

Aquatic mesocosm strategies for the environmental fate and risk assessment of engineered nanomaterials

Andrea Carboni, Danielle L. Slomberg, Mohammad Nassar, Catherine Santaella, Armand Masion, Jérôme Rose, Melanie Auffan

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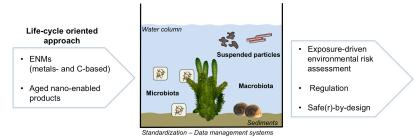
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| 1 | Aquatic mesocosm strategies for the environmental fate and risk assessment of engineered |
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| 4 | Andrea Carboni ^{1*} , Danielle L. Slomberg ¹ , Mohammad Nassar ¹ , Catherine Santaella ³ , Armand |
| 5 | Masion ¹ , Jerome Rose ^{1,2} and Melanie Auffan ^{1,2} |
| 6 | |
| 7 | ¹ CNRS, Aix-Marseille Univ., IRD, INRAE, CEREGE, 13545 Aix-en-Provence, France |
| 8 | ² Civil and Environmental Engineering Department, Duke University, 27707 Durham, NC, |
| 9 | United States |
| 10 | ³ Aix Marseille Univ, CEA, CNRS, BIAM, LEMiRE, Laboratory of Microbial Ecology of the |
| 11 | Rhizosphere, ECCOREV FR 3098, F-13108 Saint Paul-Lez-Durance, France |
| 12 | |
| 13 | |
| 14 | * Corresponding author |
| 15 | Andrea Carboni |
| 16 | Centre de Recherche et d'Enseignement de Géosciences de l'Environnement (CEREGE), |
| 17 | Technopole Environnement Arbois-Mediterranee, BP80, 13545 Aix-en-Provence, Cedex 04, |
| 18 | France |
| 19 | Phone: (+33) 04 42 97 15 00 |
| 20 | Email: carboni@cerege.fr |
| 21 | |
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| 28 | This review explores aquatic mesocosm outcomes in the study of engineered nanomaterial |
| 29 | fate and effects, across their life-cycle and in an environmental risk perspective. |
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35 Table of Contents Art

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Abstract

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In the last decade, mesocosms have emerged as a useful tool for the environmental study of 42 43 engineered nanomaterials (ENMs) as they can mimic relevant exposure scenario of contamination. Herein, we analyzed the scientific outcomes of aquatic mesocosm 44 45 experiments, with regard to their designs, the ENMs tested, and the endpoints investigated. 46 Several mesocosms designs were consistently applied in the last decade to virtually mimic 47 various contamination scenarios with regard to ecosystem setting as well as ENMs class, dose and dosing. Statistical analyses were carried out with the literature data to identify the 48 main parameters driving ENM distribution in the mesocosms and the potential risk posed to 49 benthic and planktonic communities as well as global ecosystem responses. These analyses 50 showed that at the end of the exposure, mesocosm size (water volume), experiment 51 52 duration and location indoor/outdoor had a major role in defining the ENMs/metal partitioning. Moreover, a higher exposure of the benthic communities is often observed but 53 did not necessarily translate to a higher risk due to the lower hazard posed by transformed 54 ENMs in the sediments (e.g. aggregated, sulfidized). However, planktonic organisms were 55 generally exposed to lower concentrations of potentially more reactive and toxic ENM 56 species. Hence, mesocosms can be complementary tools to existing standard operational 57 58 procedures for regulatory purposes and environmental fate and risk assessment of ENMs. To 59 date, the research was markedly unbalanced toward the investigation of metal-based 60 compared to metalloid and carbon-based ENMs but also nano-enabled product. Future 61 studies are expected to fill this gap, with special regard to high production volume and 2 62 potentially hazardous ENMs. Finally, to take full advantage of mesocosms, future studies 63 must be carefully planned to incorporate interdisciplinary approaches and ensure that the 64 large data sets produced are fully exploited.

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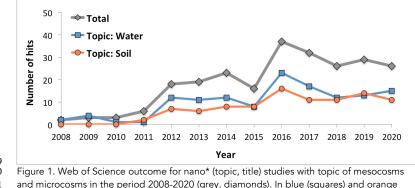
67 1. Introduction

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69 Assessing the environmental impact and hazard of engineered nanomaterials (ENMs) is 70 challenging due to their specific physico-chemical properties compared to bulk and free-ion 71 species^{1,2}. In the last decade, an extensive amount of research has been carried out to understand ENM fate and effects upon release in the environment. The majority of these 72 studies employed standardized experimental strategies, usually addressing single endpoints 73 in simplified systems (e.g. single-species ecotoxicity tests, one-parameter dependent 74 transformation), to provide straightforward information for regulatory purposes. However, 75 such approaches are often not representative of true environmental complexity and may not 76 77 reproduce mechanisms driving ENM behaviour and effects in the ecosystem. In recent years, efforts have been made on the development and use of alternative testing strategies (ATS) 78 that enable the study of ENM environmental risk in more realistic frameworks, e.g. by 79 including the aging of ENMs in natural matrices as well as their interactions with natural 80 components during trophic or trans-generational transfers^{2,3,4}. Among the ATS, mesocosms 81 are considered invaluable tools, allowing for a more comprehensive simulation of key 82 83 ecological processes compared to other single standardized test or standard operational 84 procedure (SOP)^{2,5,6,7}. 85 Mesocosms can be defined as an enclosed and essentially self-sufficient experimental environment and have been extensively used to investigate ecological processes and the 86 87 environmental fate and effects of xenobiotic chemicals^{8,9,10}. Mesocosms allow (I) self-88 sustainability for the duration of the experiment, (II) feasibility of sampling and measurement 89

of multiple bio-physico-chemical endpoints and (III) the possibility to identify direct and indirect effects of the tested compounds^{5,11,12}. Such strategies have already been implemented to generate regulatory data in the evaluation of pesticide contaminations^{13,14} and have recently been proposed for ENM environmental risk assessment (ERA). Regarding ENM ERA, mesocosms enable to simultaneously investigate ENM behavior and effects using an exposure scenario-driven approach^{5,15}. They provide exposure and hazard data that account for ecological synergies but also interdependent transformation processes⁵. This is particularly relevant, as the validity of conventional standardized tests is challenged by the

- unique properties of ENMs and their dynamic evolution upon release in the ecosystem¹. 97 Exposure and hazard data obtained in ENMs dosed mesocosms can be gathered in 98 database and fed into fate models, hazard scoring schemes, and environmental risk 99 100 assessment models^{16,17}. They can also serve as input for high throughput experiments to 101 better elucidate the mechanisms involved². 102 While a definition of mesocosms has been debated with regard to the operational time and 103 size, terms such as microcosm and mesocosms have interchangeably been used in literature 104 for describing similar systems. A literature survey in 2012 focused on ENM studies in both terrestrial and aquatic mesocosms, reporting only twelve instances of published works¹⁸. 105 106 However, during the last decade, the use of mesocosms in ENM fate and nanosafety studies has consistently increased with more than 200 research items between 2008-2020 (source: 107 Web of Science; keywords nano*, mesocosms and microcosm) (Fig. 1). These works 108 investigated ENMs in either aquatic or terrestrial mesocosms and a part of these items have 109 already been reviewed with a focus on terrestrial environments¹⁹ and ENM toxicity⁴. 110 Here, we reviewed papers focused on fate and impact of ENMs in aquatic mesocosms to 111 provide an overview of the relevant strategies, mesocosms designs, and their findings. An 112 113 extensive screening of these papers was carried out in order to select (I) experimental mesocosm investigating ENMs and (II) studies focusing on the aquatic ecosystem. A final 114 selection of about fifty peer-reviewed papers was obtained excluding short-term 115 experiments (<1 week) with no acclimation time as well as single-endpoint and single-species 116 oriented studies, which have been already reviewed by Bour et al. (2015)⁴. A detailed list of 117 the selected literature and further information of the selection criteria are reported in the 118 Supporting Information (Table S1). A further aim of the present work is to relate the ENM 119 fate and nanotoxicity for aquatic organisms to the environmental risk in relevant 120 environmental scenarios. Statistical tools were employed to identify important parameters 121 determining the long-term partitioning of ENMs in aquatic environments. This will be used as 122 the basis to discuss ENMs exposure towards benthic and planktonic compartments as well as 123 124 ENMs hazard and systemic ecosystem responses to ENM contamination. Finally, the interest 125 of aquatic mesocosms in ENM lifecycle-oriented and ERA studies, as well as the current 126 limitations and knowledge gaps, will be discussed.
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| 135 | . A large panel of ENM environmental exposure scenario | s |
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(circles) the studies focused on water and soil compartments, respectively.

2.1 Diverse aquatic mesocosm designs

Figure 2 provides an overview of the main designs found in the literature, where aquatic mesocosm strategies are categorized based on their location and the ecosystem mimicked. A detailed list of these works, with additional information regarding the ENM properties, experimental setups and endpoints addressed is provided in the Supporting Information (Table S1). In general, a major distinction in experimental approaches is achieved between indoor laboratory-scale facilities and outdoor systems operated in the natural environment (ex situ or in situ), which have been equally reported in literature. A further distinction is obtained along a salinity gradient, i.e. from freshwater (84% of the literature) to estuarine and seawater (16% of the literature) ecosystems (Fig. 2).

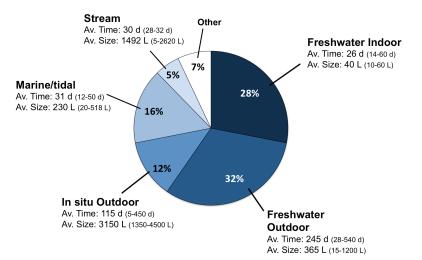


Figure 2. Mesocosm designs applied in the studies of ENM environmental fate with corresponding average experimental duration (Av. Time) and average water volume in the mesocosms (Av. Size). In parenthesis the range of minimum and maximum values applied for each design. "Other designs" refers to low-volume (<10 L) constructed wetland and sediment-oriented studies.

Freshwater indoor mesocosms have been implemented to mimic both lotic and lentic ecosystems. Relatively small aquaria (e.g. 10-15 L) were employed to study microbial communities²⁰ or specific trophic chains²¹, while larger modular set-ups (50-60 L tanks) were used to simulate complex ecological frameworks^{e.g.22,23,24}. These systems ensure the control of several parameters (e.g. pH, temperature, water volume, sediment depth) and allow high repeatability²⁵. Macro-organisms tested across these studies have included a variety of aquatic macrophytes and grazing animals as well as echinoderms, crustaceans, insects and amphibians larvae, and fishes. Authors used natural inoculums to colonize the mesocosms with primary producers sampled from the environment to be mimicked. Sediment matrices typically consisted of artificial sediments combined with sieved natural sediments. Similarly, aqueous phases consisted of commercial water with a pH and ionic strength close to the

actual environment spiked with water samples collected in the field²². This allowed controlling the water chemistry and porous media properties (texture, density, metals content) while providing natural inputs to the mesocosms. Indoor mesocosms have been extensively used to study the fate of a wide range of ENMs. The exposure to the ENMs usually took place over a period of several weeks (26 days on average) and was preceded by an acclimation phase (approx. two weeks) for the equilibration of the (eco)system (e.g. settling of suspended matter, pH/Eh stabilization, development of microbiotic communities).

Freshwater outdoor mesocosms achieve more realistic exposure scenarios at the expense of 178 179 less control and a lower feasibility compared to laboratory-scale set-ups. These experiments account for seasonal changes and varied weather conditions (e.g. precipitation, drought). 180 They are prone to colonization by organisms from the surrounding environment, resulting in 181 an enhanced complexity but diminished control of parameters such as water volume, 182 temperature and overall biodiversity. Small 10-20 L mesocosms were occasionally employed 183 outdoors to monitor algae population response over several months after ENMs 184 contamination^{26,27,28}. The most consistently applied outdoor design made use of larger 185 186 facilities (365 L on average) optimized for longer exposure testing (245 d on average)^{e.g.29,30}. While more challenging with regard to repeatability and the possibility for standardization, 187 these systems provide information about long-term contamination effects^{e.g.31} and ecosystem 188 response and recovery. For instance, freshwater, wetland ecosystems were simulated for 189 periods up to 1,5 years in rectangular-shaped containers (3.5 m³) lined with a uniform soil 190 191 layer (~20 cm) and a sloped-bed allowing humidity and redox gradients between permanently and periodically flooded areas²⁹. The watertight enclosures were open at the 192 top to allow variation of the water volume due to rainfall, evaporation and transpiration. 193 Native plankton and benthic bacteria communities were brought with natural groundwater 194 and soil matrices. Macro-organisms tested include terrestrial and aquatic macrophytes, 195 molluscs, crustaceans, and fishes as well as unintentional insects. 196

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Stream freshwater mesocosms allowed for the addition of ENMs to flowing water in order to monitor their mobility and transformation during transport. Outdoor studies employed hybrid flow-through/recirculating systems to model wastewater treatment plant and constructed wetland effluents^{32,33}. Briefly, these consisted of two collecting pools connected by 18 x 0.5 m sloped channels presenting riffle and glide sections. Engineered nanomaterialamended water in the top pool was gravity-fed to the bottom and then partially or totally

recirculated in order to maintain a stable volume and an essentially closed system. The 204 resulting mesocosms were colonized with insects, fishes, and snails and were tailored for 205 studying the effects of ENMs on the periphyton and algae populations. These systems 206 207 necessarily employ large water volumes and usually operate for short periods of time 208 (weeks), also to limit wastewater volume. Flow-through designs were also employed in an 209 indoor study, with small recirculating containers used for exposing snails and algae under 210 permanent water flowing conditions. Although less representative of whole stream 211 ecosystems, this approach presented several advantages such as the possibility to 212 accumulate large algae populations and to perform highly replicated experiments³⁴. 213

In situ mesocosms were obtained by contaminating an artificially bounded portion of a 214 natural lentic ecosystem. Furtado et al. applied such a strategy in two different studies where 215 round-shaped polypropylene sheeting, suspended from a floating ring and sealed to the 216 sediments, was directly placed in a sheltered bay of a littoral lake^{35,36}. These open-bottom 217 designs represent a straightforward approach for studying larger volumes (2400-3000 L) with 218 219 natural matrices. These studies were carried out over a relatively short-term of 30 d. Longer 220 exposure periods were achieved by Jovanovic et al., with a similar design where 1100 L 221 fiberglass tanks were attached to a platform anchored in the middle of a lake³⁷. Their mesocosms were filled with sediments and filtered lake water followed by introduction of 222 zooplankton, macroinvertebrates, macrophytes and fishes before exposure to ENMs for 223 224 three months. Larger in situ mesocosms (> 5000 L) were also used for the evaluation of community structures in contaminated ditches^{38,39,40}. These systems focused on sediments 225 and benthic community structure and recolonization of polluted areas following sediment 226 227 contamination by ENMs for periods of up to 15 months.

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229 Marine mesocosms have been less implemented compared to freshwater ones. Some pioneering works have been achieved in saltwater scenarios^{41,42,43}, which are to date the least 230 231 investigated compartments (approx. 15% of the reported literature). This can be ascribed to 232 the low predicted environmental concentrations (PEC) expected in marine environments due 233 to the ENM removal from the water phase at high salinities⁴⁴. In a few instances, indoor 234 facilities used small tanks (20 L) containing natural seawater and sediments for the study of native bacteria and plankton communities^{42,43}. More recently, larger systems (>300 L) were 235 applied to study low-dose addition of ENMs to open water⁴⁵ or coastal ecosystems 236 previously colonized with molluscs and echinoderms⁴⁶. Tidal cycles in estuarine ecosystems 237

have also been mimicked. Buffet et al. investigated the response of benthic worms and filterfeeders to ENM-contaminated intertidal sediments in outdoor facilities⁴⁷,⁴⁸. In their study,
pre-contaminated sediments in ~5L containers were introduced in a top-tank and cyclically
received aerated natural seawater from a reservoir. Similar designs (with 2 reservoirs to mimic
the tide) have also been used indoors in more complex ecological setups including plants,
snails, clams, shrimp and fish^{41,49,50}.

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245 2.2 ENM dosing scenarios and total doses

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247 When selecting the ENM to be investigated in aquatic mesocosms, a broader attention has been given to potentially hazardous materials, such as antimicrobial and (photo)catalytic Aq-, 248 Ti-, and Ce-ENMs (34%, 15%, and 16% of the aquatic mesocosms studies, respectively). Cu-, 249 Au- were also the subject of some studies, whereas little or no attention was paid to several 250 high production ENMs such as Zn-, Si-, Fe-, Al-ENMs and low production volume materials 251 such as quantum dots. For instance, Si-based ENMs were mostly neglected, despite SiO₂ 252 ENMs being one of the most produced^{51,52}. Similarly, C-based ENMs (e.g. fullerenes, 253 254 nanotubes, graphene) were investigated to a lower extent (10% of the studies) compared to metal-based ENMs. A graphene ENM was only recently included in a study despite its 255 increasing production and application rates⁵³. Carbon-based ENMs such as graphene and 256 CNTs can display enhanced toxicity when interacting with natural components^{54,55}, which 257 258 should be further characterized in realistic environmental settings. In general, the selection of ENMs should be based on criteria identifying the potentially most impactful materials in 259 environmental contexts. For instance, Arvidsson et al., identified "global production volume" 260 and "short-term ecotoxicity" as relevant proxy measures for assessing ENMs environmental 261 impact⁵¹. They further provided a proof of concept assessment classifying several ENMs on a 262 low-to-high production vs toxicity basis. Similar criteria can be used to identify the most 263 264 relevant ENM to be investigated in future aquatic mesocosms. 265 Whatever the ENMs, the absolute dose injected can strongly affect both their exposure and 266 hazards of the mimicked ecosystem, as many physico-chemical processes (aggregation, 267 dissolution, adsorption) are kinetically controlled by the concentration^{31,56}. With a few 268 exceptions, contamination of the mesocosms at low mg.L-1 levels has emerged as the best compromise between environmentally relevant concentrations and qualitative and 269

270 quantitative analyses in the complex matrices investigated. To date, half of the mesocosms

271 experiments used mg.L⁻¹ contamination levels, whereas less than 20% were carried out at the

more environmentally relevant low µg.L⁻¹ levels (Fig. 3). Although lower than the 272 273 concentrations typically applied in in vitro ecotoxicity studies, these ranges are higher than 274 the PECs, mostly expected in the range of ng.L⁻¹ and µg.Kg⁻¹ in water and sediments, 275 respectively⁴⁴. A well-known challenge in mesocosm studies performed at environmentally 276 relevant concentrations is the analytical feasibility of detecting and quantifying low 277 concentrations in matrices with high natural backgrounds (e.g. detecting Ti-ENMs within 278 sediment naturally rich in endogenous Ti). Recent studies confronted these issues by 279 applying powerful analytical methodologies using spICP-MS (for metal-based ENMs)^{35,45,57} or 280 optimized HPLC-MS (for carbon-based ENMs) able to detect pg.L⁻¹ levels⁴⁶. The use of 281 isotopically-labeled ENMs shows a great potential for detecting and discerning ENMs apart from the natural background at realistic contamination concentrations and can be applied to 282 a wide range of metal, metalloid or carbon-based ENMs. For instance, similar techniques 283 enabled the detection of ¹⁰⁹Ag-enriched Ag-ENMs⁵⁸ or multi-isotopically labelled 284 ¹¹¹Cd⁷⁷Se/⁶⁸ZnS QDs^{59,60} at concentrations as low as ng.L⁻¹ in complex matrices. Furthermore, 285 understanding the isotopic fingerprint of ENMs can help distinguish natural from 286 287 manufactured ENMs, and even the synthetic methods and manufacturers of engineered 288 ENMs⁶¹. Such methodologies have surprisingly received scarce attention in mesocosm investigations (e.g. ¹⁴¹Ce-enriched Ce-ENMs⁶²) and may represent a straightforward solution 289 for overcoming the current dosing limitations in mesocosm studies. 290 More than addressing the absolute doses, several studies assessed the role of the dosing 291 scenario in the global response of the ecosystems to ENM contamination^{e.g.22,31,33}. Two main 292 mesocosm dosing procedures were published *i.e.* (I) "pulse" treatments that mimic single 293 294 contamination events and (II) "press" treatments describing continuous addition of low 295 doses of ENMs over the duration of the experiment. These are also referred to as acute or chronic exposures, respectively. Although equally reported in literature (Fig. 3), in recent 296 297 years press contamination was increasingly applied in order to mimic the chronic exposures expected in aquatic ecosystems^{31,63}. The mesocosms dosing influenced both the ENMs fate 298 299 and effects, affecting partitioning between the environmental compartments and 300 bioaccumulation and biomagnification in organisms^{31,64}. Using multivariate analysis, Nassar et

al. recently highlighted that the dosing scenario constituted the main parameter to consider when investigating the environmental risk for a given ecosystem exposed to ENMs with

303 different sizes and coatings⁶⁵.

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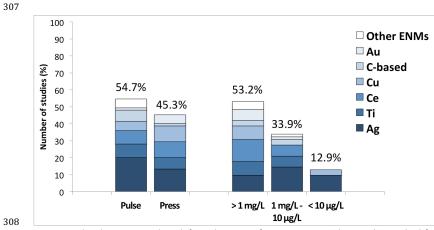


Figure 3. The dosing procedure (left) and ranges of concentrations (dose, right) applied for 309

aquatic mesocosm contamination. The dose concentration ranges refer to the total ENMs 310

input in the mesocosms at the end of the experiment. 311

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2.3 ENM partitioning within ecological compartments 313

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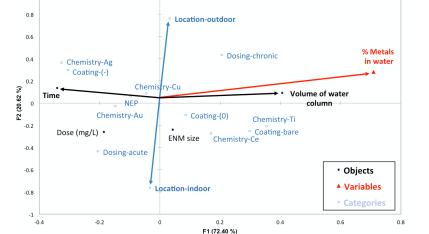
315 More than the absolute dose, the environmental risk posed by ENMs stems from their 316 accumulation and speciation within ecological compartments. ENM partitioning in 317 mesocosms is driven by bio-physico-chemical transformation processes leading to the distribution of large aggregates in the sediments and more well-dispersed ENMs or 318 319 dissolved metals in the water column. Espinasse et al., reported short aqueous residence 320 times in a wetland mesocosm for a range of ENMs (Aq > TiO_2 > SWCNT > CeO₂ ENMs) with half-lives in the water ranged from 36 h for CeO2-NPs to 10 d for Aq-NPs⁶⁶. Such an 321 322 environmental partitioning results from dynamic (bio)transformations in the ecosystem regulated by environmental factors such as the water chemistry, the ENM properties, the 323 exposure scenario and the mesocosm design. 324 To investigate the contribution of each of these parameters on the ENM partitioning, 325 multivariate analyses were performed on data extracted from the papers reviewed herein. 326 Among the fifty peer-reviewed papers initially screened, 31 that comprehensively reported 327 328 the concentrations of metals detected in the environmental compartments as well as data 329 about both the ENMs used (e.g. composition, size, coating) and experimental parameters applied (e.g. dose and dosing, mesocosm location, duration and total water volume of 330 mesocosm) were selected (Supporting Information, table S2). It must be noted that the 331 analysis did not include environmental factors that showed low variability between the 332 333 studies (e.g. pH, 8.2 ± 0.6 in average) or large daily or seasonal changes during the 334 experiments (e.g. pH, T, dissolved O_2). Parameters that were not comprehensively 335 investigated/reported across the studies under exam (e.g. organic and total suspended matter) were not included in this analysis. ENM partitioning was expressed as the percentage 336 of injected metals found in the water column at the end of the experiments (107 d on 337 338 average), which is an indicator of ENMs environmental distribution often reported in 339 literature. 340 PERMANOVA analysis (Supporting Information, Table S3) showed that at the end of the exposure mesocosm size (water volume), experiment duration and location had a major role

- 342 in defining the metal partitioning (p-value<0.05). In Canonical Correlation Analysis (Fig. 4), a
- 343 strong linear correlation was identified between the mesocosm water volume and the
- percentage of metals in the water ($R^2 = 0.163$, p-value ≤ 0.001). This is attributed to a higher 344
- water height in larger mesocosms, resulting in longer settling time and metal persistency in 345
- 346 water. Conversely, the experiment duration was inversely correlated to the percentage of

metals in water ($R^2 = 0.049$, p-value = 0.023), which is consistent with the kinetics of ENMs settling onto surficial sediments.

The mesocosm location (i.e. indoor or outdoor) had a significant impact on the system variance (PERMANOVA, $R^2 = 0.064$, p-value ≤ 0.005) but in Canonical Correlation Analysis this parameter was poorly correlated with the percentage of metals in the water. It is noteworthy that the long-term accumulation of the metals/ENMs in the sediments was observed independently of the location, suggesting that both indoor and outdoor designs were consistent in reproducing the behaviour of metal ENMs in aquatic ecosystems on the long-term.

356 It must be noted that, since the data used for this statistical analysis referred to the end of the experiments (107 d on average), the results presented here may underestimate the role 357 that parameters played over the short term. For instance, the intrinsic ENM physico-chemical 358 properties (size, chemistry, coating), the water chemistry (freshwater vs seawater), and the 359 dosing scenario had no major role in defining the metal partitioning at the end of the 360 361 experiment (p-value>0.05). For the dosing scenario, the Canonical Correlation Analysis 362 suggests that press dosing (chronic) was directly correlated with the percentage of metals in the water, whereas pulse dosing (acute) was inversely correlated (Fig. 4). This was consistent 363 with the observations reported in literature, i.e. that for a given ENM dose pulse additions 364 caused a short-term higher aqueous concentration but also rapid settling of the ENMs onto 365 the sediments, whereas press treatments lead to a higher persistence of ENMs in the water 366 367 column on the longer term. For instance, Colman et al. showed that, in wetland mesocosms, 368 water [Ag] in chronic exposures matched those of the acute treatment after three weeks, and were ~200-fold higher after 7 months at equal total doses³¹. In pond mesocosms, Ce-ENMs 369 sedimentation rates were not significantly different following pulse or press dosing, but 370 chronic addition resulted in a higher final Ce concentration in water column after 1 month²². 371 Comparable behaviors were also observed in stream³² or saltwater ecosystems^{41,43,49}. 372 373 Nonetheless, a limitation in the statistical analysis was due to the lack of data available about 374 important parameters driving ENM fate. For instance, while conductivity was often reported 375 as an estimate of mesocosm water ionic strength, the ionic species in solution were seldom 376 characterized. Similarly, while macrophytes and associated biofilm have been described as a 377 main factor for ENM fateeg.67,68, estimates of the biomass and surface area are missing. Statistical analysis such as the one presented here will benefit from more comprehensive 378 datasets including such data and future mesocosms studies are expected to fill these gaps. 379 380



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Fig. 4. Canonical Correlation Analysis showing the relationships between qualitative (blue) and quantitative (black) parameters and the amount of metals in the water column (%, red) at the end of the experiments. The arrows identify the significant parameters (p-value<0.05 in PERMANOVA) and the angle between the arrows indicates the degree of linear correlation between them.

- 389 390
- 391 3. Environmental risk for aquatic ecological niches
- 392 393
- 394 3.1 The benthic compartment: high exposure to less reactive ENMs
- 395

A predominant accumulation of the ENMs and metals onto the sediment was transversally reported across the studies, driven by homo- and/or hetero-aggregation with natural

colloidse.g.^{69,70} and interaction with various aquatic organisms⁶⁶. Thus, benthic organisms are predominantly exposed to aggregated ENMs, accumulated at the top sediment layer (0-1 cm)^{23,71,72}. Once in the sediments, ENMs can be remobilized via bioturbation, feeding, and entered the food web. Studies highlighted ENMs biomagnification and ENMs trophic transfers, e.g. from biofilm to insect larvae, molluscs and fish^{24,41,64}.

403 The risk posed to the benthic communities did not only depend on the relatively more important doses in sediments but also on the metals speciation. Studies described 404 405 (bio)transformation processes are relevant in such dynamic redox environment and after interaction with benthic biota. Oxidation/sulfidation is a major transformation pathway for 406 407 ENMs such as Aq, Zn and Cu in sub-oxic and anoxic environments^{29,56}. Lowry et al., showed that sulfidation of Aq-ENMs in freshwater sediment and organisms involves the oxidation of 408 Aq(0) to Aq(I) and subsequent formation of Aq₂S and sulfhydryl compounds²⁹. The 409 transformed Ag-ENMs remained bioavailable to benthic species^{29,31,66}. However, the 410 diminished solubility and reactivity of these by-products translated to slower metal 411 dissolution, lower uptake and weaker damage on aquatic plants, benthic microbial 412 community and (in)vertebrate compared to pristine ENMs^{73,74,75}. In the case of Cu-ENMs, 413 sulfidation could even enhance mobility and dissolution, contributing to a faster 414 transformation of the pristine ENMs via Cu-S- and Cu-O- binding with organic matter⁶⁷. 415

Grazing benthic macroorganisms such as snails and filter feeders have been shown to 416 accumulated Ag-, Ce-, Cu- and C-ENMs^{23,46,48,76} and had the highest concentration factor in 417 complex food webs exposed to Au-ENMs⁴¹. In estuarine ecosystems, evidence of DNA 418 419 damage and oxidative stress in clams exposed to Cu- and Ag-ENMs is attributed to the uptake of aggregated ENMs containing redox active metals⁴⁸. In freshwater ecosystems, 420 ENM contamination led to behavioural alterations of bivalve filtering capacity (Ce-ENM76) 421 and molluscs excretion rates (Au- and Cu-ENMs⁷⁷). Ce-ENMs also induced oxidative stress to 422 snails and genotoxic effects in amphibian larvae^{24,78}, linked to the uptake of aggregated 423 ENMs and the subsequent bioreduction of Ce(IV) in Ce(III)^{70,76}. In these studies, citrate coated 424 425 Ce-ENMs were less aggregated and more bioavailable⁷⁶, but caused lower oxidative stress,

426 likely because of the antioxidant properties of the organic ligands²³.

- 427 Macrophytes and benthic algae were found to be an important target for Ce-³², Ti-^{33,34}, Ag-³¹
- 428 and Cu-ENMs $^{67},^{79}\!\!$. Aquatic plants present a high surface area and could sequester a

429 significant fraction of ENMs in the long term, mostly due to adsorption and uptake onto

430 plants roots, tissues and associated microbiome $^{67,68,80}\!\!.$ These processes influence ENMs

431 $\,$ cycling by immobilizing the ENMs and reintroducing them in the food web^{29,66}. However,

most studies did not identify the main interaction mechanisms between ENMs and plant 432 tissues. Negative effects on periphyton endpoints (e.g. algal density, viability, diversity, 433 434 chlorophyll production, biomass growth) were also observed at relatively high doses (> 1 435 mg/L) of Ti-^{27,33}, Al-²⁶, and Fe-ENMs²⁸ in lotic and lentic ecosystems. 436 Biofilms associated with sediment and benthic organisms were also often identified as an important sink^{e.g.41,81} and can contribute to the immobilization of the ENMs as well as their 437 438 biotransformation^{68,82} and entry in the food web⁵⁰. However, the impact of ENMs on benthic 439 microbiota appeared to be limited, even in the case of the expectedly toxic Aq- and Cu-440 ENMs. Several studies found negligible effects on prokaryote abundance and diversity in 441 saltwater⁴³ and freshwater sediments⁸³. On a longer term, the stability of microbial community function and structure was associated with a developed tolerance to metals in 442 constructed wetlands⁸¹. Interestingly, other studies highlighted how resilience depends on 443 444 population density and how specific taxa were more sensitive to contamination⁸⁴.

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447 3.2 The planktonic compartment: low exposure to more reactive species

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Because ENMs primarily partition onto the sediments, most of the research focused on the
fate and effects in the benthic compartments and data available on planktonic communities
are limited. However, despite the presence of low ENM concentrations in the water column
over time, exposure of planktonic biota involved potentially more bioavailable, reactive and
consequently potentially toxic ENMs and metals.
Several works investigated the role of dissolution as a transformation pathway that might

release toxic by-products as free-metal ions. Avellan et al., recently showed the (bio)dissolution of Au-ENMs by bacteria within biofilms growing on macrophytes and the metals complexation in stable cyanide, hydroxyl or thiolic derivatives^{68,82}. A rapid dissolution

- 458 was also observed in freshwater for Cu nano-pesticides⁸⁵ and Al-ENMs²⁶ and it can be 459 expected for ENMs such as ZnO, whose toxicity has been related to ions rather than
- 460 nanoparticulate forms⁵⁶. In other studies, quantitative assessment of dissolution was difficult
- 461 to estimate due to re-complexation of the resulting ions with natural suspended matter.
- 462 Furtado et al., found relatively low dissolution rates of Ag-ENMs in freshwater lake
- 463 mesocosms at low ionic strength and high dissolved organic carbon (DOC), and concluded
- that free metals such as Ag^+ ions were removed by complexation with natural organic matter
- 465 $(NOM)^{35}$. Hence, while ENMs were observed to decrease in size, they may have formed

larger hetero-aggregates rich in metals. Other studies showed that plant derived DOC 466 stabilized the Aq- and Ce-ENMs in suspension and that the adsorption of organic ligands 467 blocked the oxidation sites and prevented dissolution^{86,87}. 468

469 Furthermore, decreasing the ENM size, due to the progressive desorption of metals, can 470 dramatically change their behavior, leading to enhanced mobility, reactivity, and toxicity. In 471 wetland mesocosms, smaller Ce-ENMs interacted more with suspended solids and were 472 significantly more bioavailable to (micro)organisms⁸⁸. Similarly, in a marine ecosystem, the 473 negative effects of Aq-ENMs to plankton communities were associated with diminishing sizes and dissolved silver ions in solution⁴⁵. Tsiola et al., showed that Aq-ENMs and the resulting 474 475 dissolved Ag ions significantly affected seawater planktonic microbial communities45. In particular, they observed noxious effects on specific species and highlighted the potential 476 damage to photosynthetic activity of cyanobacteria. In freshwater, Aq-ENMs affected the 477 abundance and had a hormesis effect on the planktonic microbial communities89. In the 478 water column, a bioaccumulation of Aq-ENMs was found in dragonfly larvae²⁹, whereas the 479 toxicity of Ce-ENMs was observed in amphibian larvae⁷⁸. A significant bioaccumulation of 480 481 Aq-ENMs and Ti-ENMs was also observed in higher organisms such as fish species^{21,29}.

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3.3 Global ecosystem response

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485 Mesocosm studies provide a global view of the ecosystem responses including toxicity arising from indirect interactions with ENMs as well as shifts in complex ecological processes 486 and ecosystem functioning^{30,78}. For instance, the noxious effects of Ce-ENMs toward 487 secondary consumers in pond mesocosms was not observed in parallel experiments with 488 direct or trophic chain exposures78. The authors attributed the toxicity to changes in the 489 microbiota community, affecting the nitrogen cycle and litter decomposition. Thus, the 490 491 ecosystem equilibria may be altered following disturbances to key ecological processes related to biota respiration or nutrient production and consumption. Toxicity to detritivores 492 493 such as chironomid larvae²⁴, combined with shifts in the structure of fungal and bacteria 494 communities^{90,91,92}, have also been shown to alter organic matter decomposition and the 495 availability of nutrients.

A decrease in chlorophyll production and algal biomass as well as variations of abiotic 496 parameters such as redox potential and dissolved oxygen were reported after exposure to 497

Fe-20,28, Al-26 and Ti-ENMs²⁷. Further studies suggested a limited impact on the long-term, 498

but effects of Ag- and Cu-ENMs on photosynthesis and methane metabolism⁸³. 499

In a few instances, the authors reported major ecosystem disruption and were able to 500 501 characterize a complex cascade of effects following contamination. A strong eutrophication 502 was observed in wetland mesocosms due to a synergistic effect of either Au- or Cu-ENMs 503 with nutrient enrichment, which led to an extended algal bloom (>50 d) in comparison with 504 nutrient only treatments⁹³. In this case, an increase in water turbidity eventually affected plant 505 growth and caused up to 92% reduction of the ecosystem primary production. Colman et al., related the toxicity of Aq-ENMs to macrophytes and phytoplankton with a general disruption 506 507 of the ecosystem resulting in increased methanogenesis³⁰. In this study, an initial damage to autotrophs led to the release of DOC and an increase of heterotrophic prokaryote 508 509 abundance and respiration. In turn, the consumption of dissolved O_2 and diminished photosynthetic activity resulted in hypoxia, which combined with elevated CO₂ and 510 decreased methane consumption led to a 40-fold increase of CH4 in the wetland. Although 511 such processes were observed after single pulse additions at relatively high doses of Ag-512 ENMs, a similar response of a lower magnitude was also identified in further experiments at 513 lower concentrations³¹. 514 515 Other studies reported minor or no changes in the ecosystem functioning, but identified 516 negative effects on specific ecological niches. In lake mesocosms, amendment of Ti-ENMs led to a significant decrease of soluble reactive phosphorus and although a wide range of 517 organisms (phytoplankton, plants and fishes) was unaffected³⁷. The authors reported a 518 significant decrease in the biomass of Rotifer species, which typically display high 519 520 reproductive rates and tolerance to environmental changes. Similarly, in two different 521 studies, the exposure to Aq-ENMs had no major consequences on most organisms under 522 investigation but affected benthic nitrifying bacteria, decreasing nitrification rates in the 523 ecosystem and altering the nitrogen cycling^{72,84}. Recently, Li et al., further investigated these processes, with a focus on the synergies between ENM and endogenous metal species⁹⁴. 524 525 526 4. Environmental perspectives and current knowledge gaps 527 528 4.1 Need for life-cycle oriented approaches

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ENMs release in the environment can occur at any stage of the value chain, i.e. from 530

synthesis to application/use and eventual disposal (Figure 5). The environmental research so 531

far has mainly focused on pristine ENMs (85% of mesocosm studies considered herein) rather 532

533 than nano-enabled products (NEPs). However, given the strict control at production and

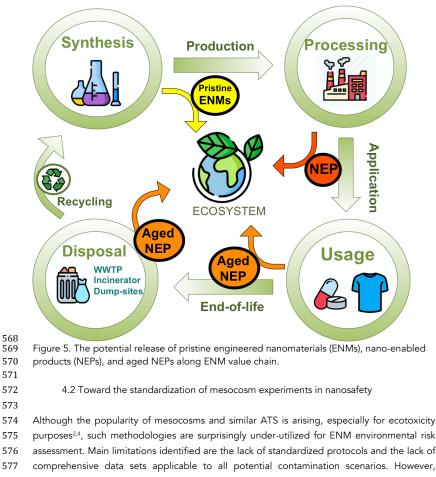
processing sites, incidental discharge into the environment may be more relevant during the 534

use and end-of-life phases. Environmental research on aged ENM by-products and NEPs is still very limited, especially in the case of mesocosms studies (11% and 4% of the studies considered herein, respectively).

538 Recent studies have stressed the need for life-cycle oriented approaches considering realistic exposure scenarios and the actual state of ENMs entering the environment^{6,95,96} and their 539 540 further aging in natural matrices^{3,97}. Aquatic mesocosms represent an invaluable tool for lifecycle oriented studies, as they implicitly account for the bio-physico-chemical aging of these 541 542 NEPs and aged ENMs and can highlight differences in environmental fate and impacts over long periods. For instance, an estuarine mesocosm experiment focusing on consumer 543 544 products containing Ag-ENMs, found a gradual release of up to 99% of the Ag ions over 60 days49. The Ag released by three different NEPs displayed higher persistence in the water 545 column compared to ionic Ag and pristine Ag-ENMs, and enhanced bioconcentration in 546 biofilm, snails and clams. On the contrary, Auffan et al., reported diminished aqueous 547 persistence of Cu-ENMs associated to fragmented paint NEPs⁹⁸. In their study, only 1% of 548 the Cu remained in the water column after 28 days, compared to 10% of the pristine ENMs. 549 550 In addition, the paint matrix prevented (bio)transformation of the ENMs, with regard to dissolution and complexation. More recently, Chatel et al., reported an enhanced 551 bioaccumulation of pristine Ti-ENMs compared to cement-embedded counterparts in 552 freshwater snails99. 553

These studies highlighted a significant difference between pristine and matrix-embedded 554 555 ENMs, suggesting that most results gathered to date should be considered with caution, as 556 they may not be representative of the actual behavior of ENMs entering the ecosystem. The characterization of pristine ENMs is still fundamental for risk assessment and management, 557 and is the basis for understanding transformed ENM properties⁵⁶. However, future research 558 should focus on NEPs especially in the context of safe(r)-by-design products¹⁰⁰. In particular, 559 priority should be given to reproducing aged NEPs from realistic sources and to the 560 application of relevant dosing for better mimicking real contamination scenarios. In this 561 562 framework, protocols/strategies have been developed for the production of suitable 563 materials, i.e. fragmented NEP and relative ENMs-free products to be used as controls^{95,96}. 564 Better use of these protocols should overcome current limitations in the understanding of 565 NEPs environmental release, exposure, risk, and life cycle assessment^{101,102}.

566 567



- 578 depending on their application, ENM contamination scenarios will likely concern specific
- 579 environmental compartments, whose assessment should be grouped and prioritized. For 580 instance, Ti-ENMs released from sunscreen consumer products accumulate in sediments in
- 581 near-shore freshwater and seawater environments and likely affect offshore environments to

a lower extent^{103,104,105}. Similarly, WWTP and constructed wetland effluents as well as river 582 sediments downstream of production sites were identified as hotspots of ENMs 583 contamination^{106,107}. In this context, mesocosms should be seen as tools for regional-scale 584 585 evaluations of such impacted areas, i.e. providing information about ENM fate and effects 586 under the most relevant conditions. While the current lack of standardization of mesocosms 587 translates to a lack of regulatory relevance, the same argument may be valid contrariwise, as 588 the lack of regulatory relevance currently hampers mesocosms use. Standardization may be 589 considered challenging, due to the overall complexity of mesocosms compared to other more classic approaches. However, it must be noted that specific designs, e.g. for mimicking 590 591 pond and wetland environments, have been consistently applied in the last decade with essentially identical procedures. Similarly, dual-tanks systems for mimicking the tidal cycle 592 have been applied and optimized to reproduce estuarine environments. These designs have 593 allowed for the study and comparison of several ENM classes in potentially affected 594 environmental compartments, and provide set up examples for future research 595 investigations. In general, no mesocosms design is a priori better than another, since the 596 597 design should be tailored to mimic the exposure scenario of interest. Nonetheless, the design must be comprehensive of the ecosystem setting and complexity, for instance with 598 regard to micro-biota and organisms, which should be representative of both the planktonic 599 and benthic compartments. Biota populations and mesocosm sizes should also be 600 harmonized, in order to avoid stress or behavioural changes e.g. due to population densities. 601 602 According with the studies reviewed here, experimental periods of several weeks or months 603 are recommended, in order to allow mid- to long-term observations of ENMs fate, ecosystem response and trajectory. Furthermore, an acclimation period is also 604 recommended prior contamination to better mimic the equilibrated ecosystem. Finally, 605 mesocosms are self-sufficient environments and once operating should not be extensively 606 manipulated except for sampling, e.g. with the addition of organisms, food sources or 607 nutrients. Future studies will need to further identify and characterize the most important 608 609 parameters influencing ENM exposure and hazard, which will in turn define the designs 610 applied to future experiments. 611

- 612
- 613 5. Conclusion
- 614

- Mesocosms are increasingly being used for the study of ENM environmental fate and effects, 615 616 and were reviewed here with a focus on aquatic ecosystems. When compared to most SOP, the advantage of mesocosms is the possibility to mimic realistic contamination scenarios, 617 618 comprehensive of natural complexity and synergies between contaminants and natural 619 components. Addressing multiple endpoints during dynamic ecological processes allows a 620 realistic, kinetic-oriented investigation of ENM exposure and hazards of target ecological 621 niches. Furthermore, mesocosms incorporate experimental periods of months to years, thus 622 allowing the long-term characterization of ENM contamination and ecosystem response and 623 recovery. 624 Different mesocosms designs were consistent in showing that ENMs entering the aquatic environment will be predominantly removed from the water column and accumulate onto 625 the sediments. Statistical analysis suggests that such an accumulation will occur in the long-626 term, independent of ENM physico-chemical properties. As a result, the potential risk for 627 aquatic benthos is defined by a higher accumulation of less reactive species (e.g. homo- / 628 hetero-aggregated, sulfidised). On the contrary, the risk for planktonic species will result 629 630 from a lower concentration of potentially more reactive ENM/metals. 631 Future studies will need to address current knowledge gaps, with special regard to the identification of the most environmentally relevant ENMs/NEPs and their realistic dosing. In 632 addition, the scientific outcome to date had a stronger focus on the characterization of the 633 ENM fate than ecological effects and ecosystem responses, which are crucial for risk 634 635 assessment. Large and heterogeneous datasets are produced within mesocosms that should be fully exploited. In this context, researchers should carefully define analytical and operative 636 637 strategies in advance, in order to generate findable, accessible, interoperable and reusable 638 data for the nanosafety community and stakeholders. 639 640
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660 References

- 661
- 662 (1)Hansen, S. F.; Jensen, K. A.; Baun, A. NanoRiskCat: A Conceptual Tool for Categorization
- 663 and Communication of Exposure Potentials and Hazards of Nanomaterials in Consumer Products.
- 664 I Nanopart Res 2014, 16 (1), 2195, https://doi.org/10.1007/s11051-013-2195-z.
- 665 (2) Hjorth, R.; Holden, P. A.; Hansen, S. F.; Colman, B. P.; Grieger, K.; Hendren, C. O. The Role of
- 666 Alternative Testing Strategies in Environmental Risk Assessment of Engineered Nanomaterials. 667
- Environ. Sci.: Nano 2017, 4 (2), 292-301. https://doi.org/10.1039/C6EN00443A.
- Mitrano, D. M.; Nowack, B. The Need for a Life-Cycle Based Aging Paradigm for 668 (3)
- 669 Nanomaterials: Importance of Real-World Test Systems to Identify Realistic Particle 670 Transformations. Nanotechnology 2017, 28 (7), 072001. https://doi.org/10.1088/1361-
- 6528/28/7/072001. 671
- 672 (4) Bour, A.; Mouchet, F.; Silvestre, J.; Gauthier, L.; Pinelli, E. Environmentally Relevant
- 673 Approaches to Assess Nanoparticles Ecotoxicity: A Review. Journal of Hazardous Materials 2015, 674
- 283, 764–777. https://doi.org/10.1016/j.jhazmat.2014.10.021. 675 (5) Auffan, M.: Masion, A.: Mounevrac, C.: de Garidel-Thoron, C.: Hendren, C. O.: Thierv, A.:
- 676 Santaella, C.; Giamberini, L.; Bottero, I.-Y.; Wiesner, M. R.; Rose, J. Contribution of Mesocosm
- Testing to a Single-Step and Exposure-Driven Environmental Risk Assessment of Engineered 677
- 678 Nanomaterials. NanoImpact 2019, 13, 66-69. https://doi.org/10.1016/j.impact.2018.12.005.
- Masion; Auffan; Rose. Monitoring the Environmental Aging of Nanomaterials: An 679 (6)
- 680 Opportunity for Mesocosm Testing? Materials 2019, 12 (15), 2447.
- 681 https://doi.org/10.3390/ma12152447.
- Desmau, M.; Carboni, A.; Le Bars, M.; Doelsch, E.; Benedetti, M. F.; Auffan, M.; Levard, C.; 682 (7)
- Gelabert, A. How Microbial Biofilms Control the Environmental Fate of Engineered 683
- 684 Nanoparticles? Front. Environ. Sci. 2020, 8, 82. https://doi.org/10.3389/fenvs.2020.00082.
- Odum, E. P. The Mesocosm. BioScience 1984, 9, 558-562. 685 (8)
- 686 https://doi.org/10.2307/1309598.
- 687 (9) Stewart, R. I. A.: Dossena, M.: Bohan, D. A.: Jeppesen, E.: Kordas, R. L.: Ledger, M. E.:
- Meerhoff, M.; Moss, B.; Mulder, C.; Shurin, J. B.; Suttle, B.; Thompson, R.; Trimmer, M.; Woodward, 688
- 689 G. Mesocosm Experiments as a Tool for Ecological Climate-Change Research. In Advances in
- 690 Ecological Research; Elsevier, 2013; Vol. 48, pp 71-181. https://doi.org/10.1016/B978-0-12-
- 691 417199-2.00002-1.

- 692 (10) Sharma, K. V.; Sarvalingam, B. K.; Marigoudar, S. R. A Review of Mesocosm Experiments on
- 693 Heavy Metals in Marine Environment and Related Issues of Emerging Concerns. Environ Sci Pollut
- 694 Res 2021, 28, 1304-1316.
- (11) Caquet, Th. Use of Aquatic Mesocosms in Ecotoxicology: State of the Art and Perspectives. 695
- Radioprotection 2002, 37 (C1), C1-173-C1-177. https://doi.org/10.1051/radiopro/2002033. 696
- 697 (12) Solomon, K. R.: Hanson, M. Mesocosms and Microcosms (Aquatic). In Encyclopedia of
- 698 Toxicology; Elsevier, 2014; pp 223-226. https://doi.org/10.1016/B978-0-12-386454-3.00568-6.
- (13) Boyle, T. P.; Fairchild, J. F. THE ROLE OF MESOCOSM STUDIES IN ECOLOGICAL RISK 699
- 700 ANALYSIS, Ecological Applications 1997, 7 (4), 1099–1102, https://doi.org/10.1890/1051-
- 701 0761(1997)007[1099:TROMSI]2.0.CO;2.
- 702 (14) Campbell, P. J.; Arnold, D. J. S.; Brock, T. C. M.; Grandy, N. J.; Heger, W.; Heimbach, F.;
- 703 Maund, S. J.: Streloke, M. Higher-Tier Aquatic Risk Assessment for Pesticides: Guidance Document
- 704 from the SETAC-Europe/OECD/EC Workshop. 1999.
- 705 (15) Holden, P. A.; Gardea-Torresdey, J. L.; Klaessig, F.; Turco, R. F.; Mortimer, M.; Hund-Rinke,
- 706 K.: Cohen Hubal, E. A.: Avery, D.: Barceló, D.: Behra, R.: Cohen, Y.: Devdier-Stephan, L.: Ferguson,
- 707 P. L.; Fernandes, T. F.; Herr Harthorn, B.; Henderson, W. M.; Hoke, R. A.; Hristozov, D.; Johnston, J.
- 708 M.; Kane, A. B.; Kapustka, L.; Keller, A. A.; Lenihan, H. S.; Lovell, W.; Murphy, C. J.; Nisbet, R. M.;
- 709 Petersen, E. J.; Salinas, E. R.; Scheringer, M.; Sharma, M.; Speed, D. E.; Sultan, Y.; Westerhoff, P.;
- 710 White, J. C.; Wiesner, M. R.; Wong, E. M.; Xing, B.; Steele Horan, M.; Godwin, H. A.; Nel, A. E.
- 711 Considerations of Environmentally Relevant Test Conditions for Improved Evaluation of
- 712 Ecological Hazards of Engineered Nanomaterials. Environ. Sci. Technol. 2016. 50 (12), 6124-
- 6145. https://doi.org/10.1021/acs.est.6b00608. 713
- (16) Kühnel, D.; Nickel, C.; Hellack, B.; van der Zalm, E.; Kussatz, C.; Herrchen, M.; Meisterjahn, 714
- 715 B.; Hund-Rinke, K. Closing Gaps for Environmental Risk Screening of Engineered Nanomaterials.
- 716 NanoImpact 2019, 15, 100173. https://doi.org/10.1016/j.impact.2019.100173.
- 717 (17) Avadi, A.: Rose, I.: de Garidel-Thoron, C.: Hendren, C.: Wiesner, M. R.: Auffan, M.
- 718 MESOCOSM: A Mesocosm Database Management System for Environmental Nanosafety. 2021.
- 719 21, 100288. https://doi.org/10.1016/j.impact.2020.100288.
- 720 (18) Yoon, S.-I.: An. Y.-I. Microcosm Studies of Nanomaterials in Water and Soil Ecosystems.
- 721 Journal of Korean Society of Environmental Engineers 2012, 34 (4), 288–294.
- 722 https://doi.org/10.4491/KSEE.2012.34.4.288.
- 723 (19) Kwak, I. L.: An. Y.-I. The Current State of the Art in Research on Engineered Nanomaterials
- 724 and Terrestrial Environments: Different-Scale Approaches. Environmental Research 2016, 151,
- 725 368-382. https://doi.org/10.1016/j.envres.2016.08.005.
- 726 (20) Barnes, R. J.; van der Gast, C. J.; Riba, O.; Lehtovirta, L. E.; Prosser, J. I.; Dobson, P. J.;
- 727 Thompson, I. P. The Impact of Zero-Valent Iron Nanoparticles on a River Water Bacterial
- 728 Community. Journal of Hazardous Materials 2010, 184 (1-3), 73-80.
- 729 https://doi.org/10.1016/i.jhazmat.2010.08.006.
- 730 (21) Asztemborska, M.; Jakubiak, M.; Stęborowski, R.; Chajduk, E.; Bystrzejewska-Piotrowska,
- 731 G. Titanium Dioxide Nanoparticle Circulation in an Aquatic Ecosystem. Water Air Soil Pollut 2018,
- 732 229 (6), 208. https://doi.org/10.1007/s11270-018-3852-8.
- 733 (22) Auffan, M.; Tella, M.; Santaella, C.; Brousset, L.; Paillès, C.; Barakat, M.; Espinasse, B.;
- 734 Artells, E.; Issartel, J.; Masion, A.; Rose, J.; Wiesner, M. R.; Achouak, W.; Thiéry, A.; Bottero, J.-Y. An
- 735 Adaptable Mesocosm Platform for Performing Integrated Assessments of Nanomaterial Risk in
- Complex Environmental Systems. Sci Rep 2014, 4 (1), 5608. https://doi.org/10.1038/srep05608. 736
- 737 (23) Tella, M.; Auffan, M.; Brousset, L.; Issartel, J.; Kieffer, I.; Pailles, C.; Morel, E.; Santaella, C.;

- Angeletti, B.; Artells, E.; Rose, J.; Thiery, A.; Bottero, J.-Y. Transfer, Transformation, and Impacts of 738
- 739 Ceria Nanomaterials in Aquatic Mesocosms Simulating a Pond Ecosystem. Environ. Sci. Technol.
- 740 2014, 48 (16), 9004-9013. https://doi.org/10.1021/es501641b.
- (24) Bour, A.: Mouchet, F.: Cadarsi, S.: Silvestre, I.: Chauvet, E.: Bonzom, I.-M.: Pagnout, C.: Clivot, 741
- H.; Gauthier, L.; Pinelli, E. Impact of CeO 2 Nanoparticles on the Functions of Freshwater 742
- 743 Ecosystems: A Microcosm Study. Environ. Sci.: Nano 2016. 3 (4), 830-838.
- https://doi.org/10.1039/C6EN00116E. 744
- 745 (25) Nassar, M.; Auffan, M.; Santaella, C.; Masion, A.; Rose, J. Robustness of Indoor Aquatic
- 746 Mesocosm Experimentations and Data Reusability to Assess the Environmental Risks of
- 747 Nanomaterials. Front. Environ. Sci. 2021. 9. 625201.
- https://doi.org/10.3389/fenvs.2021.625201. 748
- 749 (26) Pakrashi, S.: Dalai, S.: Ritika: Sneha, B.: Chandrasekaran, N.: Mukheriee, A. A Temporal
- 750 Study on Fate of Al2O3 Nanoparticles in a Fresh Water Microcosm at Environmentally Relevant
- 751 Low Concentrations. Ecotoxicology and Environmental Safety 2012, 84, 70–77.
- 752 https://doi.org/10.1016/i.ecoenv.2012.06.015.
- 753 (27) Kumar, D.; Rajeshwari, A.; Roy, R.; Pakrashi, S.; Iswarya, V.; Paul, I. E.; Mathur, A.;
- 754 Chandrasekaran, N.; Mukherjee, A. A Temporal Study on the Effects of TiO2 Nanoparticles in a
- Fresh Water Microcosm. Proc. Natl. Acad. Sci., India, Sect. B Biol. Sci. 2014, 86 (2), 415-420. 755
- 756 https://doi.org/10.1007/s40011-014-0462-0.
- 757 (28) Kumar, D.; Parashar, A.; Chandrasekaran, N.; Mukheriee, A. The Stability and Fate of
- 758 Synthesized Zero-Valent Iron Nanoparticles in Freshwater Microcosm System, 3 Biotech 2017, 7 759 (3), 227. https://doi.org/10.1007/s13205-017-0869-4.
- (29) Lowry, G. V.; Espinasse, B. P.; Badireddy, A. R.; Richardson, C. J.; Reinsch, B. C.; Bryant, L. D.; 760
- Bone, A. I.: Deonarine, A.: Chae, S.: Therezien, M.: Colman, B. P.: Hsu-Kim, H.: Bernhardt, E. S.: 761
- Matson, C. W.; Wiesner, M. R. Long-Term Transformation and Fate of Manufactured Ag 762
- 763 Nanoparticles in a Simulated Large Scale Freshwater Emergent Wetland, Environ, Sci. Technol.
- **2012**. 46 (13). 7027–7036. https://doi.org/10.1021/es204608d.
- 764
- 765 (30) Colman, B. P.; Espinasse, B.; Richardson, C. J.; Matson, C. W.; Lowry, G. V.; Hunt, D. E.;
- Wiesner, M. R.: Bernhardt, E. S. Emerging Contaminant or an Old Toxin in Disguise? Silver 766 Nanoparticle Impacts on Ecosystems. Environ. Sci. Technol. 2014, 48 (9), 5229-5236.
- 767
- https://doi.org/10.1021/es405454v. 768
- 769 (31) Colman, B. P.: Baker, L. F.: King, R. S.: Matson, C. W.: Unrine, I. M.: Marinakos, S. M.: Gorka,
- 770 D. E.; Bernhardt, E. S. Dosing, Not the Dose: Comparing Chronic and Pulsed Silver Nanoparticle
- 771 Exposures. Environ. Sci. Technol. 2018, 52 (17), 10048-10056.
- 772 https://doi.org/10.1021/acs.est.8b01700.
- 773 (32) Baker, L. F.; King, R. S.; Unrine, J. M.; Castellon, B. T.; Lowry, G. V.; Matson, C. W. Press or
- 774 Pulse Exposures Determine the Environmental Fate of Cerium Nanoparticles in Stream
- 775 Mesocosms. Environ Toxicol Chem 2016, 35 (5), 1213-1223, https://doi.org/10.1002/etc.3261.
- 776 (33) Wright, M. V.; Matson, C. W.; Baker, L. F.; Castellon, B. T.; Watkins, P. S.; King, R. S. Titanium
- Dioxide Nanoparticle Exposure Reduces Algal Biomass and Alters Algal Assemblage Composition 777
- 778 in Wastewater Effluent-Dominated Stream Mesocosms. Science of The Total Environment 2018,
- 626, 357-365. https://doi.org/10.1016/j.scitotenv.2018.01.050. 779
- 780 (34) Kulacki, K. I.: Cardinale, B. I.: Keller, A. A.: Bier, R.: Dickson, H. How Do Stream Organisms
- 781 Respond to, and Influence, the Concentration of Titanium Dioxide Nanoparticles? A Mesocosm
- Study with Algae and Herbivores. Environmental Toxicology and Chemistry 2012, 31 (10), 2414-782
- 2422. https://doi.org/10.1002/etc.1962. 783

- (35) Furtado, L. M.: Hogue, M. E.: Mitrano, D. M.: Ranville, I. F.: Cheever, B.: Frost, P. C.: 784
- 785 Xenopoulos, M. A.; Hintelmann, H.; Metcalfe, C. D. The Persistence and Transformation of Silver
- 786 Nanoparticles in Littoral Lake Mesocosms Monitored Using Various Analytical Techniques.
- 787 Environ. Chem. 2014, 11 (4), 419. https://doi.org/10.1071/EN14064.
- 788 (36) Furtado, L. M.; Norman, B. C.; Xenopoulos, M. A.; Frost, P. C.; Metcalfe, C. D.; Hintelmann, H.
- 789 Environmental Fate of Silver Nanoparticles in Boreal Lake Ecosystems. Environ. Sci. Technol.
- **2015**, 49 (14), 8441–8450. https://doi.org/10.1021/acs.est.5b01116. 790
- 791 (37) Jovanović, B.; Bezirci, G.; Cağan, A. S.; Coppens, J.; Levi, E. E.; Oluz, Z.; Tuncel, E.; Duran, H.;
- 792 Beklioğlu, M. Food Web Effects of Titanium Dioxide Nanoparticles in an Outdoor Freshwater
- 793 Mesocosm Experiment. Nanotoxicology 2016, 10 (7), 902-912.
- 794 https://doi.org/10.3109/17435390.2016.1140242.
- 795 (38) Velzeboer, I.: Kuprvianchyk, D.: Peeters, E. T. H. M.: Koelmans, A. A. Community Effects of
- 796 Carbon Nanotubes in Aquatic Sediments. Environment International 2011, 37 (6), 1126-1130.
- 797 https://doi.org/10.1016/j.envint.2011.01.016.
- 798 (39) Velzeboer, I.: Peeters, E. T. H. M.: Koelmans, A. A. Multiwalled Carbon Nanotubes at
- 799 Environmentally Relevant Concentrations Affect the Composition of Benthic Communities.
- 800 Environ. Sci. Technol. 2013, 47 (13), 7475-7482. https://doi.org/10.1021/es400777j.
- 801 (40) Iovanović, B.; Milošević, D.; Piperac, M. S.; Savić, A. In Situ Effects of Titanium Dioxide
- Nanoparticles on Community Structure of Freshwater Benthic Macroinvertebrates. 802
- 803 Environmental Pollution **2016**, 213, 278–282, https://doi.org/10.1016/i.envpol.2016.02.024.
- 804 (41) Ferry, J. L.: Craig, P.: Hexel, C.: Sisco, P.: Frey, R.: Pennington, P. L.: Fulton, M. H.: Scott, I. G.:
- Decho, A. W.; Kashiwada, S.; Murphy, C. J.; Shaw, T. J. Transfer of Gold Nanoparticles from the 805
- Water Column to the Estuarine Food Web. Nature Nanotech 2009, 4 (7), 441-444. 806
- 807 https://doi.org/10.1038/nnano.2009.157.
- (42) Mühling, M.; Bradford, A.; Readman, J. W.; Somerfield, P. I.; Handy, R. D. An Investigation 808
- 809 into the Effects of Silver Nanoparticles on Antibiotic Resistance of Naturally Occurring Bacteria in
- 810 an Estuarine Sediment, Marine Environmental Research 2009, 68 (5), 278–283.
- 811 https://doi.org/10.1016/j.marenvres.2009.07.001.
- (43) Bradford, A.; Handy, R. D.; Readman, J. W.; Atfield, A.; Mühling, M. Impact of Silver 812
- Nanoparticle Contamination on the Genetic Diversity of Natural Bacterial Assemblages in 813
- Estuarine Sediments. Environ. Sci. Technol. 2009, 43 (12), 4530-4536. 814
- 815 https://doi.org/10.1021/es9001949.
- 816 (44) Giese, B.; Klaessig, F.; Park, B.; Kaegi, R.; Steinfeldt, M.; Wigger, H.; von Gleich, A.;
- Gottschalk, F. Risks, Release and Concentrations of Engineered Nanomaterial in the Environment. 817
- 818 Sci Rep 2018, 8 (1), 1565. https://doi.org/10.1038/s41598-018-19275-4.
- (45) Tsiola, A.; Toncelli, C.; Fodelianakis, S.; Michoud, G.; Bucheli, T. D.; Gavriilidou, A.; Kagiorgi, 819
- 820 M.; Kalantzi, I.; Knauer, K.; Kotoulas, G.; Mylona, K.; Papadopoulou, E.; Psarra, S.; Santi, I.;
- 821 Tsapakis, M.: Daffonchio, D.: Pergantis, S. A.: Pitta, P. Low-Dose Addition of Silver Nanoparticles
- 822 Stresses Marine Plankton Communities. Environ. Sci.: Nano 2018, 5 (8), 1965-1980.
- https://doi.org/10.1039/C8EN00195B. 823
- 824 (46) Sanchís, I.; Llorca, M.; Olmos, M.; Schirinzi, G. F.; Bosch-Orea, C.; Abad, E.; Barceló, D.; Farré,
- M. Metabolic Responses of Mytilus Galloprovincialis to Fullerenes in Mesocosm Exposure 825
- 826 Experiments, Environ, Sci. Technol. 2018, 52 (3), 1002-1013.
- 827 https://doi.org/10.1021/acs.est.7b04089.
- (47) Buffet, P.-E.; Richard, M.; Caupos, F.; Vergnoux, A.; Perrein-Ettajani, H.; Luna-Acosta, A.; 828
- Akcha, F.; Amiard, J.-C.; Amiard-Triquet, C.; Guibbolini, M.; Risso-De Faverney, C.; Thomas-Guyon, 829

- 830 H.; Reip, P.; Dybowska, A.; Berhanu, D.; Valsami-Jones, E.; Mouneyrac, C. A Mesocosm Study of
- 831 Fate and Effects of CuO Nanoparticles on Endobenthic Species (Scrobicularia Plana, Hediste
- 832 Diversicolor). *Environ. Sci. Technol.* **2013**, 130110104824003.
- 833 https://doi.org/10.1021/es303513r.
- 834 (48) Buffet, P.-E.; Zalouk-Vergnoux, A.; Châtel, A.; Berthet, B.; Métais, I.; Perrein-Ettajani, H.;
- 835 Poirier, L.; Luna-Acosta, A.; Thomas-Guyon, H.; Risso-de Faverney, C.; Guibbolini, M.; Gilliland, D.;
- 836 Valsami-Jones, E.; Mouneyrac, C. A Marine Mesocosm Study on the Environmental Fate of Silver
- 837 Nanoparticles and Toxicity Effects on Two Endobenthic Species: The Ragworm Hediste
- B38 Diversicolor and the Bivalve Mollusc Scrobicularia Plana. *Science of The Total Environment* **2014**,
- 839 470-471, 1151-1159. https://doi.org/10.1016/j.scitotenv.2013.10.114.
- 840 (49) Cleveland, D.; Long, S. E.; Pennington, P. L.; Cooper, E.; Fulton, M. H.; Scott, G. I.; Brewer, T.;
- 841 Davis, J.; Petersen, E. J.; Wood, L. Pilot Estuarine Mesocosm Study on the Environmental Fate of
- 842 Silver Nanomaterials Leached from Consumer Products. *Science of The Total Environment* **2012**,
- 843 *421–422*, 267–272. https://doi.org/10.1016/j.scitotenv.2012.01.025.
- 844 (50) Yeo, M.-K.; Nam, D.-H. Influence of Different Types of Nanomaterials on Their
- 845 Bioaccumulation in a Paddy Microcosm: A Comparison of TiO2 Nanoparticles and Nanotubes.
- 846 *Environmental Pollution* **2013**, *178*, 166–172. https://doi.org/10.1016/j.envpol.2013.03.040.
- 847 (51) Arvidsson, R.; Baun, A.; Furberg, A.; Hansen, S. F.; Molander, S. Proxy Measures for
- 848 Simplified Environmental Assessment of Manufactured Nanomaterials. *Environ. Sci. Technol.*
- 849 **2018**, *52* (23), 13670–13680. https://doi.org/10.1021/acs.est.8b05405.
- 850 (52) Piccinno, F.; Gottschalk, F.; Seeger, S.; Nowack, B. Industrial Production Quantities and
- Uses of Ten Engineered Nanomaterials in Europe and the World. J Nanopart Res 2012, 14 (9),
 1109. https://doi.org/10.1007/s11051-012-1109-9.
- 853 (53) Evariste, L.: Mottier, A.: Lagier, L.: Cadarsi, S.: Barret, M.: Sarrieu, C.: Soula, B.: Mouchet, F.:
- 854 Flahaut, E.; Pinelli, E.; Gauthier, L. Assessment of Graphene Oxide Ecotoxicity at Several Trophic
- Levels Using Aquatic Microcosms. *Carbon* **2020**, *156*, 261–271.
- 856 https://doi.org/10.1016/j.carbon.2019.09.051.
- 857 (54) Zhao, J.; Ning, F.; Cao, X.; Yao, H.; Wang, Z.; Xing, B. Photo-Transformation of Graphene
- 858 Oxide in the Presence of Co-Existing Metal Ions Regulated Its Toxicity to Freshwater Algae. Water
- 859 Research **2020**, 176, 115735. https://doi.org/10.1016/j.watres.2020.115735.
- 860 (55) Ahamed, A.; Liang, L.; Lee, M. Y.; Bobacka, J.; Lisak, G. Too Small to Matter?
- 861 Physicochemical Transformation and Toxicity of Engineered NTiO2, NSiO2, NZnO, Carbon
- 862 Nanotubes, and NAg. Journal of Hazardous Materials **2021**, 404, 124107.
- 863 https://doi.org/10.1016/j.jhazmat.2020.124107.
- 864 (56) Lead, J. R.; Batley, G. E.; Alvarez, P. J. J.; Croteau, M.-N.; Handy, R. D.; McLaughlin, M. J.; Judy,
- 865 J. D.; Schirmer, K. Nanomaterials in the Environment: Behavior, Fate, Bioavailability, and Effects-
- An Updated Review: Nanomaterials in the Environment. *Environ Toxicol Chem* 2018, 37 (8),
 2029–2063. https://doi.org/10.1002/etc.4147.
- 868 (57) Tuoriniemi, J.; Jürgens, M. D.; Hassellöv, M.; Cornelis, G. Size Dependence of Silver
- 869 Nanoparticle Removal in a Wastewater Treatment Plant Mesocosm Measured by FAST Single
- 870 Particle ICP-MS. Environ. Sci.: Nano 2017, 4 (5), 1189–1197.
- 871 https://doi.org/10.1039/C6EN00650G.
- 872 (58) Croteau, M.-N.; Dybowska, A. D.; Luoma, S. N.; Misra, S. K.; Valsami-Jones, E. Isotopically
- 873 Modified Silver Nanoparticles to Assess Nanosilver Bioavailability and Toxicity at
- 874 Environmentally Relevant Exposures. *Environ. Chem.* **2014**, *11* (3), 247.
- 875 https://doi.org/10.1071/EN13141.

- 876 (59) Supiandi, N. I.; Charron, G.; Tharaud, M.; Cordier, L.; Guigner, J.-M.; Benedetti, M. F.; Sivrv.
- 877 Y. Isotopically Labeled Nanoparticles at Relevant Concentrations: How Low Can We Go? The Case
- 878 of CdSe/ZnS QDs in Surface Waters. *Environ. Sci. Technol.* **2019**, *53* (5), 2586–2594.
- 879 https://doi.org/10.1021/acs.est.8b04096.
- 880 (60) Supiandi, N. I.; Charron, G.; Tharaud, M.; Benedetti, M. F.; Sivry, Y. Tracing Multi-
- 881 Isotopically Labelled CdSe/ZnS Quantum Dots in Biological Media. *Sci Rep* **2020**, *10* (1), 2866.
- 882 https://doi.org/10.1038/s41598-020-59206-w.
- 883 (61) Yang, X.; Liu, X.; Zhang, A.; Lu, D.; Li, G.; Zhang, Q.; Liu, Q.; Jiang, G. Distinguishing the
- 884 Sources of Silica Nanoparticles by Dual Isotopic Fingerprinting and Machine Learning. *Nat*
- 885 *Commun* **2019**, *10* (1), 1620. https://doi.org/10.1038/s41467-019-09629-5.
- 886 (62) Zhang, P.; He, X.; Ma, Y.; Lu, K.; Zhao, Y.; Zhang, Z. Distribution and Bioavailability of Ceria
- 887 Nanoparticles in an Aquatic Ecosystem Model. *Chemosphere* **2012**, *89* (5), 530–535.
- 888 https://doi.org/10.1016/j.chemosphere.2012.05.044.
- 889 (63) Sun, T. Y.; Bornhöft, N. A.; Hungerbühler, K.; Nowack, B. Dynamic Probabilistic Modeling of
- 890 Environmental Emissions of Engineered Nanomaterials. Environ. Sci. Technol. 2016, 50 (9),
- 891 4701-4711. https://doi.org/10.1021/acs.est.5b05828.
- 892 (64) Kim, J. I.; Park, H.-G.; Chang, K.-H.; Nam, D. H.; Yeo, M.-K. Trophic Transfer of Nano-TiO2 in
- a Paddy Microcosm: A Comparison of Single-Dose versus Sequential Multi-Dose Exposures.
- 894 Environmental Pollution 2016, 212, 316–324. https://doi.org/10.1016/j.envpol.2016.01.076.
- 895 (65) Nassar, M.; Auffan, M.; Santaella, C.; Masion, A.; Rose, J. Multivariate Analysis of the
- 896 Exposure and Hazard of Ceria Nanomaterials in Indoor Aquatic Mesocosms. Environ. Sci.: Nano
- 897 **2020**, 7 (6), 1661–1669. https://doi.org/10.1039/C9EN01439J.
- 898 (66) Espinasse, B. P.; Geitner, N. K.; Schierz, A.; Therezien, M.; Richardson, C. J.; Lowry, G. V.;
- 899 Ferguson, L.; Wiesner, M. R. Comparative Persistence of Engineered Nanoparticles in a Complex
- 900 Aquatic Ecosystem. *Environ. Sci. Technol.* **2018**, *52* (7), 4072–4078.
- 901 https://doi.org/10.1021/acs.est.7b06142.
- 902 (67) Stegemeier, J. P.; Avellan, A.; Lowry, G. V. Effect of Initial Speciation of Copper- and Silver-
- 903 Based Nanoparticles on Their Long-Term Fate and Phytoavailability in Freshwater Wetland
- 904 Mesocosms. Environ. Sci. Technol. **2017**, 51 (21), 12114–12122.
- 905 https://doi.org/10.1021/acs.est.7b02972.
- 906 (68) Avellan, A.; Simonin, M.; McGivney, E.; Bossa, N.; Spielman-Sun, E.; Rocca, J. D.; Bernhardt,
- 907 E. S.; Geitner, N. K.; Unrine, J. M.; Wiesner, M. R.; Lowry, G. V. Gold Nanoparticle Biodissolution by
- 908 a Freshwater Macrophyte and Its Associated Microbiome. Nature Nanotech 2018, 13 (11), 1072-
- 909 1077. https://doi.org/10.1038/s41565-018-0231-y.
- 910 (69) Bour, A.; Mouchet, F.; Cadarsi, S.; Silvestre, J.; Baqué, D.; Gauthier, L.; Pinelli, E. CeO2
- 911 Nanoparticle Fate in Environmental Conditions and Toxicity on a Freshwater Predator Species: A
- 912 Microcosm Study. Environ Sci Pollut Res 2017, 24 (20), 17081–17089.
- 913 https://doi.org/10.1007/s11356-017-9346-1.
- 914 (70) Tella, M.; Auffan, M.; Brousset, L.; Morel, E.; Proux, O.; Chanéac, C.; Angeletti, B.; Pailles, C.;
- 915 Artells, E.; Santaella, C.; Rose, J.; Thiéry, A.; Bottero, J.-Y. Chronic Dosing of a Simulated Pond
- 916 Ecosystem in Indoor Aquatic Mesocosms: Fate and Transport of CeO ₂ Nanoparticles. *Environ*.
- 917 *Sci.: Nano* **2015**, *2* (6), 653–663. https://doi.org/10.1039/C5EN00092K.
- 918 (71) Schierz, A.; Espinasse, B.; Wiesner, M. R.; Bisesi, J. H.; Sabo-Attwood, T.; Ferguson, P. L. Fate
- 919 of Single Walled Carbon Nanotubes in Wetland Ecosystems. Environ. Sci.: Nano 2014, 1 (6), 574-
- 920 583. https://doi.org/10.1039/C4EN00063C.
- 921 (72) Jiang, H. S.; Yin, L.; Ren, N. N.; Xian, L.; Zhao, S.; Li, W.; Gontero, B. The Effect of Chronic

- 922 Silver Nanoparticles on Aquatic System in Microcosms. Environmental Pollution 2017, 223, 395-
- 923 402. https://doi.org/10.1016/j.envpol.2017.01.036.
- 924 (73) Levard, C.; Mitra, S.; Yang, T.; Jew, A. D.; Badireddy, A. R.; Lowry, G. V.; Brown, G. E. Effect of
- 925 Chloride on the Dissolution Rate of Silver Nanoparticles and Toxicity to E. Coli. Environ. Sci.
- 926 Technol. 2013, 47 (11), 5738–5745. https://doi.org/10.1021/es400396f.
- 927 (74) Yuan, L.; Richardson, C. J.; Ho, M.; Willis, C. W.; Colman, B. P.; Wiesner, M. R. Stress
- 928 Responses of Aquatic Plants to Silver Nanoparticles. Environ. Sci. Technol. 2018, 52 (5), 2558-
- 929 2565. https://doi.org/10.1021/acs.est.7b05837.
- 930 (75) Ward, C. S.; Pan, J.-F.; Colman, B. P.; Wang, Z.; Gwin, C. A.; Williams, T. C.; Ardis, A.; Gunsch,
- 931 C. K.; Hunt, D. E. Conserved Microbial Toxicity Responses for Acute and Chronic Silver
- Nanoparticle Treatments in Wetland Mesocosms. *Environ. Sci. Technol.* 2019, *53* (6), 3268–3276.
 https://doi.org/10.1021/acs.est.8b06654.
- 934 (76) Garaud, M.; Auffan, M.; Devin, S.; Felten, V.; Pagnout, C.; Pain-Devin, S.; Proux, O.; Rodius, F.;
- 935 Sohm, B.; Giamberini, L. Integrated Assessment of Ceria Nanoparticle Impacts on the Freshwater
- 936 Bivalve Dreissena Polymorpha. Nanotoxicology **2016**, 10 (7), 935–944.
- 937 https://doi.org/10.3109/17435390.2016.1146363.
- 938 (77) Perrotta, B. G.; Simonin, M.; Back, J. A.; Anderson, S. M.; Avellan, A.; Bergemann, C. M.;
- 939 Castellon, B. T.; Colman, B. P.; Lowry, G. V.; Matson, C. W.; Bernhardt, E. S.; King, R. S. Copper and
- 940 Gold Nanoparticles Increase Nutrient Excretion Rates of Primary Consumers. Environ. Sci.
- 941 *Technol.* **2020**, 54 (16), 10170–10180. https://doi.org/10.1021/acs.est.0c02197.
- 942 (78) Bour, A.; Mouchet, F.; Cadarsi, S.; Silvestre, J.; Verneuil, L.; Baqué, D.; Chauvet, E.; Bonzom,
- 943 J.-M.; Pagnout, C.; Clivot, H.; Fourquaux, I.; Tella, M.; Auffan, M.; Gauthier, L.; Pinelli, E. Toxicity of
- 944 CeO 2 Nanoparticles on a Freshwater Experimental Trophic Chain: A Study in Environmentally
- 945 Relevant Conditions through the Use of Mesocosms. *Nanotoxicology* **2015**, 1–11.
- 946 https://doi.org/10.3109/17435390.2015.1053422.
- 947 (79) Carley, L. N.; Panchagavi, R.; Song, X.; Davenport, S.; Bergemann, C. M.; McCumber, A. W.;
- 948 Gunsch, C. K.; Simonin, M. Long-Term Effects of Copper Nanopesticides on Soil and Sediment
- 949 Community Diversity in Two Outdoor Mesocosm Experiments. Environ. Sci. Technol. 2020, 54
- 950 (14), 8878–8889. https://doi.org/10.1021/acs.est.0c00510.
- 951 (80) Stegemeier, J. P.; Colman, B. P.; Schwab, F.; Wiesner, M. R.; Lowry, G. V. Uptake and
- 952 Distribution of Silver in the Aquatic Plant Landoltia Punctata (Duckweed) Exposed to Silver and
- 953 Silver Sulfide Nanoparticles. Environ. Sci. Technol. 2017, 51 (9), 4936–4943.
- 954 https://doi.org/10.1021/acs.est.6b06491.
- 955 (81) Button, M.; Auvinen, H.; Van Koetsem, F.; Hosseinkhani, B.; Rousseau, D.; Weber, K. P.; Du
- 956 Laing, G. Susceptibility of Constructed Wetland Microbial Communities to Silver Nanoparticles: A
- 957 Microcosm Study. *Ecological Engineering* **2016**, *97*, 476–485.
- 958 https://doi.org/10.1016/j.ecoleng.2016.10.033.
- 959 (82) Avellan, A.; Simonin, M.; Anderson, S. M.; Geitner, N. K.; Bossa, N.; Spielman-Sun, E.;
- 960 Bernhardt, E. S.; Castellon, B. T.; Colman, B. P.; Cooper, J. L.; Ho, M.; Hochella, M. F.; Hsu-Kim, H.;
- 961 Inoue, S.; King, R. S.; Laughton, S.; Matson, C. W.; Perrotta, B. G.; Richardson, C. J.; Unrine, J. M.;
- 962 Wiesner, M. R.; Lowry, G. V. Differential Reactivity of Copper- and Gold-Based Nanomaterials
- 963 Controls Their Seasonal Biogeochemical Cycling and Fate in a Freshwater Wetland Mesocosm.
- 964 Environ. Sci. Technol. 2020, 54 (3), 1533–1544. https://doi.org/10.1021/acs.est.9b05097.
- 965 (83) Moore, J. D.; Stegemeier, J. P.; Bibby, K.; Marinakos, S. M.; Lowry, G. V.; Gregory, K. B.
- 966 Impacts of Pristine and Transformed Ag and Cu Engineered Nanomaterials on Surficial Sediment
- 967 Microbial Communities Appear Short-Lived. *Environ. Sci. Technol.* **2016**, *50* (5), 2641–2651.

- 968 https://doi.org/10.1021/acs.est.5b05054.
- 969 (84) Welz, P. J.; Khan, N.; Prins, A. The Effect of Biogenic and Chemically Manufactured Silver
- 970 Nanoparticles on the Benthic Bacterial Communities in River Sediments. Science of The Total
- 971 Environment 2018, 644, 1380-1390. https://doi.org/10.1016/j.scitotenv.2018.06.283.
- 972 (85) Vencalek, B. E.; Laughton, S. N.; Spielman-Sun, E.; Rodrigues, S. M.; Unrine, J. M.; Lowry, G.
- 973 V.; Gregory, K. B. In Situ Measurement of CuO and Cu(OH) 2 Nanoparticle Dissolution Rates in
- 974 Quiescent Freshwater Mesocosms. Environ. Sci. Technol. Lett. 2016, 3 (10), 375–380.
- 975 https://doi.org/10.1021/acs.estlett.6b00252.
- 976 (86) Bone, A. J.; Colman, B. P.; Gondikas, A. P.; Newton, K. M.; Harrold, K. H.; Cory, R. M.; Unrine,
- 977 J. M.; Klaine, S. J.; Matson, C. W.; Di Giulio, R. T. Biotic and Abiotic Interactions in Aquatic
- 978 Microcosms Determine Fate and Toxicity of Ag Nanoparticles: Part 2–Toxicity and Ag Speciation.
- 979 Environ. Sci. Technol. 2012, 46 (13), 6925–6933. https://doi.org/10.1021/es204683m.
- 980 (87) Unrine, J. M.; Colman, B. P.; Bone, A. J.; Gondikas, A. P.; Matson, C. W. Biotic and Abiotic
- 981 Interactions in Aquatic Microcosms Determine Fate and Toxicity of Ag Nanoparticles. Part 1.
- 982 Aggregation and Dissolution. Environ. Sci. Technol. 2012, 46 (13), 6915–6924.
- 983 https://doi.org/10.1021/es204682q.
- 984 (88) Geitner, N. K.; Cooper, J. L.; Avellan, A.; Castellon, B. T.; Perrotta, B. G.; Bossa, N.; Simonin,
- 985 M.; Anderson, S. M.; Inoue, S.; Hochella, M. F.; Richardson, C. J.; Bernhardt, E. S.; Lowry, G. V.;
- 986 Ferguson, P. L.; Matson, C. W.; King, R. S.; Unrine, J. M.; Wiesner, M. R.; Hsu-Kim, H. Size-Based
- 987 Differential Transport, Uptake, and Mass Distribution of Ceria (CeO 2) Nanoparticles in Wetland
- 988 Mesocosms. Environ. Sci. Technol. 2018, 52 (17), 9768–9776.
- 989 https://doi.org/10.1021/acs.est.8b02040.
- 990 (89) Auffan, M.; Santaella, C.; Brousset, L.; Tella, M.; Morel, E.; Ortet, P.; Barakat, M.; Chaneac, C.;
- 991 Issartel, J.; Angeletti, B.; Levard, C.; Hazemann, J.-L.; Wiesner, M.; Rose, J.; Thiéry, A.; Bottero, J.-Y.
- 992 The Shape and Speciation of Ag Nanoparticles Drive Their Impacts on Organisms in a Lotic
- 993 Ecosystem. Environ. Sci.: Nano 2020, 7 (10), 3167-3177. https://doi.org/10.1039/D0EN00442A.
- 994 (90) Pradhan, A.; Sahadevan, S.; Pascoal, C.; Cassio, F. Can Metal Nanoparticles Be a Threat to
- 995 Microbial Decomposers of Plant Litter in Streams? *Microbial Ecology* **2011**, *62(1)*, 58–68.
- 996 https://doi.org/DOI: 10.1007/s00248-011-9861-4.
- 997 (91) Du, J.; Zhang, Y.; Yin, Y.; Zhang, J.; Ma, H.; Li, K.; Wan, N. Do Environmental Concentrations
- 998 of Zinc Oxide Nanoparticle Pose Ecotoxicological Risk to Aquatic Fungi Associated with Leaf
- 999 Litter Decomposition? Water Research 2020, 178, 115840.
- 1000 https://doi.org/10.1016/j.watres.2020.115840.
- 1001 (92) Du, J.; Zhang, Y.; Qv, M.; Yin, Y.; Zhang, W.; Zhang, J.; Zhang, H. Different Phototoxicities of
- 1002 ZnO Nanoparticle on Stream Functioning. *Sci. Total Environ.* **2020**, *10*, 725:138340.
- 1003 https://doi.org/doi: 10.1016/j.scitotenv.2020.138340.
- 1004 (93) Simonin, M.; Colman, B. P.; Anderson, S. M.; King, R. S.; Ruis, M. T.; Avellan, A.; Bergemann,
- 1005 C. M.; Perrotta, B. G.; Geitner, N. K.; Ho, M.; Barrera, B.; Unrine, J. M.; Lowry, G. V.; Richardson, C. J.;
- 1006 Wiesner, M. R.; Bernhardt, E. S. Engineered Nanoparticles Interact with Nutrients to Intensify
- 1007 Eutrophication in a Wetland Ecosystem Experiment. Ecol Appl 2018, 28 (6), 1435–1449.
- 1008 https://doi.org/10.1002/eap.1742.
- 1009 (94) Li, Y.; Zhao, R.; Wang, L.; Niu, L.; Wang, C.; Hu, J.; Wu, H.; Zhang, W.; Wang, P. Silver
- 1010 Nanoparticles and Fe(III) Co-Regulate Microbial Community and N2O Emission in River
- 1011 Sediments. Science of The Total Environment 2020, 706, 135712.
- 1012 https://doi.org/10.1016/j.scitotenv.2019.135712.
- 1013 (95) Amorim, M. J. B.; Lin, S.; Schlich, K.; Navas, J. M.; Brunelli, A.; Neubauer, N.; Vilsmeier, K.;

- 1014 Costa, A. L.; Gondikas, A.; Xia, T.; Galbis, L.; Badetti, E.; Marcomini, A.; Hristozov, D.; Kammer, F.
- 1015 von der; Hund-Rinke, K.; Scott-Fordsmand, J. J.; Nel, A.; Wohlleben, W. Environmental Impacts by
- 1016 Fragments Released from Nanoenabled Products: A Multiassay, Multimaterial Exploration by the
- 1017 SUN Approach. Environ. Sci. Technol. **2018**, 52 (3), 1514–1524.
- 1018 https://doi.org/10.1021/acs.est.7b04122.
- 1019 (96) Nowack, B.; Boldrin, A.; Caballero, A.; Hansen, S. F.; Gottschalk, F.; Heggelund, L.; Hennig,
- 1020 M.; Mackevica, A.; Maes, H.; Navratilova, J.; Neubauer, N.; Peters, R.; Rose, J.; Schäffer, A.; Scifo, L.;
- 1021 Leeuwen, S. van; von der Kammer, F.; Wohlleben, W.; Wyrwoll, A.; Hristozov, D. Meeting the
- 1022 Needs for Released Nanomaterials Required for Further Testing—The SUN Approach. *Environ.*
- 1023 Sci. Technol. **2016**, 50 (6), 2747–2753. https://doi.org/10.1021/acs.est.5b04472.
- 1024 (97) Lowry, G. V.; Gregory, K. B.; Apte, S. C.; Lead, J. R. Transformations of Nanomaterials in the 1025 Environment. *Environ. Sci. Technol.* **2012**, *46* (13), 6893–6899.
- 1026 https://doi.org/10.1021/es300839e.
- 1027 (98) Auffan, M.; Liu, W.; Brousset, L.; Scifo, L.; Pariat, A.; Sanles, M.; Chaurand, P.; Angeletti, B.;
- 1028 Thiéry, A.; Masion, A.; Rose, J. Environmental Exposure of a Simulated Pond Ecosystem to a CuO
- Nanoparticle-Based Wood Stain throughout Its Life Cycle. *Environ. Sci.: Nano* 2018, 5 (11), 2579–
 2589. https://doi.org/10.1039/C8EN00712H.
- 1031 (99) Châtel, A.; Auffan, M.; Perrein-Ettajani, H.; Brousset, L.; Métais, I.; Chaurand, P.; Mouloud,
- 1032 M.; Clavaguera, S.; Gandolfo, Y.; Bruneau, M.; Masion, A.; Thiéry, A.; Rose, J.; Mouneyrac, C. The
- $1033 \qquad \text{Necessity of Investigating a Freshwater-Marine Continuum Using a Mesocosm Approach in}$
- 1034 Nanosafety: The Case Study of TiO2 MNM-Based Photocatalytic Cement. *NanoImpact* 2020, 20,
 100254. https://doi.org/10.1016/j.impact.2020.100254.
- 1036 (100) Rose, J.; Auffan, M.; de Garidel-Thoron, C.; Artous, S.; Auplat, C.; Brochard, G.; Capron, I.;
- 1037 Carriere, M.; Cathala, B.; Charlet, L.; Clavaguera, S.; Heulin, T.; Labille, J.; Orsiere, T.; Peyron, S.;
- 1038 Rabilloud, T.; Santaella, C.; Truffier-Boutry, D.; Wortham, H.; Masion, A. The SERENADE Project; a
- $1039 \qquad \text{Step Forward in the Safe by Design Process of Nanomaterials: The Benefits of a Diverse and} \\$
- 1040 Interdisciplinary Approach. *Nano Today* **2021**, *37*, 101065.
- 1041 https://doi.org/10.1016/j.nantod.2020.101065.
- 1042 (101) Pourzahedi, L.; Vance, M.; Eckelman, M. J. Life Cycle Assessment and Release Studies for 15
- 1043 Nanosilver-Enabled Consumer Products: Investigating Hotspots and Patterns of Contribution.
- 1044 Environ. Sci. Technol. 2017, 51 (12), 7148–7158. https://doi.org/10.1021/acs.est.6b05923.
- 1045 (102) Moeta, P. J.; Wesley-Smith, J.; Maity, A.; Thwala, M. Nano-Enabled Products in South Africa
- 1046and the Assessment of Environmental Exposure Potential for Engineered Nanomaterials. SN Appl.1047Sci. 2019, 1 (6), 577. https://doi.org/10.1007/s42452-019-0584-3.
- 1048 (103) Reed, R.; Martin, D.; Bednar, A.; Montano, M.; Westerhoff, P.; Ranville, J. Multi- Day Diurnal
- 1049 Measurements of Ti-Containing Nanoparticle and Organic Sunscreen Chemical Release during
- 1050 Recreational Use of a Natural Surface Water. *Environ. Sci.: Nano* **2017**, *4*, 69–77.
- 1051 https://doi.org/10.1039/C6EN00283H.
- 1052 (104) Rand, L. N.; Bi, Y.; Poustie, A.; Bednar, A. J.; Hanigan, D. J.; Westerhoff, P.; Ranville, J. F.
- 1053 Quantifying Temporal and Geographic Variation in Sunscreen and Mineralogic Titanium-
- 1054 Containing Nanoparticles in Three Recreational Rivers. Science of The Total Environment **2020**, 1055 743 140045 https://doi.org/10.1006/j.j.j.j.j.good.usu.ic/
- 1055 743, 140845. https://doi.org/10.1016/j.scitotenv.2020.140845.
- 1056 (105) Labille, J.; Slomberg, D.; Catalano, R.; Robert, S.; Apers-Tremelo, M.-L.; Boudenne, J.-L.;
- 1057 Manasfi, T.; Radakovitch, O. Assessing UV Filter Inputs into Beach Waters during Recreational
- 1058 Activity: A Field Study of Three French Mediterranean Beaches from Consumer Survey to Water
- 1059 Analysis. *Science of The Total Environment* **2020**, *706*, 136010.

- 1060 https://doi.org/10.1016/j.scitotenv.2019.136010.
- 1061 (106) Slomberg, D. L.; Auffan, M.; Guéniche, N.; Angeletti, B.; Campos, A.; Borschneck, D.;
- 1062 Aguerre-Chariol, O.; Rose, J. Anthropogenic Release and Distribution of Titanium Dioxide
- 1063 Particles in a River Downstream of a Nanomaterial Manufacturer Industrial Site. Front. Environ.
- 1064 *Sci.* **2020**, *8*, 76. https://doi.org/10.3389/fenvs.2020.00076.
- 1065 (107) Markus, A. A.; Krystek, P.; Tromp, P. C.; Parsons, J. R.; Roex, E. W. M.; Voogt, P. de; Laane, R.
- 1066 W. P. M. Determination of Metal-Based Nanoparticles in the River Dommel in the Netherlands via
- 1067 Ultrafiltration, HR-ICP-MS and SEM. *Science of The Total Environment* **2018**, 631–632, 485–495.
- 1068 https://doi.org/10.1016/j.scitotenv.2018.03.007. 1069
- 1070