elongated and are constituted of a dense array of consecutive loops of average size 100 kbp without the need for BEs, recapitulating the initial results of Alipour and Marko [39] but with a more realistic and quantitative model. In particular, they observed that loop extrusion by condensin facilitates the disentanglement of entangled sister chromatids [70], in line with other works suggesting that loop extrusion may actively unknot polymeric chains [31, 71, 72]. A more recent study, coupling polymer modeling with loop extrusion and high-resolution Hi-C experiments performed at different stages of mitosis, allowed to quantify the distinct roles of condensin I and II in mitotic chromosome formation in mammals [73]: condensin II may act first by creating a helical central scaffold with large consecutive loops ( $\sim 400$  kbp), then, around this central axis, condensin I would compact the chromosome with shorter nested loops ( $\sim 80$  kbp). The loop-extrusion-induced chromosome compaction by condensin is also believed to occur in bacteria, in which polymer models [106, 113] accounting for actively translocating LEFs loaded close to the origin of replication, can recapitulate the properties of bacterial genome folding observed in Hi-C experiments [113].

## 2.2 Loop extrusion via diffusing slip-links

Despite numerous experimental studies in the last decade, the exact molecular mechanism of the formation of loops is not fully understood. Recent single-molecule studies illustrated the capacity of yeast condensins to actively extrude loops of naked dsDNA [40, 41], demonstrating the potential translocating motor activity of SMCs. However, it is still unclear if cohesin is capable of such a movement, in particular on chromatin [74]. Rather, *in vitro* experiments on cohesin observed a random, diffusion-like dynamics [75–77, 105].

To test whether the diffusion of LEFs may lead to TAD formation, Marenduzzo's group [78, 79, 107] developed a nonequilibrium model where LEFs bind and dissociate stochastically from chromatin, are free to diffuse as molecular slip-links [81] and are stabilized when reaching BEs (Fig.2E). Using first a 1D formulation and then generalizing it to integrate the 3D polymeric



structure (chromatin as a confined semi-flexible standard 30nm-fiber [80] with 3 kbp per monomer and LEFs as very rigid handcuffs), Brackley et al [78] showed that, in case of random loading of LEFs along the chain, the slip-links are distributed uniformly along the chromatin, which favors the formation of consecutive loop formation. As these loops compete with one another, they exhibit slow growth and the typical size of loops closed by the LEF decreases with the density of bound slip-links augments. This is incompatible with the formation of hundreds of kbps-long cohesin-CTCF loops observed for mammalian genomes. However, when LEF loading is limited to one specific site within a TAD, the model predicts the formation of nested loops. For the first-loaded slip-link, presence of all other bound diffusive LEFs on one of its sides and the total absence of LEFs on the other creates an osmotic pressure for the loop that may push the LEF towards its unoccupied side, i.e. towards the BEs. This osmotic ratchet leads to a dynamical, effective loop extrusion process and, contrary to random loading, generates a positive effect: more loaded slip-links result in larger loops. In particular, Brackley et al predicted that such positive osmotic effect may be observed if the 1D diffusion constant  $D$  of slip-links is greater than  $D \approx 0.025 \mu\text{m}^2/\text{s}$  which is well below the typical value ( $D \sim 0.1 - 0.25 \mu\text{m}^2/\text{s}$ ) measured *in vitro* [75–77]. They also observed that for a density of 1 LEF every 500 kbp and  $D \approx 5 \text{ kbp}^2/\text{s}$ , the formation of TADs with corner-peaks or stripes as large as 1.5 Mbp is possible via this process. A similar model based on a discrete 3D lattice developed by Miermans & Broderz [106] showed that the ratchet effect may also explain bacterial genome organization.

Concurrently to this work and following the same basic principle, a slightly different model was proposed by Yamamoto and Schiessel [82]. As in the work of Alipour and Marko [39], chromatin is treated as a 1D lattice, neglecting the 3D polymeric nature of the fiber. Each TAD is composed of a loading site at its center where cohesins may attach and unloading sites at domain borders where they dissociate. In addition to the loading of diffusive cohesin handcuffs (or dimers) as in [78], cohesin monomers (i.e. encompassing only one DNA strand, Fig.2E) may also associate at the same site and diffuse in 1D along the chain. These monomers also contribute to the osmotic pressure and thus reinforces the growth of loops entrapped by dimers, thus leading to loop extrusion. Hence, due to this extra contribution of monomers, the same osmotic pressure may be achieved with less loaded dimers.

While the mechanism of loop extrusion via diffusive slip-links is in line with *in vitro* experiments of the movement of single cohesin unit on chromatin [75–77] and shows qualitative agreement with Hi-C data in mammals and in bacteria, it is still unclear whether such a mechanism is able to quantitatively predict the intra-TAD and inter-TAD scaling laws for the contact probability  $P(s)$  (Fig.1C,D) as well as to predict TAD organization and loops depending on CTCF motif orientations or perturbation experiments in which CTCF or cohesin concentrations were modified [34]. Moreover, the osmotic ratchet effect requires the presence of only one loading site per TAD. While Chip-Seq profiles of Nipl (cohesin loader) exhibit peaks in TADs surrounding active genes at gene promoters, Nipl may have many secondary fixing sites or TADs with several active genes will have several Nipl peaks. However, a recent version of the diffusive slip-link model suggests that LEF loading at random positions may also lead to corner-peaked TADs [107].

### 2.3 Loop extrusion via transcription-induced supercoiling

As in the previous section, based on the lack of experimental proof for a direct translocation activity of cohesin SMCs, Stasiak's group suggested that transcription-induced negative supercoiling might be the motor for loop extrusion by cohesin [83, 84]. Previously, inspired by

several studies suggesting that interphase chromosomes are torsionally stressed [85–87], the same group has developed a polymer model that accounts for possible supercoiling present within TADs [88]. They modeled chromatin as a confined (with periodic boundary conditions) bead-on-string self-avoiding polymer sensitive to torsional and bending stress, [with](#) TAD borders playing [the](#) role of anchors by maintaining a fixed stress within one domain. They showed that imposing a linking number difference of -2 turns per 100 kbp leads to the formation of plectonemes that enhances the intra-TAD contact frequency with scaling laws compatible with experiments (Fig.1C), but the model failed to generate corner-peaks.

To better describe corner-peak TADs and the important role of cohesin and CTCF in TAD formation, Racko et al proposed (Fig.2F) that cohesins are loaded between the extremities of a transcribed region (a gene or an enhancer for example) within a TAD [83] and that a global negative torsional stress is produced by transcribing RNA polymerases [85], which can be relaxed by topoisomerase II B present at TAD borders. Transcription-induced stress leads to the formation of plectonemes between the active region and the cohesin SMC, transient stabilization of these plectonemes being made possible [by the large hydrodynamic drag of the SMC anchoring the forming loop](#). This results in growing plectonemes that may push cohesin complexes towards BEs where the stress is [eventually](#) released by TOP2B. Cohesins reaching well-oriented BEs are then stabilized by interactions with bound CTCF. To test this mechanism, Racko et al developed a polymer model (hundreds of kbp in length with 400 bp per monomer) having a strong torsional persistence length that allows [for](#) the formation of plectonemes and limits the relaxation of the stress by internal twist; generation of negative supercoiling (5 to 10 turns) is performed by an active, rigid swivel positioned at the transcribed region; relaxation at BEs is allowed by passive swivels; cohesin SMCs being modeled as rigid handcuffs as in [78]. Movies extracted from molecular simulations of this model illustrated how the resulting flux of supercoiling (balance between creation at the transcribed region and relaxation at BE) may push SMCs in TAD border directions. Though the transcription is essential for the TAD formation, the model suggested that once the extrusion is complete and cohesin complexes have reached both BEs, the transcription is not required any more to maintain the TADs.

However, the lack of quantitative comparisons with Hi-C data and of a precise impact analysis of model parameters makes it difficult to evaluate the validity of the proposed mechanism. While the enrichment of TOP2B at CTCF BEs is well documented [89, 90], the association between supercoiling domains observed in [85, 86] and TADs is still unclear. For example, a recent study showed that TOP2B activity at BEs is mainly transcription-independent and suggested that topological stress tends to occur at both sides of the BEs but not intra-TAD [90]. [The main](#) ingredient in the Stasiak's group model is the capacity of the polymer to absorb supercoiling stress by forming plectonemes instead of by twisting. This requires a strong torsional persistence length ([~100 nm corresponding to bare DNA](#)) well beyond the actual experimental estimations (5 nm) [91] showing that short pieces of chromatin (10 kbp) can absorb many turns (40) before forming plectonemes [92]. Moreover, it is unclear how this mechanism of transcription-induced loop extrusion accounts for the cohesin-mediated formation of TADs with corner-peak in untranscribed or heterochromatic regions [44–46] and how it is consistent with the many perturbation experiments on BE locations and CTCF or cohesin levels.



## Conclusion and Discussion

Polymer models with various mechanisms and parameters have been proposed to model the generation of loops by extruding factors. All these models state that the formation of TADs and sub-structures like corner-peaks emerge from the dynamic translocation of SMC complexes. SMCs encompass two strands of chromatin and slide from the interior of the TAD to its boundary elements. However, the main physical ingredient leading directly or indirectly to this translocation differs between models: in [31, 32], cohesins are directly actively and unidirectionally translocated; in [78, 82], diffusive cohesins are translocated towards a preferential direction due to the osmotic pressure created by other cohesins diffusing on chromatin; in [83], transcription-induced supercoiling generates growing plectonemes that will push cohesin up to boundary elements. [In addition](#), the stoichiometry of the extruding complexes (one or two SMC rings), the number of walking legs of [an extruder](#), the possible self-interaction between motors and the molecular events leading to ATP-driven DNA capture and translocation are still unclear [74, 93–102]. Quantitative comparisons between these different modeling studies suffer from the use of different null polymer models to describe chromosomes, each assuming or using different properties to coarse-grain and simulate chromatin fibers. [The](#) development of a common theoretical framework based on a realistic null model [65] will certainly help to test the various existing hypotheses on loop extrusion under the same conditions. Another key point in these models is that, in order to explain the formation of corner-peaks only between convergent CTCF motifs, extruding factors are assumed to have differential interactions with these motifs depending on their orientations: either these sites act as a road-block or are passive in [31, 32], either they interact with extruders or reflect them in [78]. [The](#) molecular details of this asymmetric, orientation-dependent effect still need to be investigated.

All this [highlights](#) the strong uncertainties existing about the correct molecular mechanisms responsible for loop extrusion. [The most recent molecular experiments tend to suggest an active mechanism with only one walking leg \[40\] without steric hindrance between LEFs \[41\], but it was observed only for yeast condensin on dsDNA. Does it apply to cohesin whose motion seems more diffusive \[75-77\]? Does it happen on chromatin \[100\]? Future single-molecule studies on reconstituted chromatin in the presence of key controlled components \(CTCF, TOP2B, etc.\) measuring the dynamics but also the involved forces and torques will certainly help to resolve these ambiguities.](#)

While other mechanisms driving chromosome organization like epigenomic-driven interactions, phase-separation or promoter-enhancer loops also start to be well characterized, it remains unclear how these different processes might interfere with loop extrusion. For example, Hi-C experiments on cohesin-depleted cells illustrated the loss of TAD organization replaced by a stronger compartmentalization between active and inactive compartments driven by epigenetics [45], indicating that loop extrusion process may destabilize compartment formation via active mixing [62]. Similarly, chromatin is also the template for many other motors like RNA polymerases during transcription, DNA polymerases during replication or translocases like Rad54 during DNA repair. It will be of course of interest to study the possible crosstalk between these processes. Along this line, recent experiments showed that [transcription may influence cohesin localization \[108,109\] or translocation \[77, 103\]](#) but may also act as moving barriers against condensin progression [104]. Polymer physics-based models would certainly be very useful to [probe](#) such coupling and to suggest new exciting experiments. This will help to [elucidate](#) the contribution of loop extrusion in

chromosome organization during interphase and mitosis and within different organisms, from bacteria to humans, in which SMC proteins play multiple roles, suggesting that the mechanisms of dynamic loop formation are generic and may be crucial to genome-folding regulation in all kingdoms of life.

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## Author biography

Surya K. Ghosh is Assistant Professor at SRM Institute of Science and Technology. His research interest is on the modeling of various biological phenomena: from the diffusion of single molecules to the organization of long polymers.

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## Figures

**FIG. 1:** (A, B) Pieces of Hi-C maps of chromosome 4 (A) and 14 (B) for the human cell line GM12878 [7] illustrating the presence of corner-peaks (arrows in A) or of stripes (arrows in B) at TAD boundaries. For visual inspection, corresponding Chip-seq signals for CTCF and cohesin, as well as the position and orientation (green: sense, red: antisense) of CTCF motifs were added. Panels A and B were built using the webtool Juicebox [50]. (C, D) Average contact frequency  $P(s)$  between two loci separated by a genomic distance  $s$ : intra-TAD frequencies for TADs of different sizes in presence or not of corner-peaks (C) and inter-TAD frequency (D). Dashed lines represent scaling laws  $P(s) \propto s^{-\gamma}$ .

**FIG. 2:** Different models for loop extrusion. (A) If the genomic distance between two oriented motifs is larger than the torsional persistence length of chromatin, no preferential orientation of interaction will be observed for the pair. (B) Nasmyth & Riggs model for mitotic condensation by condensins resembling a loop extrusion process: condensins bind to chromatin and form an array of consecutive loops and an axial core. (C) Loop extrusion by active, unidirectional translocating motors: LEFs are loaded to and unloaded from chromatin randomly, the two 'legs' of a LEF walk in opposite directions, one leg is halted when reaching a CTCF binding motif oriented opposite to its translocation direction. (D) Representation of different types of local organization as a function of LEF concentration and processivity. For low concentrations and processivities, sparse arrays of contiguous loops of small sizes dominate. As the concentration is increased, arrays become denser; as the processivity is increased, loops become larger, with eventually, the presence of stacked loops. (E) Loop extrusion by diffusive slip-links: SMCs are loaded as dimers or monomers at specific sites and diffuse along the chromatin, dimers interact with boundary elements oriented opposite to their motion while being reflected by BE in the other direction. (F) Loop extrusion by transcription-induced supercoiling: LEFs bind to a specific – transcribed – region, supercoiling

generated by transcription leads to plectoneme formation that push LEFs which are stabilized at BE.

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