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Enrique González-Bautista, Enrique Alarcón-Gutierrez, Nathalie Dupuy, Isabelle Gaime-Perraud, Fabio Ziarelli, et al.. Influence of yeast extract enrichment and *Pycnoporus sanguineus* inoculum on the dephenolisation of sugar-cane bagasse for production of second-generation ethanol. *Fuel*, 2020, 260, pp.116370. 10.1016/j.fuel.2019.116370 . hal-03176788

HAL Id: hal-03176788

<https://hal.science/hal-03176788>

Submitted on 22 Mar 2021

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Short communication

Influence of yeast extract enrichment and *Pycnoporus sanguineus* inoculum on the dephenolisation of sugar-cane bagasse for production of second-generation ethanol

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A B S T R A C T

Pretreatment of sugar-cane bagasse (SCB) by laccases from *Pycnoporus sanguineus* under solid-state fermentation (SSF) can favour dephenolisation of the substrate prior to ethanol production. To define adequate conditions for this biological pretreatment, an experimental design and surface plot analysis were performed using as independent variables: yeast extract, fungal inoculum and substrate amounts after different incubation times. Laccase and cellulase activities and chemical characterisation of the substrate by ¹³C CP/MAS (cross-polarization at magic angle spinning) NMR (nuclear magnetic resonance) were used as responses. After 60 days of SSF, yeast extract enrichment and low amounts of inoculum enhanced dephenolisation of SCB. Moreover, yeast extract reduced the quantity of crystalline cellulose without strongly promoting cellulases and polysaccharide degradation. High amounts of inoculum decreased laccase activities and substrate dephenolisation.

1. Introduction

Production of second-generation bioethanol is based on the valorisation of agricultural wastes, like sugarcane bagasse (SCB). For instance, 18 Mt of SCB were produced in 2018 in Mexico [1]. Since ethanol production yield can reach 0.25 g/g of dry biomass of SCB [2], production of second-generation ethanol based on SCB can be considered as a relevant and sustainable technology that could feed Mexican demand for this type of resource. However, lignocellulosic material requires a pretreatment i.e. dephenolisation, in order to make the cellulose fibres accessible for the next step of fermentation. Enzymatic pretreatment appears to be a suitable eco-friendly technology, not requiring chemical or physical treatments, using low-energy input and mild operating conditions [3].

Dephenolisation of the complex polyphenolic structure of lignin can be performed by white-rot fungus like *Pycnoporus sanguineus*, which produces mainly laccases as phenoloxidases [4]. Laccases are particularly interesting enzymes for industrial applications since they do not require any co-substrate (neither H₂O₂ nor Mn), and their activities remain stable for several days [5]. In general, production of laccases has

been shown to depend on the nature of nitrogen sources as well as on C/N ratio [6]. Generally, inorganic nitrogen sources lead to low laccase activities, while organic nitrogen sources result in high laccase yields [7]. Moreover, it seems that laccase activities could increase by supplementation of an enriched organic nitrogen source [8]. There are several reports using yeast extract as an enrichment to enhance production of laccase during SSF. Niladevi et al. [9], studied laccase activities of *Streptomyces psammoticus*, and showed an increase in these enzyme activities of 86% using yeast extract compared to NH₄Cl and of 39% compared to tryptone. Also, Fenice et al. [10], increased up to 22% laccase activities and up to 15% biomass production of *Panus tigrinus* with an enrichment of 0.1% of yeast extract and 0.5% of saccharose. Moreover, enrichment with yeast extract can induce different isoforms of laccases and enhance laccase activities overall [11]. The time of incubation and the amount of initial inoculum can also influence the expression of laccases and consequently dephenolisation of the SCB. It has been observed that during the first steps of SSF, laccase activities increased [8], and aromatic degradation can be observed days after inoculation occurred [12].

This study aimed to define the conditions of substrate pretreatment

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which favoured dephenolisation of SCB. A response-surface experimental design was set up to test the effects of yeast extract, the amount of fungal inoculum and of substrate, on the kinetic of SCB dephenolisation by *Pycnoporus sanguineus*. As responses, laccase and cellulase activities were measured and transformation of lignocellulose was evaluated via solid-state ^{13}C CP/MAS NMR.

2. Materials and methods

2.1. Strains and inoculum

The strain of *Pycnoporus sanguineus* used was isolated from a wild mushroom found in the sugar-cane fields of Jalcomulco, Veracruz, Mexico (19° 20' 00" N, 96° 46' 00" W), registered as F. Ramírez-Guillén 932 in the XAL herbarium (INCOL, A.C.) and identified by analysis of ITS (internal transcribed spacer) fragments 1 and 4 (Access Genebank KR013138). The fungal strain was reactivated in plates of malt extract agar 1.5% at 30 °C for seven days. Then, five agar pieces of 1 cm diameter were used as inoculum for mycelial colonisation of wheat grains (10 days at 30 °C). Wheat grains were prepared as follows: 100 g of wheat grains were complemented with 150 ml of water and 0.5 g of gypsum and then sterilized by autoclaving [13].

2.2. Mesocosm preparation

The substrate was composed of 90% of SCB complemented with 10% of coffee pulp used as an inductor of laccases [14,15], which was previously milled and sterilized twice by autoclave. Dried and milled SCB (particles from 0.5 to 3 cm) was received from Mahuixtlan sugar refinery, located at Mahuixtlan, Veracruz, Mexico. The composition of this by-product was analysed by ^{13}C CP/MAS NMR by Hernández et al. (2017): O-Alkyl 79.76%, aromatics 12.87%, COOH 3.57% and Alkyl 2.45%. The coffee pulp was obtained from a shade-grown commercial coffee culture with the dominant variety *Coffea arabica* [16], located in the Tlacontla farm, San Marcos de León, municipality of Xico, Veracruz, Mexico. Each mesocosm was composed of 70 g of substrate (dry weight, DW) in 0.5 L aluminium plates (10*14*5 cm³), covered with an aluminium foil. For each treatment, three replicates were made, to obtain a total of 99 mesocosms and incubated at 30 °C at 80% of the water holding capacity (WHC) in the dark. 33 mesocosms were analysed after 20, 40 or 60 days of incubation.

2.3. Experimental design

Three-factor experimental design was performed using as independent variables: the amount of yeast extract, inoculum and substrate using 4 levels and a full-cubic model (Table 1). The concentrations of yeast extract ranged between 0.7–9.1% DW (from 0.7 to 9.1 g) and the concentrations of inoculum from 5.0 to 14.3% DW (from 5.0 to

Table 1

Factors and levels evaluated in the experimental design (yeast extract ranging from 0.7–9.1% DW and fungal inoculum from 5.0 to 14.3% DW and substrate from 85.6–94.3%).

Conditions	Inoculum (%)	Yeast extract (%)	Substrate (%)
1	8.1	6.3	85.1
2	8.1	3.5	88.4
3	5.0	9.1	85.9
4	14.3	0.7	85.0
5	5.0	6.3	88.7
6	8.1	0.7	91.2
7	5.0	3.5	91.5
8	5.0	0.7	94.3
9	8.1	3.5	88.4
10	11.2	0.7	88.1
11	11.2	3.5	85.3

14.3 g). The rest of the mesocosm was composed of the substrate to keep the mesocosm weight constant (85.6 – 94.3%). Sampling was performed at 3 incubation times (20, 40 and 60 days of incubation). The dependent variables measured were: laccase and cellulase activities, and chemical properties of the substrate by solid-state ^{13}C CP/MAS NMR.

2.4. Enzymatic activities

Laccases and cellulases were quantified after 20, 40 and 60 days of incubation. The reaction mixtures for laccase were performed according to Farnet et al. [17], with syringaldazine (15 μM in methanol) as substrate. Cellulase activity was assayed using CarboxyMethylCellulose (CMC) at 1% as substrate according to Somogyi Nelson method modified by Farnet et al. [18]. One unit (U) of enzyme activities is defined as one μmole of the product formed per min and per g of dry weight (DW).

2.5. Solid-state ^{13}C CP/MAS NMR

The chemical composition of SCB, coffee pulp and of each mesocosm was characterized by solid-state ^{13}C CP/MAS NMR, on a spectrophotometer Bruker DSX 400 MHz operating at 100.7 MHz. Samples (400 mg) were spun at 10 kHz at the magic angle. Contact times of 2 ms were applied with a pulse width of 2.8 μs and a recycle delay of 3 s. Chemical shift values were referenced to tetra-methyl-silane and calibrated to glycine carbonyl signal set at 176.03 ppm. The relative C distribution in ^{13}C NMR spectra was determined by integrating the signal intensity in different chemical shift regions [19], with an integration routine supplied with Dmfit 2003 software. Seven common chemical shift regions were defined according to Mathers and Xu [20]. A decomposition index (Alkyl-C/O-Alkyl-C) was calculated according to Baldock et al. [21], as the humification ratio and the crystallinity of cellulose was also measured using the relative percentage of crystalline cellulose peak (90 ppm) vs the sum of the relative percentages of amorphous (83 ppm) and crystalline cellulose [22].

2.6. Statistical analyses

Correlations between variables and the factors tested were determined using UNSCRAMBLER 10*3 (CAMO). To find out whether the independent factors had significant effects on each response, a three-way ANOVA (analysis of variance) was carried out. Homoscedasticity and normality were analysed previously. The interaction effects of yeast extract, inoculum and substrate amounts were analysed by response surface methodology [23]. Pearson's correlation coefficients were calculated between all the NMR peaks from each 33 mesocosms after 60 days of incubation and a principal-component analysis (PCA) was performed to describe chemical changes of the substrate depending on the conditions tested.

3. Results and discussion

3.1. Enhancing laccase activities and dephenolisation of SCB

According to the surface plots (Fig. 1A), laccase activities were favoured by higher amounts of yeast extract during fermentation ($P = 0.001$). This complementation rich in amino acids, vitamins, sugars and ions like Cu^{2+} could enhance fungal metabolism [24] and increase production of multicopper oxidases such as laccases [25]. Here, yeast extract was used to test the effect of substrate complementation by a complex source of nitrogen and growth factors and this study revealed that it can indeed enhanced laccase activities. Nevertheless, for an industrial approach, a sustainable and affordable complementation using agroindustrial by-products should be implemented. Zimbardi et al. [4], used wheat straw (a culture substrate

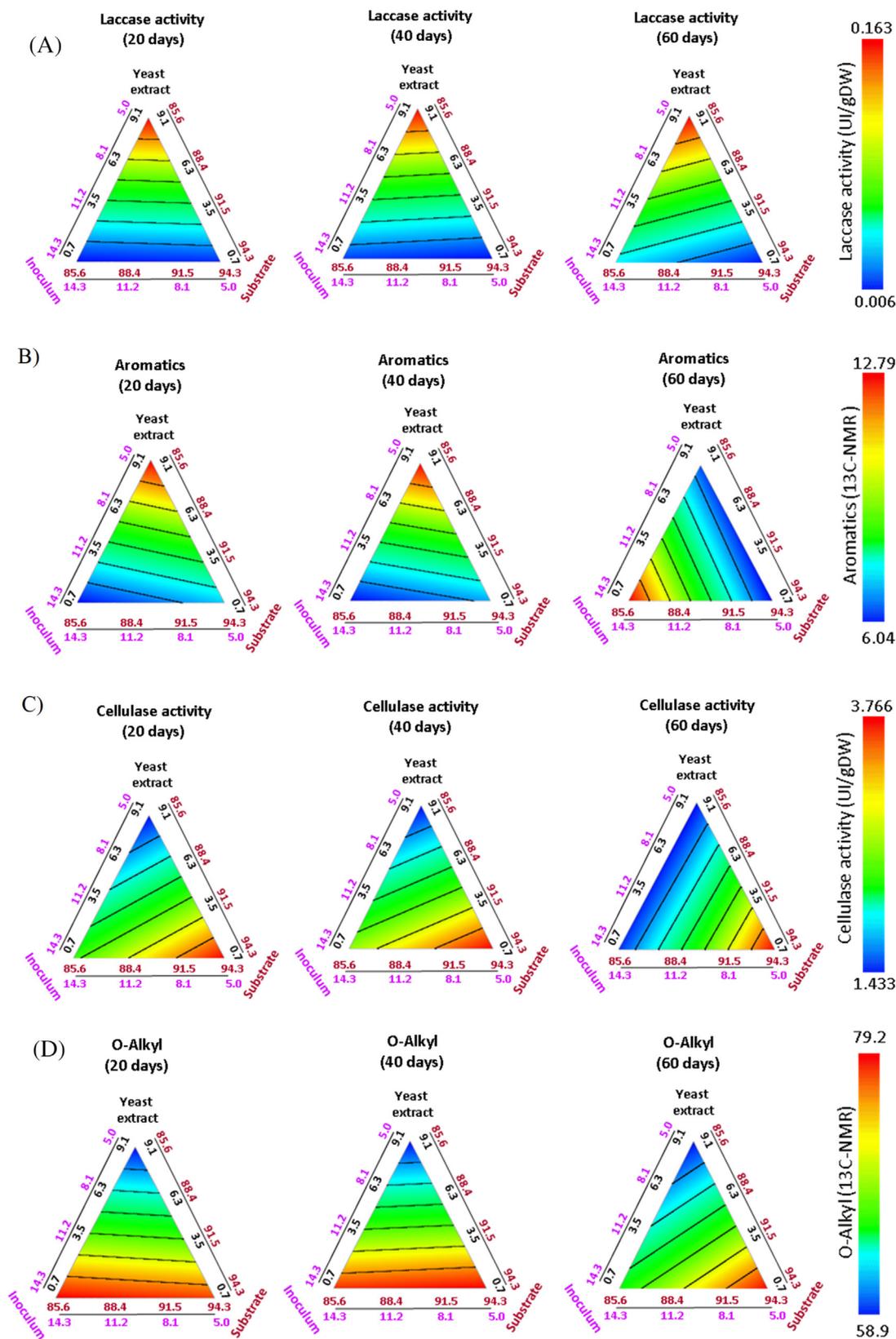


Fig. 1. Surface plots of the interaction between yeast extract (YE), amount of inoculum and substrate for A) Laccase activities, B) Total aromatics, C) Cellulase activities and D) O-Alkyl responses after 20, 40 and 60 days of incubation.

poor in nitrogen) complemented with malt extract, to increase laccase production in *Phlebia floridensis* by 16% compared with control (no complementation). The addition of wheat bran can also be a suitable option to enrich the substrate, since it is composed of 18% of proteins

and is not broadly consumed by humans [26]. Moreover, we found that low amounts of inoculum enhanced laccase activities. Several studies about kinetics of laccase production reported a quick increase in laccase activity during the first days of SSF followed by a decrease in activity.

For instance, Asgher et al. [27], found higher laccase activities after only 5 days of solid-state culture of *Trametes versicolor* IBL-04, followed by a 30% decrease after 10 days of fermentation. Increasing the proportion of inoculum during SSF could accelerate this trend and thus rapidly lead to a decrease in laccase production. Moreover, here, a high concentration of inoculum (almost 15%) is linked to a lower amount of substrate (composed of 90% SCB and 10% coffee pulp). Phenolic compounds from coffee pulp are known to induce laccase activities [28]. In our previous study [15], we found that addition of 10% of coffee pulp to SCB induced the expression of a new isoform of laccase in *Pycnoporus sanguineus* as revealed by SDS-PAGE (an induced isoform with Rf = 0.19 compared to laccases constitutively produced with Rf = 0.23 and 0.27). During substrate autoclaving, an extraction of monophenolic compounds from coffee pulp have probably occurred, which can increase laccase activities of white-rot fungi [29], and thus a reduction in coffee pulp (in the cultivation substrate) could provoke a decrease in such activities.

Analysing the surface plots of total aromatics in the substrate, we observed a favourable effect ($P = 0.0002$) of yeast extract complementation (47.2% of decrease in aromatics) between 40 and 60 days (Fig. 1B). Thus, the effect of laccase enhancement (0.16 U/gDW) by adding yeast extract on dephenolisation was observed after 40 days of incubation. Meza et al. [30], observed delignification of SCB after 30 days of incubation of *Pycnoporus cinnabarinus* after adding different laccase enhancers (ferulic acid and ethanol vapours). Moreover, Dong et al. [31], monitored laccase production kinetic using different white-rot fungi (*Pleurotus ostreatus*, *Lentinula edodes* and *Phanerochaete chrysosporium*) under SSF of SCB and obtained high laccase activities and delignification after 42 days of incubation. These findings are of huge importance since substrate dephenolisation is a key step to enhance cellulose availability and thus hydrolysis yields for production of bioethanol. Rico et al. [32], pretreated chips of wood (*Eucalyptus globulus*), reaching dephenolisation up to 50%. The pretreatment enhanced by 40% the amount of glucose obtained after saccharification. Higher concentrations of free fermentable sugars can further increase bioethanol yields.

3.2. Cellulase activities and polysaccharide degradation

The surface plot analysis of cellulase activities showed that adding yeast extract reduced these enzymatic activities ($P = 0.003$) over all the incubation time (Fig. 1C). This could be related to catabolic repression due to monosaccharides from yeast extract [33]. For instance, Khalil et al. [34], found lower activities of CMCase, β -glucosidase and xylanases when 1% of glucose was added under SSF of SCB. Interestingly, high amounts of inoculum (14.3%) favoured cellulase activities (from 1.43 to 3.76 U/gDW) at the beginning of the incubation (from 1 to 40 days). According to Dinis et al. [35], production of cellulases from white-rot fungi mainly occurred during the first days of SSF in *Phlebia rufa*, *Trametes versicolor* and *Bjerkandera adusta*. Moreover, this result is in accordance with Kachlishvili et al. [8], who reported a link with high fungal biomass and high cellulase activities.

Polysaccharide degradation of the substrate was analysed by surface plots of O-Alkyl (Fig. 1D) and showed that significant differences were found after 20 ($P < 0.05$) and 60 ($P < 0.001$) days of incubation. When high concentrations of yeast extract were added (9.1%), O-Alkyl decreased (25.7%) and this cannot be related to cellulases: these activities were lower with high amounts of yeast extract as described above. This may be explained by lower amounts of inoculum and substrate (5% and 85% respectively) both rich in polysaccharides (starch and cellulose respectively). On the other hand, when high amounts of inoculum were used, (14.3%) a high quantity of O-Alkyl signal was found during the first days of fermentation (1 to 40 days) and a decrease was observed after 60 days of incubation. This can be explained by the high levels of cellulase activities observed (1.43 to 3.76 U/gDW) with high amounts of inoculum, leading to

polysaccharide degradation at the end of the SSF i.e. 60 days (Fig. 1D). Quiroz-Castañeda et al. [36], found that the level of cellulases activities of *Pycnoporus sanguineus* and *Bjerkandera adusta* was related to the diameter of fungal growth in Petri dishes using different lignocellulosic substrates as carbon sources. This is corroborated by the variation in crystallinity ratio (qualifying the degree of depolymerisation of cellulose) which followed the same pattern over incubation time ($P < 0.05$, after 40 days and $P < 0.0001$ after 60 days, data not shown).

Another interesting result is linked to the Alkyl/COOH ratio (Data not shown). This marker of fungal biomass [37] increased (up to 1.67) with higher amounts of inoculum (14.3%) after 20 days of incubation ($P < 0.05$). This result can be related with high cellulase activities (3.76 U/gDW) observed in these conditions. Dong et al., [31], found a direct relationship between fungal biomass production and cellulases activities during SSF. This increase in cellulase activities provided more carbon to produce fungal biomass [38]. Interestingly, after 40 days of incubation ($P < 0.0001$), yeast extract addition led to an increase in fungal biomass, probably because of the role of nutrient enrichment. Kachlishvili et al. [8], proved that adding yeast extract during SSF increased the biomass of *Funalia trogii*, *Lentinus edodes* and *Pleurotus dryinus*.

Fig. 2 shows the PCA performed from all the peaks defining the spectra of solid-state ^{13}C NMR data of mesocosms after 60 days of incubation. The first two principal components (PCs) accounted for 65.6% of the total variation in the data (PC1 accounts for 36.0% and PC2 for 29.6%). Projections with the higher amount of yeast extract (from 6.3 to 9.1%) and lower amount of inoculum (from 5 to 8.1%) were discriminated from others. They were positively correlated to a peak (116 ppm) from the aromatic region and were negatively correlated to a peak (159 ppm) from phenol region. This result gives evidence of dephenolisation of the SCB under these conditions. This can be related with the previous results observed i.e. a decrease in total aromatics (aromatics plus phenols) after 60 days of incubation (Fig. 1B) when a high amount of yeast extract and a low amount of inoculum are used.

4. Conclusion

Our findings defined more favourable conditions for SCB pretreatment prior to saccharification and ethanol fermentation: high amounts

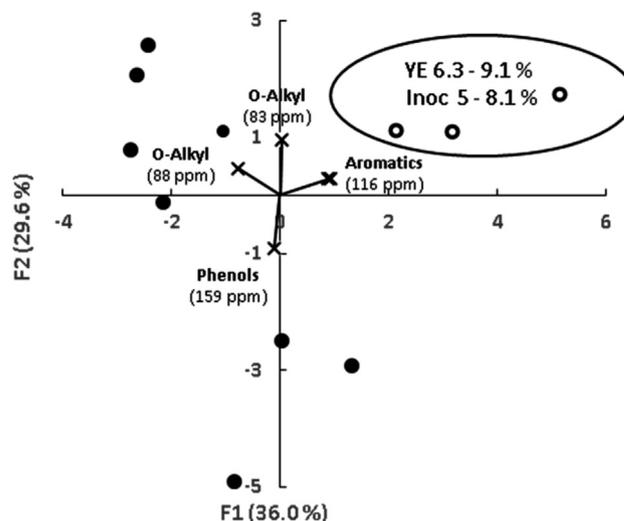


Fig. 2. Principal component analysis of the CP/MAS NMR peaks from the spectrum of each mesocosm after 60 days of incubation. White circles correspond to conditions with lower amount of inoculum and higher amount of yeast extract, black circles represent other concentrations. Each circle is a centroid representing the average of three replicates, standard deviations were below 0.1. Explaining variables (CP/MAS NMR peaks) are indicated.

of yeast extract, low amounts of inoculum and an incubation time of 60 days of SSF. Under these conditions, laccase activities of *Pycnoporus sanguineus* and dephenolisation of the substrate were enhanced, meanwhile no increase in cellulase activities and thus no extensive hydrolysis of polysaccharides were observed. These pretreatment conditions are thus beneficial for the further step of saccharification, since making cellulose more available will help at enhancing bioethanol production yields.

Acknowledgments

The authors are grateful to the National Council of Science and Technology (CONACYT) of Mexico, for the doctoral scholarship of Enrique González Bautista no. 440596 and to ECOS-ANUIES-CONACYT Program through the ECOS project M13A02, for financial support. We would like to thank Mrs. Lisa Foli for her very helpful technical assistance.

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