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1 **TITLE:**
2 Purification and Quality Control of Recombinant Septin Complexes for Cell-Free Reconstitution
3

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25 **SUMMARY:**

26 *In vitro* reconstitution of cytoskeletal proteins is a vital tool to understand the basic functional
27 properties of these proteins. The present paper describes a protocol to purify and assess the
28 quality of recombinant septin complexes, which play a central role in cell division and migration.
29

30 **ABSTRACT:**

31 Septins are a family of conserved eukaryotic GTP-binding proteins that can form cytoskeletal
32 filaments and higher-order structures from hetero-oligomeric complexes. They interact with
33 other cytoskeletal components and the cell membrane to participate in important cellular
34 functions such as migration and cell division. Due to the complexity of septins' many interactions,
35 the large number of septin genes (13 in humans), and the ability of septins to form hetero-
36 oligomeric complexes with different subunit compositions, cell-free reconstitution is a vital
37 strategy to understand the basics of septin biology. The present paper first describes a method
38 to purify recombinant septins in their hetero-oligomeric form using a two-step affinity
39 chromatography approach. Then, the process of quality control used to check for the purity and
40 integrity of the septin complexes is detailed. This process combines native and denaturing gel
41 electrophoresis, negative stain electron microscopy, and interferometric scattering microscopy.
42 Finally, a description of the process to check for the polymerization ability of septin complexes
43 using negative stain electron microscopy and fluorescent microscopy is given. This demonstrates

44 that it is possible to produce high-quality human septin hexamers and octamers containing
45 different isoforms of septin_9, as well as *Drosophila* septin hexamers.

46

47 **INTRODUCTION:**

48 The cytoskeleton has been classically described as a three-component system consisting of actin
49 filaments, microtubules, and intermediate filaments¹, but recently, septins have been
50 acknowledged as a fourth component of the cytoskeleton¹. Septins are a family of GTP-binding
51 proteins that are conserved in eukaryotes². Septins are involved in many cellular functions such
52 as cell division³, cell-cell adhesion⁴, cell motility⁵, morphogenesis⁶, cellular infection⁷, and the
53 establishment and maintenance of cell polarity⁸. Despite their important functions, how septins
54 are involved in such processes is poorly understood.

55

56 The septin family of proteins is subdivided into several subgroups (four or seven, depending on
57 the classification) based on protein sequence similarity². Members of different subfamilies can
58 form palindromic hetero-oligomeric complexes, which are the building blocks of filaments and
59 which, in turn, assemble into higher-order structures such as bundles, rings, and meshworks<sup>1,9–
60 12</sup>. Further molecular complexity arises from the presence of different splice variants, an example
61 being human SEPT9, where there is evidence for specific functions of different splice variants<sup>13–
62 15</sup>. Additionally, the length of the hetero-oligomers depends on species and cell type. For
63 instance, *Caenorhabditis elegans* septins form tetramers¹⁶, *Drosophila melanogaster* septins
64 form hexamers¹⁷ (**Figure 1A**), *Saccharomyces cerevisiae* septins form octamers¹⁸, and human
65 septins form both hexamers and octamers¹⁹ (**Figure 1A**). The ability of septin isoforms and splice
66 variants from the same subfamily to substitute each other in the complex and the (co-)existence
67 of differently sized hetero-oligomers have made it difficult to delineate the cellular functions of
68 different hetero-oligomeric complexes¹².

69

70 Another interesting ability of septins is their ability to interact with many binding partners in the
71 cell. Septins bind the plasma membrane and membranous organelles during interphase and cell
72 division^{20–22}. In dividing cells, septins cooperate with anillin^{23–25} and actin and myosin during
73 cytokinesis^{26,27}. At the late stages of cytokinesis, septins seem to regulate the endosomal sorting
74 complexes required for the transport (ESCRT) system for midbody abscission²⁸. Additionally,
75 there is also evidence of septin located on the actin cortex and actin stress fibers of cells in
76 interphase cells^{29–31}. In specific cell types, septins also bind and regulate the microtubule
77 cytoskeleton^{32,33}.

78

79 All of these features make septins a very interesting protein system to study, but also a
80 challenging one. The combination of the large number of septin subunits (13 genes in humans
81 without counting splice variants²) with the potential of septin subunits from the same subfamily
82 to substitute each other and form differently sized hetero-oligomers makes it difficult to draw a
83 conclusion on the cellular function of a specific septin by genetic manipulation. Furthermore, the
84 multiple interactions of septins make interpreting the effects of common research tools such as
85 drugs³⁴ directed at cytoskeletal or membrane components a hard task.

86

87 A way to overcome this situation is to complement research in cells with *in vitro* (cell-free)
88 reconstitution of septins. *In vitro* reconstitution allows for the isolation of a single type of septin
89 hetero-oligomers with a specific subunit composition and length^{18,35–37}. This complex can then
90 be studied in a controlled environment, either alone to discover the basic structural and
91 physicochemical properties of septins^{38–40}, or in combination with desired partners such as model
92 biomembranes^{11,41,42}, actin filaments^{10,27}, or microtubules^{32,36} to decipher the nature of their
93 interactions.

94
95 Therefore, a reliable method to purify different septin complexes efficiently is vital for septin
96 research. However, even using the same protocol, different purifications can give proteins with
97 different activity/functionality or even integrity. For commercially available proteins such as
98 enzymes, the functionality and enzymatic activity are carefully validated⁴³. Implementing careful
99 quality control for cytoskeletal or structural proteins such as septins can be challenging, but it is
100 essential to make experiments across labs comparable.

101
102 This paper describes a robust method to purify high-quality recombinant septins in their hetero-
103 oligomeric form based on the simultaneous expression of two vectors containing mono- or bi-
104 cistronic constructs (**Table 1**) in *Escherichia coli* cells. The method consists of a two-step affinity
105 chromatography approach to capture septin hetero-oligomers containing both a his₆-tagged
106 septin and a Strep-II-tagged septin (**Figure 1B,C**). This protocol, first described in Iv et al.¹⁰, has
107 been used to purify *Drosophila* septin hexamers^{11,27,35}, human septin hexamers¹⁰, and several
108 human septin octamers containing different native (isoform 1, 3, and 5)^{10,32} or mutated SEPT9
109 isoforms³². Furthermore, a description of a set of techniques to assess the quality of the purified
110 septins is given. First, the integrity and correct stoichiometry of the septin subunits is checked
111 using denaturing electrophoresis and transmission electron microscopy (TEM). Then, the
112 presence of hetero-oligomers of the correct molecular mass and the presence of monomers or
113 smaller oligomers indicative of complex instability are examined by native electrophoresis and
114 mass photometry via interferometric scattering microscopy (iSCAT). Finally, the last step consists
115 of the assessment of the polymerizing activity of the septins using fluorescence microscopy and
116 TEM.

117
118 [Place **Figure 1** here]

119
120 **PROTOCOL:**

121
122 **1. Purification of septin hetero-oligomers**

123
124 1.1. Co-transformation of bacterial cells with the expression vectors

125
126 1.1.1. Select a combination of one pnEA and one pnCS plasmid⁴⁴ that will be used for expression.
127 Choose the combination depending on the desired subunit composition of the septin hetero-
128 oligomer^{10,35} and whether or not fluorescent tagging is required.

129

130 NOTE: C-terminally tagged monomeric superfolder GFP (msfGFP)-tagged SEPT2 (for human
131 septins) or msfGFP- or monomeric enhanced GFP (mEGFP)-DSep2 (for *Drosophila* septins) is used
132 here (**Table 1**).

133

134 1.1.2. Pipette 1 μL of each plasmid ($\sim 1 \text{ ng}/\mu\text{L}$) into 100 μL of competent BL21 *Escherichia coli* cells
135 and incubate on ice for 20 min.

136

137 1.1.3. Place the cells in a water bath at 42 $^{\circ}\text{C}$ for 40 s and then immediately incubate them for 3
138 min on ice.

139

140 1.1.4. Add 0.9 mL of lysogeny broth (LB) medium to the cell suspension and let the cells grow for
141 1 h at 37 $^{\circ}\text{C}$. Plate 100 μL of cells on warm LB-agar plates containing 100 $\mu\text{g}/\text{mL}$ ampicillin and
142 100 $\mu\text{g}/\text{mL}$ spectinomycin and incubate overnight at 37 $^{\circ}\text{C}$.

143

144 1.2. Grow bacterial pre-culture

145

146 1.2.1. Fill a 250 mL Erlenmeyer flask with 100 mL of Terrific broth (TB) or LB medium containing
147 100 $\mu\text{g}/\text{mL}$ ampicillin and 100 $\mu\text{g}/\text{mL}$ spectinomycin.

148

149 1.2.2. Pick out a single colony from the LB-agar plate with a sterile inoculation loop and transfer
150 it to fresh media from step 1.2.1.

151

152 1.2.3. Incubate at 37 $^{\circ}\text{C}$ in a rotary shaker incubator, either overnight or for at least 6 h.

153

154 NOTE: From this culture, a glycerol stock can be prepared by mixing the bacterial suspension 1:1
155 with glycerol and stored at -80°C . This stock can be used in step 1.2.2. instead of a freshly
156 transformed colony.

157

158 1.3. Bacterial culture and protein expression induction

159

160 1.3.1. Transfer 100 mL of grown bacteria into 5 L of TB or LB containing 50 $\mu\text{g}/\text{mL}$ ampicillin and
161 50 $\mu\text{g}/\text{mL}$ spectinomycin.

162

163 1.3.2. Grow this culture at 37 $^{\circ}\text{C}$ in a shaker incubator until it reaches an optical density (OD)
164 measured at a wavelength of 600 nm in the range of 2–3 for unlabeled septins or 0.6–0.8 for
165 msfGFP/mEGFP-labelled septins and induce protein expression by adding a final concentration
166 of 0.5 mM IPTG. The lower OD for the labeled septins is to avoid reaching the death phase in their
167 longer expression time, as detailed in the next step.

168

169 1.3.3. Incubate the cells expressing unlabeled septin hetero-oligomers for 3 h at 37 $^{\circ}\text{C}$ or the cells
170 expressing msfGFP-labelled hetero-oligomers overnight at 17 $^{\circ}\text{C}$.

171

172 NOTE: The short protein expression time for unlabeled complexes, facilitated by the use of the
173 richer TB medium, is chosen to prevent protein degradation. The longer expression time

174 combined with lower temperature for labeled complexes is chosen to allow for correct folding of
175 the msfGFP tag.

176

177 1.4. Bacterial lysis and lysate clarification

178

179 NOTE: From this point onwards in the purification procedure, keep the protein-containing
180 solution on ice or at 4 °C at all times to prevent proteolytic protein degradation or loss of activity.

181

182 1.4.1. Collect the cultured cells by centrifuging at 4,000 x *g* for 20 min at 4 °C. Discard the
183 supernatant.

184

185 1.4.1.1. Optionally, snap-freeze the pellet in this step and store at –80 °C for around 6 months. If
186 this option is chosen, make sure to thaw the pellet on ice before continuing.

187

188 1.4.2. Dissolve the pellet in 100 mL of lysis buffer (**Table 2**) and lyse the cells. Choose one of the
189 two options below

190

191 1.4.2.1. Sonicate in 7 cycles of 30 s ON and 59 s OFF with a tip sonicator using 30% amplitude
192 (note that the settings are sonicator-dependent).

193

194 1.4.2.2. Break down the cells in the French press by passing them at least 3x.

195

196 1.4.3. Clarify the cell lysate by centrifuging at 20,000 x *g* for 30 min at 4 °C and keep the
197 supernatant. It is recommended to start with step 1.5.1. during this centrifugal step.

198

199 1.4.4. Optionally, take a sample for denaturing electrophoresis, as described in section 2.

200

201 1.5. Affinity chromatography for His-tagged proteins

202

203 NOTE: This step yields complexes containing human SEPT2 or *Drosophila* Sep1 using a nickel
204 column (**Figure 1B**).

205

206 1.5.1. Equilibrate a pre-packed nickel sepharose high-performance chromatography column with
207 septin buffer (**Table 2**).

208

209 1.5.2. Load the clarified supernatant onto the column at 1 mL/min and wash the bound protein
210 with at least three column volumes of septin buffer.

211

212 1.5.3. Elute the septin complexes with 50% HisTrap elution buffer (**Table 2**) at 1 mL/min while
213 collecting 0.5 mL fractions to yield an imidazole concentration of 250 mM.

214

215 1.5.4. Pick the fractions containing septin complexes, as indicated by the optical absorbance of
216 the eluate at 280 nm monitored online with a fast protein liquid chromatography (FPLC) system
217 or after the purification with a microvolume spectrophotometer.

218

219 NOTE: Imidazole absorbs light at 280 nm. This probably explains why the protein peak does not
220 go back to zero absorbance after the septin elution (**Figure 2A**).

221

222 1.6. Affinity chromatography for Strep-II-tagged proteins

223

224 NOTE: This step yields complexes containing either human SEPT7 (hexamers), human SEPT9
225 (octamers), or *Drosophila* peanut using a Strep-Tactin column (**Figure 1B**). The chromatography
226 column is based on a modified biotin-streptavidin system. The protein is tagged with modified
227 biotin (Strep-II-tag), and the column contains an engineered streptavidin (Strep-Tactin). Despite
228 being modified from the biotin-streptavidin system, there is no interference between the Strep-
229 Tactin-Strep-II-tag system and the biotin-streptavidin system. The described system is used to
230 avoid interference with reconstitution assays using biotin and streptavidin.

231

232 1.6.1. Equilibrate a pre-packed StrepTactin sepharose high-performance chromatography
233 column with septin buffer (**Table 2**). Load the septin-containing fractions recovered from the
234 nickel column at 1 mL/min and wash the bound protein with at least three column volumes of
235 septin buffer.

236

237 1.6.2. Elute the septin complexes with 100% StrepTrap elution buffer (**Table 2**) at 1 mL/min while
238 collecting 0.5 mL fractions to yield a concentration of 2.5mM desthiobiotin.

239

240 NOTE: The desthiobiotin in the StrepTrap elution buffer must be dissolved fresh.

241

242 1.6.3. Pick the fractions containing septin complexes, as indicated by the optical absorbance of
243 the eluate at 280 nm monitored online with an FPLC system or after the purification with a
244 microvolume spectrophotometer.

245

246 NOTE: The denaturing electrophoresis is usually done at this point with samples of the column
247 washings and septin fractions. The order of columns can be inverted with indistinguishable
248 results, i.e., the clarified lysate after step 1.4. can be subjected to Strep-Tactin affinity
249 chromatography followed by nickel affinity chromatography.

250

251 1.7. Dialysis and storage

252

253 1.7.1. To remove the desthiobiotin from the final storage solution, dialyze the septin complexes
254 in a ~1:300 sample-to-buffer volume ratio against septin buffer (**Table 2**) supplemented with 1
255 mM DTT overnight, or for at least 4 h, at 4 °C using a 30 kDa MWCO dialysis membrane.

256

257 1.7.2. Optionally, concentrate the septins using a 30 kDa MWCO centrifugal concentration
258 column up to the desired concentration. Aim for a concentration of 5–7 µM, as measured via the
259 solution optical absorbance at 280 nm and using a theoretical extinction coefficient calculated
260 via ProtParam (**Table 3**).

261

262 1.7.3. Aliquot the protein complexes into the desired aliquot size, snap-freeze the aliquot, and
263 store it at -80°C .

264
265 NOTE: It is recommended not to store the protein for more than 6 months. In addition, it is
266 recommended to perform regular quality control experiments, especially if the protein is stored
267 for longer than the recommended time.

268 **2. Quality control of purity and integrity of the septin hetero-oligomer**

270
271 NOTE: The hetero-oligomer quality control consists of a set of biochemical and imaging
272 techniques that allow for the detection of the mass and integrity of the septin complexes present
273 in the solution.

274
275 2.1. Denaturing electrophoresis to check for the formation of the septin hetero-oligomer with
276 the correct components

277
278 2.1.1. Mix $10\ \mu\text{L}$ of the selected fractions with $10\ \mu\text{L}$ of 2x SDS sample buffer, load them onto a
279 precast 4%–15% TGX gel, and fill the system with Tris/glycine/SDS running buffer.

280
281 2.1.2. Run the electrophoresis for 35 min at 200 V and stain the gel (**Table of Materials**) to
282 visualize the results. The molecular weights of the individual septin proteins and septin hetero-
283 oligomeric complexes can be found in **Table 3**.

284
285 2.1.3. Measure the relative intensity of each band inside each lane containing purified septins in
286 a contrast-inverted image. Do this by calculating the mean intensity of equally sized rectangles
287 around each band and of an equally sized rectangle on a region without any band in the same
288 lane. Then, normalize the values by dividing the intensity of each band by the intensity of the
289 region without bands.

290
291 NOTE: If the intensity is saturated (for example, values of 255 for an 8-bit image on a contrast-
292 inverted image), skip the lane.

293
294 2.2. Ensemble-averaged native size distribution *via* native electrophoresis

295
296 2.2.1. Prepare 800 mL of anode buffer and 200 mL of light blue cathode buffer the day before
297 and store them in the fridge. To prepare the anode buffer, dilute 40 mL of 20x running buffer
298 with 760 mL of type-I deionized water (I-water). To prepare the light blue cathode buffer, dilute
299 10 mL of 20x running buffer and 1 mL of 20x cathode additive with 189 mL of I-water. The running
300 buffer and cathode additive come with a kit (**Table of Materials**).

301
302 2.2.2. Prepare $10\ \mu\text{L}$ of the sample by mixing $\sim 500\ \text{ng}$ of septin with the needed amount of sample
303 buffer ($2.5\ \mu\text{L}$ in this case, due to the use of a 4x sample buffer; see **Table of Materials**) and
304 enough I-water to reach a volume of $10\ \mu\text{L}$.

305

306 2.2.3. Load the samples onto the gel and fill the system with the ice-cold anode and cathode
307 buffers.

308
309 2.2.4. Run the electrophoresis for around 115 min at 150 V with a power supply that does not
310 stop at low currents and stain the gel (**Table of Materials**) to visualize the results. The molecular
311 weights of the single proteins and complexes calculated based on the sequence can be found in
312 **Table 3**.

313
314 2.3. Single-molecule mass distribution using mass photometry via interferometric scattering
315 microscopy

316
317 2.3.1. Wash #1.5 glass slides by sonicating them in an ultrasonic cleaner for 5 min in I-water, 5
318 min in isopropanol, and finally, 5 min in I-water.

319
320 2.3.2. Dry two glass slides with a gentle stream of nitrogen gas and place a 7 μL drop of 0.01%
321 Poly-L-Lysine (PLL) solution on the center of one of the slides. Then, place the center of the other
322 slide on top of the PLL drop, orienting the two slides orthogonally for easy separation. Incubate
323 for 30 s.

324
325 2.3.2.1. Wash by immersing in a beaker with I-water 1x and by directly applying a stream of I-
326 water 2x. Then, dry the two slides with a flow of nitrogen gas. These slides can be stored
327 afterward for around 6 weeks at room temperature in dry conditions.

328
329 NOTE: Label the side of the slide that is treated with the PLL-PEG to correctly run the experiment.

330
331 2.3.3. Just before the experiment, cut a piece of 2 x 2, 3 x 2, or 3 x 3 gaskets (that yield 4, 6, or 9
332 imaging chambers/slide, respectively) and stick it on the PLL-PEG-treated part of a glass slide
333 while avoiding the glass slide and the gaskets contacting any dirty surface. Place the slide on a
334 light-duty wiper tissue and press on the gaskets with a pipette tip to stick them with the
335 protecting plastic still on the gaskets.

336
337 2.3.4. Warm septin buffer (**Table 2**) to room temperature and thaw the proteins in hand (keep
338 them on ice afterward).

339
340 NOTE: iSCAT shows the signal of some detergents and small molecules that resemble protein
341 signals⁴⁵. DTT is one of those small molecules, and that is why it is not used for this experiment.
342 There is only a trace of DTT coming from the stored septin.

343
344 2.3.5. Place the slide with gaskets on the commercial mass photometry system containing 19 μL
345 of septin buffer and focus the microscope using the autofocus option. Follow the manufacturer's
346 instructions to check if the found focus is correct.

347 2.3.5.1. Optimize the frame rate and the exposure time by pressing on **Optimize illumination** and
348 refocus with the autofocus option with the new settings. The standard 100x objective that is part
349 of the setup is used here.

350
351 2.3.6. Create or load a project folder to store the data using **File > New Project** or **File > Load**
352 **Project**.

353
354 2.3.7. Pipette 1 μL of sample onto the 19 μL of septin buffer drop (step 2.3.5) used to focus and
355 mix while minimizing the movement of the slide by not touching anything while doing so. Then,
356 record a 6,000-frame video by clicking on **Record**.

357
358 2.3.7.1. For correct analysis, record the following samples: septin buffer, protein mass standard
359 for the calibration of the signal-to-mass ratio (if a recent calibration is available and the
360 environmental conditions have not changed, this sample can be skipped), and 250 nM of septin
361 complexes diluted in septin buffer without DTT (this gives a final concentration of ~ 12.5 nM).

362
363 2.3.8. Analyze the videos using the manufacturer's software to obtain the protein mass
364 distribution. Check for good quality data as follows.

365
366 2.3.8.1. If the peaks of different septin hetero-oligomers sizes are overlapping too much or too
367 many events are detected ($>3,500$ events for a 6,000-frames video with the regular field of view
368 of 128 pixels x 34 pixels spanning $10.8 \mu\text{m}$ x $2.9 \mu\text{m}$), decrease the final septin concentration and
369 measure again.

370
371 2.3.8.2. If there are not enough counts of single molecules measured (at least 2,500–3,500 for a
372 6,000-frame video with the regular field of view), increase the septin concentration and measure
373 again.

374
375 2.4. Direct imaging of septin complexes via negative stain transmission electron microscopy
376

377 2.4.1. Dilute samples to a concentration of about 50 nM in septin buffer and prepare the staining
378 solution (2% uranyl formate or uranyl acetate in I-water).

379
380 NOTE: Uranyl formate must be prepared fresh.

381
382 2.4.2. Pipette 4 μL of diluted septins onto a glow discharged electron microscopy grid and
383 incubate for 30 s.

384
385 2.4.3. Remove most of the protein solution using a filter paper and wash the grid 2x with septin
386 buffer and 1x with I-water to remove loosely adsorbed septins.

387
388 2.4.4. Stain with 2% uranyl acetate or uranyl formate solution in I-water for 1 min, absorb the
389 staining solution with a filter paper, and air-dry the grid for a few minutes.

390
391 2.4.5. Screen the grid using a properly aligned transmission electron microscope to search for
392 regions of enhanced stain and collect about 100 images within these selected areas.

393

394 2.4.6. Collect images at a magnification of at least 50,000x to obtain a pixel size of about 2 Å/pixel
395 and with a defocus varying from $-1\ \mu\text{m}$ to $-2\ \mu\text{m}$. Use an acceleration voltage of 200 kV.
396 Preferably, use an automated procedure to collect the data, which will depend on the acquisition
397 software available.

398

399 2.4.7. Perform 2D image processing using dedicated software

400

401 2.4.7.1. Box out at least 2,000 particles using a dedicated software⁴⁶.

402

403 2.4.7.2. Perform two-dimensional alignment and classification iteratively until classes are
404 obtained without further improvement. The first alignment and classification step should be
405 reference-free to avoid any bias in the classification.

406

407 2.4.7.3. Use the averages obtained from the first reference-free classification as new references
408 to carry out an extra round of classification. Repeat this process iteratively until no further
409 improvement is achieved. Ensure that each class is based on 50 to 100 picked particles, and
410 individual subunits are clearly visible. Different software tools can be used (Spider, Eman, or
411 Relion)⁴⁶⁻⁴⁸.

412

413 **3. Septin functional quality control via polymerization analysis**

414

415 NOTE: The functionality quality control consists of a set of imaging techniques that allow for the
416 detection of polymerized septin complexes. Below, unlabelled septins are referred to as “dark”
417 septins, and the buffer used for polymerizing unlabelled septins is referred to as “dark” septin
418 polymerization buffer (SPB).

419

420 3.1. Septin bundle imaging via fluorescence microscopy

421

422 3.1.1. Prepare the 5x fluoSPB (**Table 2**) and a septin mix consisting of 90% dark septin and 10%
423 msfGFP-septin at six times higher concentration than the desired final concentration in septin
424 buffer + 1 mM DTT. A typical concentration for this assay is 300 nM, and, therefore, the
425 concentration is 1,800 nM for this mix.

426

427 3.1.2. Polymerize the septin by mixing, in this specific order, I-water (enough to top up to the
428 final desired volume), 20% 5xfluoSPB (a final dilution of 1:5), 0.05 μM PCD, and 16.67% septin
429 mix (a final dilution of 1:6). For 10 μL , mix 6.23 μL of I-water, 2 μL of 5xfluoSPB, 0.1 μL of PCD
430 (with a stock of 5 μM), and 1.67 μL of septin mix. Incubate this mix for at least 30 min at room
431 temperature.

432

433 3.1.3. Add the samples to an imaging chamber washed with fluoSPB (**Table 2**) and image the
434 septin bundles. PLL-PEG-passivated flow channels, as described in previous research^{10,32}, work
435 well for this experiment.

436

437 **3.2. Septin bundle imaging via negative stain transmission electron microscopy**

438
439 3.2.1. Prepare the 5x darkSPB (**Table 2**) and a septin mix consisting of 100% dark septin at six
440 times higher concentration than the desired final concentration in septin buffer + 1 mM DTT. A
441 typical concentration for this assay is 300 nM, and, therefore, the concentration is 1,800 nM for
442 this mix.

443
444 3.2.2. Polymerize the septin by mixing, in this specific order, I-water (enough to top up to the
445 final desired volume), 20% 5xdarkSPB, and 16.67% septin mix. For 5 μ L, mix 3.16 μ L of I-water, 1
446 μ L of 5xfluSPB, and 0.83 μ L of septin mix. Incubate this mix for at least 30 min at room
447 temperature.

448
449 3.2.3. Add 3–5 μ L of sample to a glow-discharged electron microscopy grid and incubate for 1
450 min. Then, wash the grid 2x with darkSPB (**Table 2**) by absorbing the liquid with a filter paper and
451 adding a drop of darkSPB buffer, wash 1x with I-water, incubate for \sim 30 s with 2% uranyl acetate,
452 blot the stain, and air-dry the sample for a few minutes.

453
454 3.2.4. Image the septin bundles at 120 kV and magnifications between 5,000x and 60,000x with
455 a defocus of between 1–2 μ m.

456
457 **REPRESENTATIVE RESULTS:**

458 As mentioned in the protocol, 5 L of *E. coli* cells co-transformed with the two septin-expressing
459 plasmids were grown, and the expression of septins was induced by adding IPTG. After 3 h, the
460 cells were collected by centrifugation, resuspended in lysis buffer, and lysed by sonication. The
461 lysate was then clarified by centrifugation, and the clarified solution was applied to a HisTrap
462 column (**Figure 2A**). After the first purification, the septin-containing fractions were pooled and
463 applied onto a StrepTrap column (**Figure 2B**). This typically yields around 3–5 mL of \sim 1 μ M septin
464 complex. Before pooling the septin-containing fractions, denaturing gel electrophoresis can be
465 used to check for the integrity of the septin subunits and the equimolar stoichiometric ratio
466 between the different septin subunits forming the complex. (**Figure 3A**). If the gel shows similarly
467 intense bands corresponding to the molecular weights (**Table 3**) of the septin subunits, the
468 protocol can be continued. If not, it is recommended to start the protocol again. In the example
469 shown for human septin octamer with SEPT9_i1, **Figure 3A** clearly shows bands corresponding
470 to SEPT9_i1, SEPT6, SEPT7, and SEPT2 (in the order from top to bottom) with similar intensities;
471 the 99% confidence interval of the normalized intensity was 1.128 ± 0.048 for SEPT2, $1.092 \pm$
472 0.034 for SEPT6, 1.108 ± 0.040 for SEPT7, and 1.067 ± 0.029 for SEPT9. If SEPT2 is tagged with
473 msfGFP, it will shift up very close below SEPT9_i1. Depending on the electrophoresis system used
474 and the presence of the C-terminal TEV-Strep tag for SEPT7 (which makes it migrate more slowly
475 than untagged SEPT7), the SEPT7 and SEPT6 bands sometimes merge due to their comparable
476 molecular weights. The next step is to pool the fractions and dialyze them against septin buffer
477 with DTT. After the dialysis, if the concentration is too low ($<2 \mu$ M) or a higher concentration is
478 needed for the experiments, a concentration step can be included, as described in the protocol.
479 Concentrations below 1 μ M usually indicate the bad functional quality of the septins. A final
480 septin complex concentration between 3.5 μ M and 7 μ M works well for most *in vitro* assays.

481 These concentrations are usually obtained when the volume after concentration reaches 0.5–1
482 mL.

483
484 [Place **Figure 2** here]

485
486 To continue with the quality control, native electrophoresis, as described in the protocol, was
487 performed (**Figure 3B**). In the gels, a major band corresponding to the intact hetero-oligomers
488 and, usually, a minor band corresponding to trimers or tetramers can be observed. Human
489 hexamers are found a little bit above the 242 kDa marker band, while octamers are found above
490 the 480 kDa band, above their calculated molecular mass. The location of these bands was
491 checked by western blot analysis of eukaryotic cell extracts³². Tagging with msfGFP couples each
492 SEPT2 with an msfGFP protein. This causes an increase in the molecular weight of septin
493 complexes of 53.4 kDa (26.7 kDa/msGFP molecule). Nevertheless, on the native electrophoresis
494 gel, the apparent molecular weight of the msfGFP-tagged complexes is indistinguishable from
495 that of the untagged complexes.

496
497 A complementary technique to test whether the septin complexes are intact is mass photometry
498 by iSCAT microscopy. iSCAT monitors the light scattering of molecules landing on a glass slide
499 amplified by interference with reference light, typically the reflection of the laser on the bottom
500 of the glass slide. Then, a background subtraction approach is used to give contrast to the
501 particles. Due to this correction, the signal shows positive and negative values depending on
502 whether the particles land on the glass or move away from it⁴⁹. The detected signal is directly
503 proportional to the molecular weight of proteins⁵⁰. Therefore, a signal-to-mass calibration with
504 a mass standard can determine the mass of the sample proteins. **Figure 3C** shows an example of
505 human septin octamers containing SEPT9_i1. Most of the detected single particles (~50%) are of
506 a molecular weight expected for complete octamers containing SEPT9_i1 (423 kDa) (**Figure 3C**).
507 There are also particles with masses between 150–300 kDa, but no clear peak is observed,
508 indicating the possible presence of other septin species in low abundance. Similarly, most of the
509 detected single particles for mEGFP-tagged *Drosophila* hexamers are of a molecular weight
510 expected for intact hexamers (361 kDa) (**Figure 3D**). An additional clear peak at 241 kDa indicates
511 the presence of stable tetramers containing two peanut proteins, one DSep1 and one mEGFP-
512 DSep2. Finally, both the human and the fly septin complexes show a peak around 80 kDa that
513 could be a mix of monomers and dimers, possibly amplified by a trace of DTT or any other small
514 molecule that aggregates, showing a peak in the positive side of the plot⁴⁵.

515
516 [Place **Figure 3** here]

517
518 Given that both native gels and iSCAT provide ensemble-averaged information only, class
519 averaging of transmission electron microscopy images of single septin oligomers was used to
520 check the integrity and purity of the complexes by direct visualization. In TEM images of septin
521 complexes in septin buffer, rods of 24 nm (hexamers) or 32 nm (octamers) in length can be
522 observed. An example of a human septin octamer containing SEPT9_i1 can be seen in **Figure 3E**.
523 When class averaging them, each of the subunits can be observed and counted, as seen for the
524 msfGFP-tagged human octamer with SEPT9_i1 in **Figure 3F**. In case the oligomer is fluorescently

525 labeled, extra densities that correspond to the SEPT2-msfGFP can be observed at the end of the
526 rods (**Figure 3F**).

527

528 The combination of the above techniques proves that octamers (or hexamers) with the correct
529 stoichiometric ratio and high purity can be purified using the described protocol. Finally, the last
530 quality control check is for the functionality of the septin complexes in terms of their
531 polymerization ability. In the presence of low salt concentration (<150 mM KCl with the described
532 buffer⁹), if septins are not in the presence of other proteins or negatively charged lipid
533 membranes, they self-assemble into bundles⁹. Septins are prevented from polymerization by
534 keeping them in the storage buffer, which has a high (300 mM) KCl concentration. The septin
535 hetero-oligomers are then diluted at a 1:6 volume ratio in a buffer of the same composition but
536 without KCl to achieve a final KCl concentration of 50 mM. To do fluorescence imaging, this buffer
537 is complemented with an oxygen scavenging system to protect from photobleaching and with a
538 blinking suppressor. In TIRF microscopy, small clusters of proteins can be observed within the
539 shallow TIRF field (~100 nm; **Figure 4A,B**). On a confocal microscope, large clusters of filamentous
540 structures can be seen floating higher up in solution (**Figure 4C**). Finally, with TEM, small bundles
541 of septin (**Figure 4D**), corresponding to the clusters observed by TIRF, and large bundles (**Figure**
542 **4E**), corresponding to the structures observed by confocal microscopy, can be observed. The
543 insets of **Figure 4D,E** reveal that both types of structures consist of long, thin filaments that run
544 in parallel, forming bundles with tapered ends. Together, the fluorescence and TEM images prove
545 that the purified septin complexes can polymerize into filaments, which in turn self-assemble into
546 bundles.

547

548 [Place **Figure 4** here]

549

550 **FIGURE AND TABLE LEGENDS:**

551 **Figure 1: Purification strategy.** (A) Schematics of the septin hetero-oligomers that exist in human
552 (left) and *Drosophila* (right) cells. Numbers denote septin subunits from the indicated groups,
553 and P denotes peanut. Human SEPT9 can be any of its isoforms. The septin subunits have an
554 asymmetric shape and are longitudinally associated with two distinct interfaces, the NC:NC and
555 the G:G interface, as denoted by NC and G, respectively, on top of the human hexamer. (B,C)
556 Schematic illustration of the two-step chromatography strategy, shown for (B) human septin
557 hexamers and (C) octamers. H indicates the his-tags, while S indicates the Strep-II-tags.

558

559 **Figure 2: Example chromatograms corresponding to the purification of dark human septin**
560 **octamers_9i1.** (A) HisTrap column chromatogram. After the septin elution peak, the absorbance
561 does not go back to zero, likely due to the presence of imidazole in the buffer. The pooled fraction
562 went from the start of the elution peak until the absorbance stabilized at around 250 mL. (B)
563 StrepTrap column chromatogram. The pooled fraction went from the start of the elution peak
564 until the absorbance went back to around 0 at 50 mL.

565

566 **Figure 3: Examples of results of the oligomer quality control.** (A) Example of denaturing gel
567 showing different fractions of the elution peak from the purification of dark human septin
568 octamers_9i1. (B) Example of native electrophoresis of different septin complexes. (C,D)

569 Different examples of histogram results of mass photometry at 12.5 nM of septin complexes: (C)
570 dark human septin octamers_9i1 and (D) DSep1-msfGFP *Drosophila* septin hexamers. Lines are
571 Gaussian fits. (E) TEM image of 25 nM dark human septin octamers_9i1 in septin buffer. Scale
572 bar = 200 nm. (F) Class average image of SEPT2-msfGFP human septin octamers_9i1. The msfGFP
573 tags are visible as fuzzy densities on the two ends. Scale bar = 10 nm. Panels (E) and (F) are the
574 copyright of The Company of Biologists and have been adapted from Iv et al.¹⁰ with permission.
575

576 **Figure 4: Examples of results of the polymerization ability quality control.** (A) TIRF image of 300
577 nM human septin hexamers (10% msfGFP-labelled hexamers) in fluoSPB. (B) TIRF image of 300
578 nM human septin octamers containing SEPT9_i1 (10% msfGFP-labelled octamers9_i1) in fluoSPB.
579 (C) Confocal maximum-intensity projections of Z-stacks across ~30 μm with 0.5 μm of spacing of
580 300 nM human septin octamers_9i3 in fluoSPB. (A–C) Scale bar = 10 μm and inverted grayscale.
581 (D,E) Example TEM images of (D) small and (E) large bundles of human septin octamers_9i1 in
582 darkSPB. Insets show regions where clear filaments running parallel within the bundle can be
583 observed. Scale bars = 500 nm. Panels (C–E) are copyright of The Company of Biologists and have
584 been adapted from Iv et al.¹⁰ with permission.
585

586 **Table 1: List of plasmids.** Plasmids to purify septin oligomers following this protocol. All plasmids
587 have been deposited in Addgene (first column).
588

589 **Table 2: List of buffers.** Buffer compositions used for the purification and quality control of septin
590 oligomers.
591

592 **Table 3: Molecular weights and extinction coefficients.** List of molecular weights (MW) and
593 optical extinction coefficients (ϵ) at a wavelength of 280 nm calculated with ProtParam based on
594 the sequences of the complex, assuming linear fusion of the septin subunits, the different septin
595 complexes, and the unique septin subunits (only MW) that can be purified with the plasmids
596 listed in **Table 1**.
597

598 **DISCUSSION:**

599 The method described here allows for the robust purification and quality control of pre-formed
600 septin hetero-oligomers. Some of the key issues to consider for the correct application of the
601 method are as follows. During the elution steps in the chromatographic separations, it is
602 important to use the recommended (or lower) flow rate to minimize the dilution of the septin
603 complexes. Additionally, to maximize the recovery of protein during the final concentration step,
604 the concentrator column is oriented in such a way that the solution is not pushed against the
605 filter (when there is only a filter on one side). If the solution goes directly to the filter, the protein
606 sticks much more to it, substantially decreasing the final yield. It is also important to consider
607 that the concentration step is not always necessary. Picking fractions only from a narrow range
608 around the peak in the chromatogram usually gives a high enough stock concentration (>3,000
609 nM) for many reconstitution applications (which usually operate between 10–300 nM). Finally,
610 for the quality control of the functionality of the septin complexes by fluorescence microscopy,
611 it is important to correctly passivate the surface of the microscopy slides, since septin complexes

612 avidly stick to glass. Passivation of the glass slides can be done either via PLL-PEG
613 functionalization or by the formation of neutral (100% DOPC) supported lipid bilayers^{11,32}.

614

615 Compared to the original purification protocol first described in Iv et al.¹⁰, there is a change in
616 the buffer compositions (**Table 2**). The concentration of MgCl₂ has been reduced from 5 mM to
617 2 mM, and the concentration and pH of Tris-HCl have been reduced from 50 mM to 20 mM and
618 from 8.0 to 7.4, respectively. These changes were made to make the buffer conditions compatible
619 with studies of the interactions of human septins with lipid bilayers, actin filaments, and
620 microtubules^{10,11,32}. This is because the authors formed supported lipid bilayers and polymerized
621 actin in the F-buffer, whose composition is identical to that of darkSPB, apart from the presence
622 of ATP in the F-buffer. The buffer change did not produce any changes in the quality or lifetime
623 of the purified septins compared to the original buffers.

624

625 This method of purification still has several limitations. First, different purification attempts can
626 vary in yield (0.5–1 mL of 2–5 μM septin complexes) and functional quality, as checked by the
627 bundle formation ability of the purified septin complexes. That is why it is very important to
628 consistently perform the quality checks described in this paper. Controlling very well the times
629 of expression and the optical density of the bacterial culture can help mitigate the difference in
630 yield. Second, this purification pipeline cannot distinguish between trimers and hexamers or
631 between tetramers and octamers (**Figure 1B**). However, the quality control experiments can be
632 used to prove that the majority of septin complexes are in their long oligomer form. In case an
633 even narrow oligomer size distribution is required, size exclusion chromatography can be
634 inserted in between step 1.6. and step 1.7. of the purification protocol. This optional step,
635 however, dramatically decreases the yield, and it is not recommended unless it is strictly needed.
636 A last, more fundamental, limitation comes from the use of *E. coli* as an expression system for
637 recombinant septin complexes. Naturally, this system does not allow for post-translational
638 modifications (PTMs), which have been reported in animal cells, such as phosphorylation,
639 acetylation, and sumoylation^{6,51–53}. These posttranslational modifications could be added by
640 implementing a similar purification strategy in insect or human cells. Furthermore, this paper has
641 only discussed the reconstitution of septins by themselves, but studies in cells indicate that
642 regulatory proteins such as proteins from the Borg family^{54,55} and anillin^{24,25,56} can have
643 substantial yet poorly understood effects on the assembly and functions of septins and are,
644 therefore, important to eventually incorporate in *in vitro* studies. Protocols for the purification
645 of Borg proteins and anillin have been reported^{54,57}.

646

647 The septin purification protocol reported here offers a standardized way to purify septins in their
648 oligomer form with the correct subunit stoichiometry, offering an important advance over many
649 earlier *in vitro* studies relying on single septin subunits. Even though some septins in specific
650 contexts can act as a single subunit², the current body of literature strongly suggests that, in
651 animal cells, septins mostly function in complexes^{9,58}. Therefore, the use of pre-formed hetero-
652 oligomers, such as the ones described in this paper and others^{10,11,18,32,35–37}, is of great importance
653 to study the structural and biophysical properties of septins via *in vitro* reconstitution to dissect
654 their functions in the cell. Furthermore, septins are self-assembling proteins with many
655 interaction partners, including the membrane and the cytoskeleton, which makes them of great

656 interest for bottom-up synthetic biology^{59–61} and studies of protein-induced changes in
657 membrane biophysical properties such as curvature^{42,62,63}.

658

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668

669 **DISCLOSURES:**

670 The authors declare no competing or financial interests.

671

672 **REFERENCES:**

- 673 1. Mostowy, S., Cossart, P. Septins: The fourth component of the cytoskeleton. *Nature*
674 *Reviews Molecular Cell Biology*. **13** (3), 183–194 (2012).
- 675 2. Shuman, B., Momany, M. Septins from protists to people. *Frontiers in Cell and*
676 *Developmental Biology*. **9**, 3802 (2022).
- 677 3. Bridges, A. A., Gladfelter, A. S. Septin form and function at the cell cortex. *Journal of*
678 *Biological Chemistry*. **290** (28), 17173–17180 (2015).
- 679 4. Smith, C. et al. Septin 9 exhibits polymorphic binding to F-actin and inhibits myosin and
680 cofilin activity. *Journal of Molecular Biology*. **427** (20), 3273–3284 (2015).
- 681 5. Golden, J. K., Peck, S., Chen, Y. C. M., Krummel, M. F. The septin cytoskeleton facilitates
682 membrane retraction during motility and blebbing. *Journal of Cell Biology* **196** (1), 103–114
683 (2012).
- 684 6. Marquardt, J., Chen, X., Bi, E. Architecture, remodeling, and functions of the septin
685 cytoskeleton. *Cytoskeleton*. **76** (1), 7–14 (2018).
- 686 7. Van Ngo, H., Mostowy, S. Role of septins in microbial infection. *Journal of Cell Science*.
687 **132** (9), jcs226266 (2019).
- 688 8. Fung, K. Y. Y., Dai, L., Trimble, W. S. Cell and molecular biology of septins. *International*
689 *Review of Cell and Molecular Biology*. **310**, 289–339 (2014).
- 690 9. Kinoshita, M., Field, C. M., Coughlin, M. L., Straight, A. F., Mitchison, T. J. Self- and actin-
691 templated assembly of mammalian septins. *Developmental Cell*. **3** (6), 791–802 (2002).
- 692 10. Iv, F. et al. Insights into animal septins using recombinant human septin octamers 2 with
693 distinct SEPT9 isoforms. *Journal of Cell Science*. **134** (15), jcs258484 (2021).
- 694 11. Szuba, A. et al. Membrane binding controls ordered self-assembly of animal septins. *eLife*.
695 **10**, e63349 (2021).
- 696 12. Kinoshita, M. Assembly of mammalian septins. *Journal of Biochemistry*. **134** (4), 491–496
697 (2003).
- 698 13. Connolly, D. et al. Septin 9 isoform expression, localization and epigenetic changes during
699 human and mouse breast cancer progression. *Breast Cancer Research*. **13** (4), R76 (2011).

- 700 14. Connolly, D. et al. Septin 9 amplification and isoform-specific expression in peritumoral
701 and tumor breast tissue. *Biological Chemistry*. **395** (2), 157–167 (2014).
- 702 15. Estey, M. P., Di Ciano-Oliveira, C., Froese, C. D., Bejide, M. T., Trimble, W. S. Distinct roles
703 of septins in cytokinesis: SEPT9 mediates midbody abscission. *Journal of Cell Biology*. **191** (4),
704 741–749 (2010).
- 705 16. John, C. M. et al. The *Caenorhabditis elegans* septin complex is nonpolar. *EMBO Journal*.
706 **26** (14), 3296–3307 (2007).
- 707 17. Field, C. M. et al. A purified *Drosophila* septin complex forms filaments and exhibits
708 GTPase activity. *Journal of Cell Biology*. **133** (3), 605–616 (1996).
- 709 18. Bertin, A. et al. *Saccharomyces cerevisiae* septins: Supramolecular organization of
710 heterooligomers and the mechanism of filament assembly. *Proceedings of the National Academy
711 of Sciences of the United States of America*. **105** (24), 8274–8279 (2008).
- 712 19. Sellin, M. E., Sandblad, L., Stenmark, S., Gullberg, M. Deciphering the rules governing
713 assembly order of mammalian septin complexes. *Molecular Biology of the Cell*. **22** (17), 3152–
714 3164 (2011).
- 715 20. Akil, A. et al. Septin 9 induces lipid droplets growth by a phosphatidylinositol-5-phosphate
716 and microtubule-dependent mechanism hijacked by HCV. *Nature Communications*. **7**, 12203
717 (2016).
- 718 21. Tanaka-Takiguchi, Y., Kinoshita, M., Takiguchi, K. Septin-mediated uniform bracing of
719 phospholipid membranes. *Current Biology*. **19** (2), 140–145 (2009).
- 720 22. Omrane, M. et al. Septin 9 has two polybasic domains critical to septin filament assembly
721 and Golgi integrity. *iScience*. **13**, 138–153 (2019).
- 722 23. Carim, S. C., Kechad, A., Hickson, G. R. X. Animal cell cytokinesis: The rho-dependent
723 actomyosin-anilloseptin contractile ring as a membrane microdomain gathering, compressing,
724 and sorting machine. *Frontiers in Cell and Developmental Biology*. **8**, 575226 (2020).
- 725 24. Amine, N. El, Kechad, A., Jananji, S., Hickson, G. R. X. Opposing actions of septins and
726 Sticky on Anillin promote the transition from contractile to midbody ring. *Journal of Cell Biology*.
727 **203** (3), 487–504 (2013).
- 728 25. Renshaw, M. J., Liu, J., Lavoie, B. D., Wilde, A. Anillin-dependent organization of septin
729 filaments promotes intercellular bridge elongation and Chmp4B targeting to the abscission site.
730 *Open Biology*. **4** (1), 130190 (2014).
- 731 26. Vogt, E. T. et al. The ultrastructural organization of actin and myosin II filaments in the
732 contractile ring: new support for an old model of cytokinesis. *Molecular Biology of the Cell*. **28**
733 (5), 613–623 (2017).
- 734 27. Mavrakis, M. et al. Septins promote F-actin ring formation by crosslinking actin filaments
735 into curved bundles. *Nature Cell Biology*. **16** (4), 322–334 (2014).
- 736 28. Karasmanis, E. P. et al. A septin double ring controls the spatiotemporal organization of
737 the ESCRT machinery in cytokinetic abscission. *Current Biology*. **29** (13), 2174–2182.e7 (2019).
- 738 29. Hagiwara, A. et al. Submembranous septins as relatively stable components of actin-
739 based membrane skeleton. *Cytoskeleton*. **68** (9), 512–525 (2011).
- 740 30. Calvo, F. et al. Cdc42EP3/BORG2 and septin network enables mechano-transduction and
741 the emergence of cancer-associated fibroblasts. *Cell Reports*. **13** (12), 2699–2714 (2015).
- 742 31. Salameh, J., Cantaloube, I., Benoit, B., Poüs, C., Baillet, A. Cdc42 and its BORG2 and BORG3
743 effectors control the subcellular localization of septins between actin stress fibers and

744 microtubules. *Current Biology*. **31** (18), 4088–4103.e5 (2021).

745 32. Kuzmić, M. et al. Septin-microtubule association via a motif unique to isoform 1 of septin
746 9 tunes stress fibers. *Journal of Cell Science*. **135** (1), jcs258850 (2022).

747 33. Shindo, A. et al. Septin-dependent remodeling of cortical microtubule drives cell
748 reshaping during epithelial wound healing. *Journal of Cell Science*. **131** (12), jcs212647 (2018).

749 34. Hu, Q., Nelson, W. J., Spiliotis, E. T. Forchlorfenuron alters mammalian septin assembly,
750 organization, and dynamics. *Journal of Biological Chemistry*. **283** (43), 29563–29571 (2008).

751 35. Mavrakis, M., Tsai, F. C., Koenderink, G. H. Purification of recombinant human and
752 *Drosophila* septin hexamers for TIRF assays of actin–septin filament assembly. *Methods in Cell*
753 *Biology*. **136**, 199–220 (2016).

754 36. Nakos, K., Radler, M. R., Spiliotis, E. T. Septin 2/6/7 complexes tune microtubule plus-end
755 growth and EB1 binding in a concentration- and filament-dependent manner. *Molecular Biology*
756 *of the Cell*. **30** (23), 2913–2928 (2019).

757 37. Kaplan, C. et al. Absolute arrangement of subunits in cytoskeletal septin filaments in cells
758 measured by fluorescence microscopy. *Nano Letters*. **15** (6), 3859–3864 (2015).

759 38. Castro, D. K. S. V. et al. A complete compendium of crystal structures for the human SEPT3
760 subgroup reveals functional plasticity at a specific septin interface. *IUCr*. **7** (Pt 3), 462–479
761 (2020).

762 39. Jiao, F., Cannon, K. S., Lin, Y.-C., Gladfelter, A. S., Scheuring, S. The hierarchical assembly
763 of septins revealed by high-speed AFM. *Nature Communications*. **11** (1), 1–13 (2020).

764 40. Bertin, A. et al. Phosphatidylinositol-4,5-bisphosphate promotes budding yeast septin
765 filament assembly and organization. *Journal of Molecular Biology*. **404** (4), 711–731 (2010).

766 41. Bridges, A. A., Jentzsch, M. S., Oakes, P. W., Occhipinti, P., Gladfelter, A. S. Micron-scale
767 plasma membrane curvature is recognized by the septin cytoskeleton. *Journal of Cell Biology*. **213**
768 (1), 23–32 (2016).

769 42. Beber, A. et al. Membrane reshaping by micrometric curvature sensitive septin filaments.
770 *Nature Communications*. **10**, 420 (2019).

771 43. Zhou, R., Shi, Y., Yang, G. Expression, purification, and enzymatic characterization of
772 intramembrane proteases. in *Methods in Enzymology*. **584**, 127–155 (2017).

773 44. Diebold, M. L., Fribourg, S., Koch, M., Metzger, T., Romier, C. Deciphering correct
774 strategies for multiprotein complex assembly by co-expression: Application to complexes as large
775 as the histone octamer. *Journal of Structural Biology*. **175** (2), 178–188 (2011).

776 45. Lebedeva, M. A., Palmieri, E., Kukura, P., Fletcher, S. P. Emergence and rearrangement of
777 dynamic supramolecular aggregates visualized by interferometric scattering microscopy. *ACS*
778 *Nano*. **14** (9), 11160–11168 (2020).

779 46. Ludtke, S. J., Baldwin, P. R., Chiu, W. EMAN: Semiautomated software for high-resolution
780 single-particle reconstructions. *Journal of Structural Biology*. **128** (1), 82–97 (1999).

781 47. Zivanov, J. et al. New tools for automated high-resolution cryo-EM structure
782 determination in RELION-3. *eLife*. **7**, e42166 (2018).

783 48. Frank, J. et al. SPIDER and WEB: Processing and visualization of images in 3D electron
784 microscopy and related fields. *Journal of Structural Biology*. **116** (1), 190–199 (1996).

785 49. Young, G., Kukura, P. Interferometric scattering microscopy. *Annual Review of Physical*
786 *Chemistry*. **70**, 301–322 (2019).

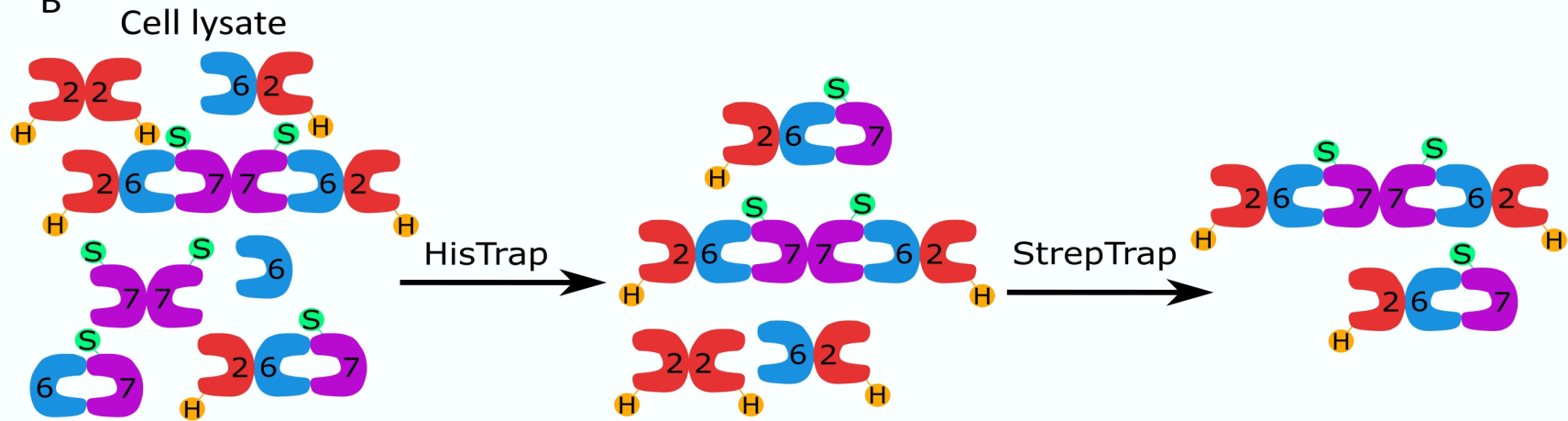
787 50. Young, G. et al. Quantitative mass imaging of single biological macromolecules. *Science*.

788 **360** (6387), 423–427 (2018).
789 51. Hernández-Rodríguez, Y., Momany, M. Posttranslational modifications and assembly of
790 septin heteropolymers and higher-order structures. *Current Opinion in Microbiology*. **15** (6), 660–
791 668 (2012).
792 52. Ribet, D. et al. SUMOylation of human septins is critical for septin filament bundling and
793 cytokinesis. *Journal of Cell Biology*. **216** (12), 4041–4052 (2017).
794 53. Sinha, I. et al. Cyclin-dependent kinases control septin phosphorylation in *Candida*
795 *albicans* hyphal development. *Developmental Cell*. **13** (3), 421–432 (2007).
796 54. Sheffield, P. J. et al. Borg/Septin interactions and the assembly of mammalian septin
797 heterodimers, trimers, and filaments. *Journal of Biological Chemistry*. **278** (5), 3483–3488 (2003).
798 55. Joberty, G. et al. Borg proteins control septin organization and are negatively regulated
799 by Cdc42. *Nature Cell Biology*. **3** (10), 861–866 (2001).
800 56. Chen, X., Wang, K., Svitkina, T., Bi, E. Critical roles of a RhoGEF-anillin module in septin
801 architectural remodeling during cytokinesis. *Current Biology*. **30** (8), 1477-1490.e3 (2020).
802 57. Kučera, O. et al. Anillin propels myosin-independent constriction of actin rings. *Nature*
803 *Communications*. **12** (1), 1–12 (2021).
804 58. Hsu, S. C. et al. Subunit composition, protein interactions, and structures of the
805 mammalian brain sec6/8 complex and septin filaments. *Neuron*. **20** (6), 1111–1122 (1998).
806 59. Olivi, L. et al. Towards a synthetic cell cycle. *Nature Communications*. **12** (1), 1–11 (2021).
807 60. Hürtgen, D., Härtel, T., Murray, S. M., Sourjik, V., Schwille, P. Functional modules of
808 minimal cell division for synthetic biology. *Advanced Biosystems*. **3** (6), e1800315 (2019).
809 61. Jia, H., Schwille, P. Bottom-up synthetic biology: Reconstitution in space and time. *Current*
810 *Opinion in Biotechnology*. **60**, 179–187 (2019).
811 62. Cannon, K. S., Woods, B. L., Crutchley, J. M., Gladfelter, A. S. An amphipathic helix enables
812 septins to sense micrometer-scale membrane curvature. *The Journal of Cell Biology*. **218** (4),
813 1128–1137 (2019).
814 63. Lobato-Márquez, D., Mostowy, S. Septins recognize micron-scale membrane curvature.
815 *Journal of Cell Biology*. **213** (1), 5–6 (2016).
816

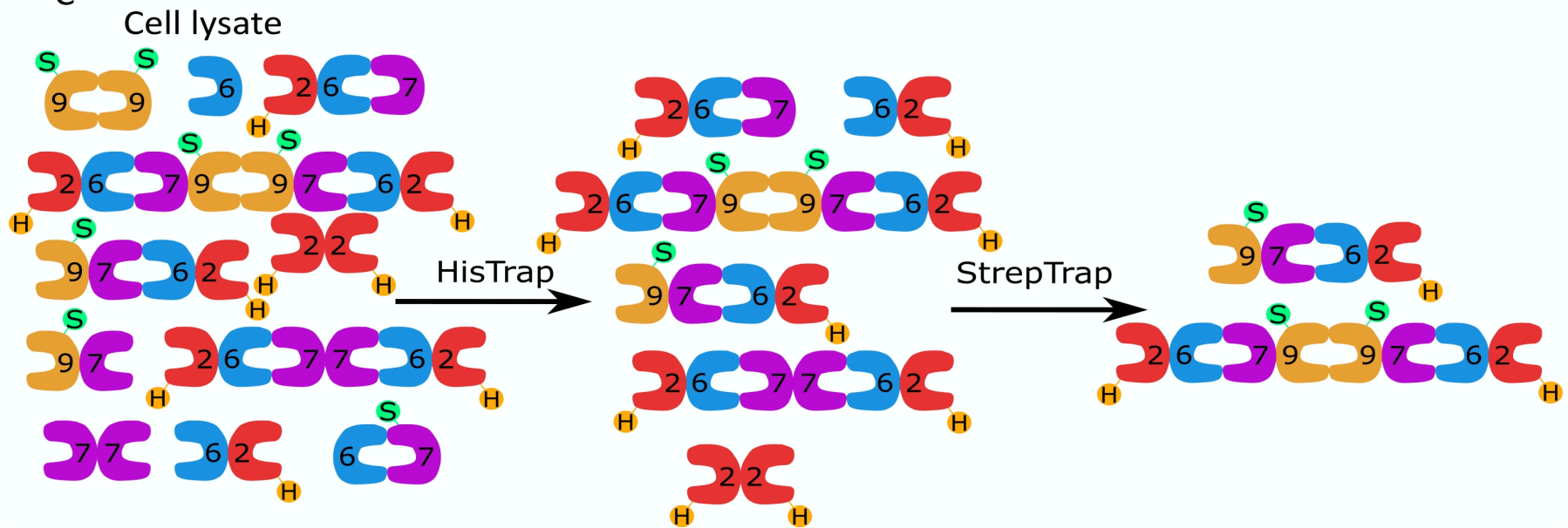
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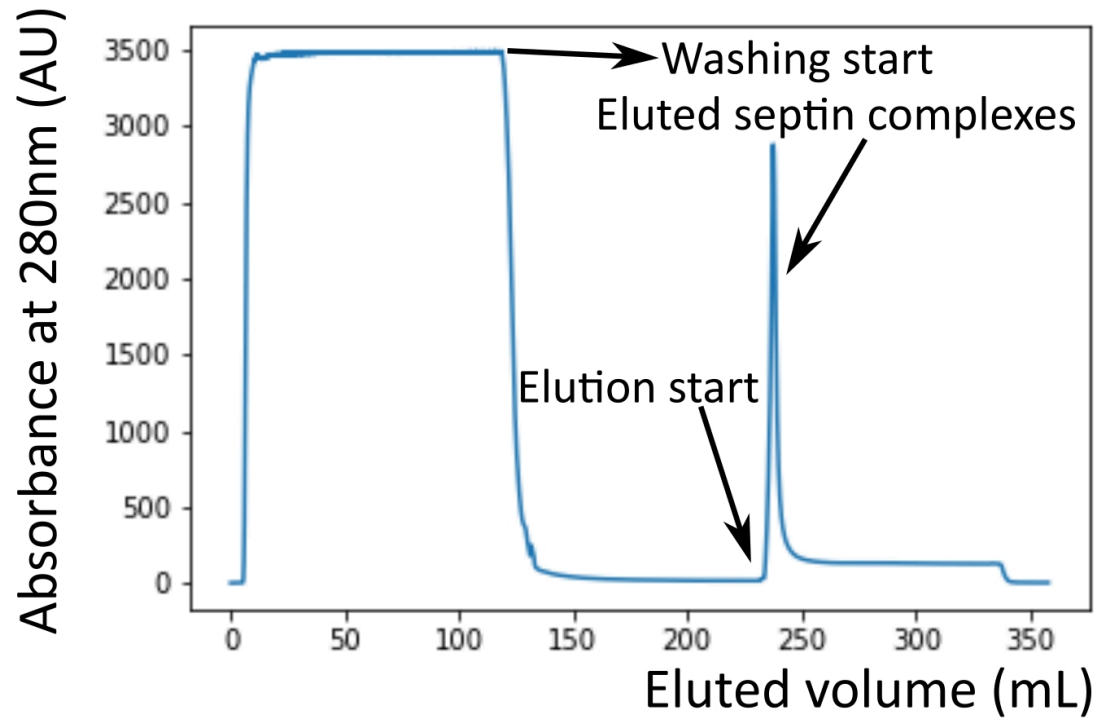


B



C



A**HisTrap Chromatogram****B****StrepTrap Chromatogram**