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Battle for Metals: Regulatory RNAs at the Front Line

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Metal such as iron, zinc, manganese, and nickel are essential elements for bacteria. These nutrients are required in crucial structural and catalytic roles in biological processes, including precursor biosynthesis, DNA replication, transcription, respiration, and oxidative stress responses. While essential, in excess these nutrients can also be toxic. The immune system leverages both of these facets, to limit bacterial proliferation and combat invaders. Metal binding immune proteins reduce the bioavailability of metals at the infection sites starving intruders, while immune cells intoxicate pathogens by providing metals in excess leading to enzyme mismetallation and/or reactive oxygen species generation. In this dynamic metal environment, maintaining metal homeostasis is a critical process that must be precisely coordinated. To achieve this, bacteria utilize diverse metal uptake and efflux systems controlled by metalloregulatory proteins. Recently, small regulatory RNAs (sRNAs) have been revealed to be critical post-transcriptional regulators, working in conjunction with transcription factors to promote rapid adaptation and to fine-tune bacterial adaptation to metal abundance. In this mini review, we discuss the expanding role for sRNAs in iron homeostasis, but also in orchestrating adaptation to the availability of other metals like manganese and nickel. Furthermore, we describe the sRNA-mediated interdependency between metal homeostasis and oxidative stress responses, and how regulatory networks controlled by sRNAs contribute to survival and virulence.

Keywords: Regulatory RNA, metal ions, metal homeostasis, nutritional immunity, oxidative stress

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INTRODUCTION

Trace metals like iron, zinc, manganese, or nickel are essential nutrients for bacteria (Begg, 2019), being cofactors and/or structural components of ~40% of proteins (Andreini et al., 2008). At the same time, these essential nutrients can also be toxic. Since they cannot be synthesized or degraded, bacteria, including pathogens, must adapt to their presence and absence, which is particularly important in the context of infection. During infection, the host renders metals inaccessible to invaders, a process termed "nutritional immunity" (Hood and Skaar, 2012; Nairz et al., 2018; Núñez et al., 2018). This includes extracellular metal withholding through systemic and locally secreted metal-binding proteins such as transferrin, lactoferrin and calprotectin, and metal depletion from

phagosomes by host transporters such as NRAMP1. The host also harnesses the toxicity of metals and intoxicates pathogens with high metal levels (Imlay, 2003; Imlay, 2014; Djoko et al., 2015). This mini review will focus on how pathogens adapt to metal limitation and the role of regulatory RNAs in this response and maintaining metal homeostasis. However, it is important to note that the need to adapt changing metal abundance is not restricted to pathogens and that common strategies are used by both pathogenic and environmental microbes.

Bacterial Countermeasures

Bacteria have evolved multiple mechanisms to maintain metal homeostasis in response to their ever-changing environments. This includes metal importers and exporters, metal storage proteins, and alternative enzymes/pathways to preserve critical enzymatic and metabolic functions (Merchant and Helmann, 2012; Chandrangsu et al., 2017).

Metal import and export systems are critical for bacterial survival and virulence. For instance, in *Staphylococcus aureus*, the uptake of manganese is mediated by the NRAMP homolog MntH and the ABC-type transporter MntABC, while its efflux is carried out by MntE. These systems are crucial not only to maintain manganese homeostasis, but also to resist oxidative stress (Grunenwald et al., 2019; Radin et al., 2019). To efficiently extract metal from their environment, many pathogens also secrete metallophores, molecules having a higher metal-binding affinity than host proteins. Siderophores, iron-chelating secondary metabolites, are important virulence factors used to obtain iron (Kramer et al., 2020; Khasheii et al., 2021). More recently, metallophores that promote zinc uptake and contribute to pathogenesis were identified in *Pseudomonas aeruginosa* and *S. aureus* (Grim et al., 2017; Lhospice et al., 2017). Another approach to obtain these essential nutrients is to steal them from host metal-containing proteins like transferrin, lactoferrin or hemoglobin for iron (Barber and Elde, 2015) and calprotectin for zinc (Stork et al., 2013).

While important for pathogenesis, the expression of importers is insufficient to enable infection. To cope with metal limitation, bacteria can release iron from storage proteins like ferritin and ferritin-related proteins (Chandrangsu et al., 2017; Bradley et al., 2020). They also alter the composition of the cytoplasm, which buffers metal ions, potentially facilitating metal acquisition by metalloproteins. In response to scarcity, bacteria also reduce their need for the limiting metal, by switching to alternative isozymes or even entire pathways that are not dependent on it (Merchant and Helmann, 2012). A classic example is the use of paralogs of the superoxide dismutase (SOD) associated with distinct metal cofactors to rescue the reactive oxygen species (ROS) detoxification pathway (see below and **Figure 1**).

While beneficial when metal availability is restricted, these adaptations can be deleterious when metals are abundant. Similarly, the adaptations used to overcome metal intoxication, reviewed by Djoko et al. (2015) and Bradley et al. (2020), can be detrimental when metals are limiting. Thus, use of all above-mentioned mechanisms is carefully coordinated by regulators.

METAL-RESPONSIVE TRANSCRIPTION FACTORS

Classically, the bacterial response to environmental metal abundance is controlled by regulators that directly interact with the target metal. These regulators can be divided into two classes; those that coordinate the response to metal limitation and those that respond to intoxication. Transcription factors that coordinate the response to metal limitation generally repress expression when their target metal is abundant, with converse logic being used to respond to intoxication (Waldron and Robinson, 2009). Metal limitation is frequently sensed by members of either the Fur (Ferric uptake regulator) or DtxR (Diphtheria toxin regulator) family of metal sensing regulators, with divergence enabling the same family to be used to sense different metals (**Table 1**). While primarily negative regulators, positive regulation by these families can occur, for example two MntRs that positively regulate the expression of manganese efflux systems have been reported (Huang et al., 2017; Grunenwald et al., 2019). The Fur family has also been co-opted to coordinate their response to peroxide stress *via* PerR (Bsat et al., 1998; Lee and Helmann, 2006; Pinochet-Barros and Helmann, 2018). In addition, bacteria can leverage protein scaffolds not generally considered metal-responsive to sense metals such as the MarR family proteins, AdcR and ZitR (Varela et al., 2019). While coordinating the response to metal limitation was once thought to be the purview of metal sensing regulators, it is now apparent that metal independent sensors also critically contribute (Nairn et al., 2016; Radin et al., 2016; Harper et al., 2018; Párraga Solórzano et al., 2019; Lonergan et al., 2020). Similarly, there is a growing appreciation for post-transcriptional regulation *via* small regulatory RNAs (sRNAs).

WHAT ARE BACTERIAL REGULATORY RNAS?

To enable tighter and fine-tuned regulation, transcription factors are frequently associated with sRNAs into so-called mixed regulatory circuits (Nitzan et al., 2017). Regulatory RNAs in bacteria are generally non-coding and range in size from ~30 to >1,000 nts (Barrientos et al., 2021; Li and Stanton, 2021). Hundreds of sRNAs have been identified in diverse bacteria (Boutet et al., 2022). These critical post-transcriptional regulators control, amongst others, bacterial physiology, stress responses and virulence in response to specific internal or external stimuli such as nutrient availability, oxidative stress, or antibiotics exposure (Chakravarty and Massé, 2019; Mediati et al., 2021).

Regulatory RNAs are mainly categorized in two classes, cis- and trans-encoded. The cis-encoded sRNAs include antisense RNAs and riboswitches that originate from genes located at the same locus as the targeted mRNA. Antisense RNAs are encoded on the opposite DNA strand and, consequently, regulate their cognate mRNA target *via* perfect base-pairings. They are notably involved in toxin-antitoxin systems (Sarpong and Murphy, 2021). Riboswitches are regulatory elements embedded within

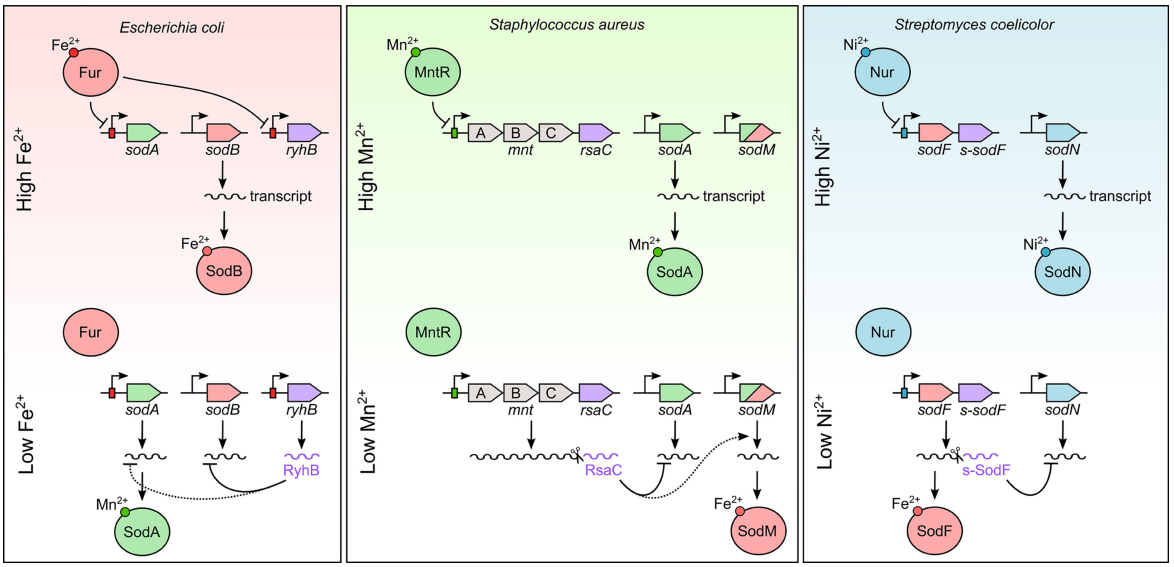


FIGURE 1 | Mixed regulatory circuits between transcription factors and sRNAs to control SOD synthesis according to metal bioavailability. Fur (red), MntR (green) and Nur (blue) boxes are indicated by a rectangle overlapping the promoter. sRNAs are indicated in purple. Dotted lines represent putative or indirect regulation in need of further experimentation. See text for more details.

the 5' untranslated region (5'UTR) of mRNA targets (Breaker, 2022). Through their aptamer domain, riboswitches sense metabolites or metal ions. Upon ligand recognition, riboswitches are subject to structural modifications which modulate the transcription and/or translation of downstream gene(s).

Trans-encoded sRNAs and their respective targets are located at distant loci. These regulatory RNAs usually regulate multiple targets, from a few to several tens, through imperfect base-pairings (Carrier et al., 2018; Jørgensen et al., 2020). A broad range of regulatory mechanisms are used to control targeted mRNAs positively or negatively. For instance, sRNAs can pair with the Shine-Dalgarno sequence of a specific mRNA, which restricts access to the ribosome binding site and thus blocks mRNA translation. Conversely, sRNAs binding can mask cleavage sites or induce conformational changes promoting mRNA translation. Several RNA-binding proteins, such as Hfq, ProQ and CsrA, play crucial roles in the sRNA-dependent regulation, especially in Gram-negative bacteria. Their roles have been widely discussed in Christopoulou and Granneman (2021) and Quendera et al. (2020).

SRNAS AND METAL HOMEOSTASIS

A mere 20 years ago, Massé and Gottesman described the first metal-responsive sRNA, RyhB, which helps reestablish iron homeostasis in starved *E. coli*. Since then, accumulating data

TABLE 1 | Transcription factors and sRNAs that respond to metal limitation.

	Metal[a]	Cognate sRNA	Exemplar species
Fur family[c]			
Fur	Fe	RyhB and analogs	*E. coli*, *S.* Typhimurium, *P. aeruginosa*, *B. subtilis*...
Zur	Zn	NR	
Mur	Mn	NR	
Nur	Ni	s-SodF	*S. coelicolor*
Irr	Heme	NR	
PerR	Fe, Mn, H_2O_2	NR	
DtxR family[b,d]			
DtxR	Fe	NR	
MntR	Mn	RsaC	*S. aureus*
MarR family[e]			
AdcR/ZitR	Zn	NR	
Ribbon-helix-helix family[f]			
NikR	Ni	NikS	*H. pylori*

NR. None reported. [a]Metal(s) or stimuli that modulate activity in the native organism. [b]The nomenclature of the DtxR family is heterogeneous with considerable species specificity. Reviewed by: [c]Sevilla et al. (2021), [d]Merchant and Spatafora (2014), [e]Varela et al. (2019), [f]Li and Zamble (2009).

revealed that sRNA-mediated adaptive responses to metal fluctuations is not restricted to the model organism *E. coli*, or to iron.

Iron Homeostasis and RyhB-like sRNAs

RyhB responds to intracellular iron levels as it is directly under the control of Fur (**Table 1**). While Fe^{2+}-Fur efficiently represses *ryhB* expression, Fur becomes inactive upon intracellular iron depletion (Masse and Gottesman, 2002). Once produced, RyhB directly interacts with a large set of mRNAs to boost iron import, reduce cellular demands, and redirect iron to essential biological processes (e.g., respiration, DNA synthesis). For more details about this sRNA-dependent iron-sparing response, please refer to Chareyre and Mandin (2018).

Multiple analogs of RyhB have been discovered among bacteria [e.g., PrrF1/2 in *P. aeruginosa* (Wilderman et al., 2004) and FsrA in *Bacillus subtilis* (Gaballa et al., 2008)]. While RyhB-like sRNAs share little to no sequence similarities, they remarkably control a similar set of transcripts and belong to the Fur regulon. In pathogens, RyhB-like sRNAs interlink iron homeostasis and virulence. Porcheron et al. (2014) demonstrated that the deletion of *ryhB* gene in the uropathogenic *E. coli* strain CFT073 leads to a significant reduction of bladder colonization. In *Salmonella* Typhimurium, RyhB-1/2 sRNAs modulate bacterial replication within macrophages and presumably promote immune evasion (Peñaloza et al., 2021). Additional examples are provided by Porcheron and Dozois (2015) and Chareyre and Mandin (2018).

Similar to other adaptations to metal limitation, the synthesis of RyhB sRNA can cause adverse effects on cell physiology and colicin resistance (Lalaouna et al., 2015). Therefore, its production is tightly controlled at both transcriptional and post-transcriptional levels. Lalaouna et al. (2015) identified a tRNA-derived fragment, which protects *E. coli* cells from deleterious effects by "sponging" the transcriptional noise of *ryhB* gene in non-inducing conditions. Other RyhB-like sponging mechanisms have been thereafter described in *E. coli* (AspX; Chen et al., 2021) and in *P. aeruginosa* (SkatA; Han et al., 2016).

Fur-Independent sRNAs Involved in Iron Homeostasis

In addition to RyhB, several other sRNAs including CsrB/C, CyaR and FnrS have been linked with iron homeostasis.

The RNA-binding protein CsrA and its homologs are pleiotropic post-transcriptional regulators, which control multiple cellular processes including carbon metabolism, stress response, mobility, and virulence (Romeo et al., 1993; Pourciau et al., 2020). In *E. coli*, CsrA also plays a role in iron homeostasis (Pourciau et al., 2019). CsrA reportedly modulates iron storage protein synthesis to remobilize iron during the exponential phase of growth. CsrA also seems to lower RyhB sRNA level *via* an unknown mechanism (Potts et al., 2017). The activity of CsrA is modulated by two sRNAs, named CsrB and CsrC (Liu et al., 1997; Weilbacher et al., 2003), in response to short-chain carboxylic acids and carbon nutritional status (Pannuri et al., 2016; Alvarez et al., 2021). In stark contrast with above-mentioned mechanisms of action, CsrB/C sRNAs bind the CsrA protein *via* recognition motif mimicry, preventing it from interacting with its 'true' targets. Noteworthy, both CsrB/C and CsrA are not responsive to iron bioavailability (Pourciau et al., 2019). Hence, CsrA, and by extension, CsrB/C sRNAs could link cellular responses to iron homeostasis notably by integrating additional signals.

Other sRNAs control specific iron-related mRNA targets as recently exemplified by Sy and Tree (2022). The *chuAS* operon encodes a haem receptor and haem oxygenase, involved in haem uptake, and then iron dissociation. This operon is not only regulated by RyhB and Fur in enterohemorrhagic *E. coli*, but also by CyaR and FnrS sRNAs. CyaR activates ChuA translation in response to high cyclic AMP levels, while FnrS sRNA represses ChuS translation during anaerobiosis. Hence, *chuAS* expression is fine-tuned at the infection site by integrating distinct parameters such as oxygen and nutrient bioavailability.

Manganese-Responsive RNAs

Investigations into metal-responsive RNAs has largely focused on the intersection with iron homeostasis. However, it is now apparent that sRNAs broadly contribute to metal homeostasis. The manganese-responsive sRNA RsaC originates from the 3'UTR of *mntABC* transcript coding for the main manganese ABC transporter in *S. aureus* and is tightly repressed by Mn^{2+}-MntR (**Table 1**) (Lalaouna et al., 2019). When *S. aureus* faces manganese starvation, both the MntABC transporter and RsaC sRNA are produced. Lalaouna et al. (2019) demonstrated that, after its release from its precursor through an endonucleolytic cleavage, RsaC modulates the switch between manganese-dependent and iron-utilizing superoxide dismutases (SODs), linking manganese homeostasis to oxidative stress response. More details are provided below and in **Figure 1**. While further investigation is necessary, RsaC could also be a bridge between distinct metal-related networks. In response to manganese limitation, RsaC potentially regulates the transcription factor Zur, the zinc transporter AdcABC, the Fe^{3+}-siderophore transporter SstABCD and the iron-sulfur cluster biosynthesis system Suf (Lalaouna et al., 2019).

Regulatory RNAs also contribute to resisting intoxication as the synthesis of bacterial manganese tolerance/efflux systems are controlled by MntR and/or the Mn^{2+}-sensing *yybP-ykoY* riboswitch (Dambach et al., 2015; Zeinert et al., 2018; Waters, 2020). Upon Mn^{2+} binding, the *yybP-ykoY* riboswitch either releases the ribosome binding site or forms an antitermination structure allowing the translation or transcription of the downstream gene, respectively. Remarkably, this riboswitch is highly conserved, as over 1,300 bacterial genomes contain one or two *yybP-ykoY* motifs (Zeinert et al., 2018).

Nickel Homeostasis and Virulence

Regulatory RNAs also contribute to nickel homeostasis and pathogenesis. *Helicobacter pylori*, responsible for benign to severe stomach pathologies, must survive the acidic conditions encountered in the stomach. To that end, *H. pylori* neutralizes acid by producing a nickel-dependent urease, converting urea

into ammonia and bicarbonate. The transcription of *ureAB* mRNA encoding the two structural subunits of urease is activated by NikR in presence of Ni^{2+}. To prevent advert effects of alkalinization, an antisense RNA to *ureB* is produced in response to elevated pH to lower urease synthesis (Wen et al., 2011; Wen et al., 2013).

More directly connecting nickel-responsive sRNAs and virulence, in 2017, (Vannini et al. (2017) identified three putative NikR-regulated sRNAs. One of these, HPnc4160, renamed NikS, especially controls major virulence and colonization factors including the vacuolating cytotoxin VacA and the carcinogenic protein CagA (Eisenbart et al., 2020; Kinoshita-Daitoku et al., 2021). These examples highlight the role of metal-responsive sRNAs in cell survival and virulence, but also the necessity to integrate multiple environmental signals *via* mixed regulatory circuits to adapt to the host environment.

A conserved metal-sensing riboswitch, named *czcD* and located upstream genes encoding putative cation efflux pumps, has been described as nickel, cobalt and iron-responsive. However, the nature of its ligand is under debate and its physiological role still needs to be clarified *in vivo* (Furukawa et al., 2015; Xu and Cotruvo, 2020).

The Regulation of Superoxide Dismutases, an Illustrative Example

Superoxide dismutases are ubiquitous enzymes that detoxify superoxide, a toxic compound generated either by aerobic respiration or by immune cells (Abreu and Cabelli, 2010). SODs rely on diverse metal cofactors to function (i.e., manganese, iron, nickel, or zinc/copper). Many bacteria have more than one SOD, each reliant on a different metal cofactor, to cope with fluctuations in metal bioavailability. Regulatory RNAs (RyhB, RsaC and s-SodF) and their cognate transcription factors (Fur, MntR and Nur) (**Figure 1**; **Table 1**) have critical roles in coordinating expression of these multiple SODs in response to metal abundance.

E. coli possesses two cytoplasmic (SodA and SodB) and one periplasmic SODs (SodC), which differ in their metal needs and in their temporal regulation. The iron-dependent SodB and the manganese-dependent SodA enzymes are tightly and divergently controlled in response to iron and oxygen abundance (Fee, 1991; Compan and Touati, 1993). As mentioned above, RyhB directly targets several mRNAs coding for non-essential iron-containing proteins such as *sodB* mRNA to spare intracellular iron (Masse and Gottesman, 2002) during iron limitation (**Figure 1**). Under these conditions, the manganese-dependent SOD enzyme, SodA, is induced due to loss of Fe^{2+}-Fur mediated repression (Niederhoffer et al., 1990; Compan and Touati, 1993), reestablishing the ROS detoxification pathway. However, RyhB also seems to negatively regulate *sodA* mRNA (Jacques et al., 2006; Argaman et al., 2012). While the molecular rationale for this contrasting regulation is unknown, Semsey (2014) proposed that regulation by both Fur and RyhB allows a faster SodA-dependent response to superoxide stress during iron starvation. This paradoxical regulation of SodA for an sRNA as well studies as RyhB highlights the broader need for continued investigation into the role of sRNAs in metal homeostasis. It should be also noted that there is no evidence that RyhB could control *sodC* mRNA, even in large-scale analyses (Lalaouna et al., 2015; Melamed et al., 2016).

The *S. aureus* genome encodes two SOD enzymes (**Figure 1**), the manganese-dependent SodA and the cambialistic enzyme SodM using either manganese or iron to function (Garcia et al., 2017). As discussed above, the MntR-dependent RsaC sRNA directly blocks SodA synthesis during manganese starvation and facilitates its substitution by the iron-associated SodM (Lalaouna et al., 2019). RsaC likely enables *S. aureus* to spare manganese for essential manganese-containing proteins, avoid the synthesis of a non-functional enzyme and reestablish the ROS detoxification pathway. It is noteworthy that both *rsaC* and *sodM* genes have been acquired exclusively by *S. aureus* and closely related strains, a clear advantage under harsh and selective conditions compared to other staphylococcal species.

SOD regulation also highlights the broad importance of sRNA mediated regulation to non-pathogenic microbes. Indeed, two distinct SODs are encoded in *Streptomyces coelicolor* genome, the nickel-containing SodN and the iron-containing SodF (Kim et al., 2014). These enzymes are antagonistically regulated in response to the cellular level of nickel (Ahn et al., 2006). The nickel-responsive transcriptional regulator of the Fur family, Nur, directly binds to *sodF* promoter and switches its transcription off, while indirectly turns on SodN production (**Figure 1**). Kim et al. (2014) raised the veil on this mystery by identifying s-SodF, an sRNA originating from the 3'UTR of *sodF* mRNA during nickel starvation. This 3'UTR-derived sRNA pairs with *sodN* transcript, induces its degradation and impedes its translation, favoring a nickel-dependent detoxification pathway. Consequently, Nur activates *sodN* transcription *via* the negative regulation of *sodF* transcription.

CONCLUSIONS

Metal-responsive transcription factors and associated sRNAs are involved in intricate regulatory networks, enabling to finely regulate metal homeostasis in response to environmental fluctuations and metal-based immune strategies. This phenomenon is not restricted to *E. coli* and is rather widely distributed among bacteria. These mixed regulatory circuits also play important roles in the virulence and survival of pathogenic strains.

As exemplified above, sRNA-dependent regulations are not limited to iron homeostasis. Manganese and nickel intracellular levels are also tightly regulated *via* trans-encoded RNAs and riboswitches. It is also very likely that other sRNA-dependent metal-sparing responses exist due to the imperative need to balance the import and export of zinc, cobalt or copper. We also highlighted that virulence, metal homeostasis and oxidative stress responses are intimately linked *via* transcription factors and their cognate sRNAs, allowing bacteria to cope with multifactorial environmental fluctuations.

AUTHOR CONTRIBUTIONS

MC, GG-E, TK-F and DL contributed to the conceptualization and writing of the manuscript. All authors have read and approved the submitted version.

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REFERENCES

Abreu, I. A., and Cabelli, D. E. (2010). Superoxide Dismutases-a Review of the Metal-Associated Mechanistic Variations. *Biochim. Biophys. Acta* 1804 (2), 263–274. doi: 10.1016/j.bbapap.2009.11.005

Ahn, B. E., Cha, J., Lee, E. J., Han, A. R., Thompson, C. J., and Roe, J. H. (2006). Nur, a Nickel-Responsive Regulator of the Fur Family, Regulates Superoxide Dismutases and Nickel Transport in Streptomyces Coelicolor. *Mol. Microbiol.* 59 (6), 1848–1858. doi: 10.1111/j.1365-2958.2006.05065.x

Alvarez, A. F., Rodríguez, C., González-Chávez, R., and Georgellis, D. (2021). The Escherichia Coli Two-Component Signal Sensor BarA Binds Protonated Acetate *via* a Conserved Hydrophobic-Binding Pocket. *J. Biol. Chem.* 297 (6), 101383. doi: 10.1016/j.jbc.2021.101383

Andreini, C., Bertini, I., Cavallaro, G., Holliday, G. L., and Thornton, J. M. (2008). Metal Ions in Biological Catalysis: From Enzyme Databases to General Principles. *J. Biol. Inorg. Chem.* 13 (8), 1205–1218. doi: 10.1007/s00775-008-0404-5

Argaman, L., Elgrably-Weiss, M., Hershko, T., Vogel, J., and Altuvia, S. (2012). RelA Protein Stimulates the Activity of RyhB Small RNA by Acting on RNA-Binding Protein Hfq. *Proc. Natl. Acad. Sci. U.S.A.* 109 (12), 4621–4626. doi: 10.1073/pnas.1113113109

Barber, M. F., and Elde, N. C. (2015). Buried Treasure: Evolutionary Perspectives on Microbial Iron Piracy. *Trends Genet.* 31 (11), 627–636. doi: 10.1016/j.tig.2015.09.001

Barrientos, L., Mercier, N., Lalaouna, D., and Caldelari, I. (2021). Assembling the Current Pieces: The Puzzle of RNA-Mediated Regulation in Staphylococcus Aureus. *Front. Microbiol.* 12. doi: 10.3389/fmicb.2021.706690

Begg, S. L. (2019). The Role of Metal Ions in the Virulence and Viability of Bacterial Pathogens. *Biochem. Soc. Trans.* 47 (1), 77–87. doi: 10.1042/bst20180275

Boutet, E., Djerroud, S., and Perreault, J. (2022). Small RNAs Beyond Model Organisms: Have We Only Scratched the Surface? *Int. J. Mol. Sci.* 23 (8), 4448. doi: 10.3390/ijms23084448

Bradley, J. M., Svistunenko, D. A., Wilson, M. T., Hemmings, A. M., Moore, G. R., and Le Brun, N. E. (2020). Bacterial Iron Detoxification at the Molecular Level. *J. Biol. Chem.* 295 (51), 17602–17623. doi: 10.1074/jbc.REV120.007746

Breaker, R. R. (2022). The Biochemical Landscape of Riboswitch Ligands. *Biochemistry* 61 (3), 137–149. doi: 10.1021/acs.biochem.1c00765

Bsat, N., Herbig, A., Casillas-Martinez, L., Setlow, P., and Helmann, J. D. (1998). Bacillus Subtilis Contains Multiple Fur Homologues: Identification of the Iron Uptake (Fur) and Peroxide Regulon (PerR) Repressors. *Mol. Microbiol.* 29 (1), 189–198. doi: 10.1046/j.1365-2958.1998.00921.x

Carrier, M. C., Lalaouna, D., and Masse, E. (2018). Broadening the Definition of Bacterial Small RNAs: Characteristics and Mechanisms of Action. *Annu. Rev. Microbiol.* 72, 141–161. doi: 10.1146/annurev-micro-090817-062607

Chakravarty, S., and Massé, E. (2019). RNA-Dependent Regulation of Virulence in Pathogenic Bacteria. *Front. Cell. Infect. Microbiol.* 9. doi: 10.3389/fcimb.2019.00337

Chareyre, S., and Mandin, P. (2018). Bacterial Iron Homeostasis Regulation by sRNAs. *Microbiol Spectr.* 6(2). doi: 10.1128/microbiolspec.RWR-0010-2017

Chandrangsu, P., Rensing, C., and Helmann, J. D. (2017). Metal Homeostasis and Resistance in Bacteria. *Nat. Rev. Microbiol.* 15 (6), 338–350. doi: 10.1038/nrmicro.2017.15

Chen, J., To, L., de Mets, F., Luo, X., Majdalani, N., Tai, C. H., et al. (2021). A Fluorescence-Based Genetic Screen Reveals Diverse Mechanisms Silencing Small RNA Signaling in E. Coli. *Proc. Natl. Acad. Sci. U. S. A.* 118 (27), e2106964118. doi: 10.1073/pnas.2106964118

Christopoulou, N., and Granneman, S. (2021). The Role of RNA-Binding Proteins in Mediating Adaptive Responses in Gram-Positive Bacteria. *FEBS J* 289(7), 1746–1764. doi: 10.1111/febs.15810

Compan, I., and Touati, D. (1993). Interaction of Six Global Transcription Regulators in Expression of Manganese Superoxide Dismutase in Escherichia Coli K-12. *J. Bacteriol.* 175 (6), 1687–1696. doi: 10.1128/jb.175.6.1687-1696.1993

Dambach, M., Sandoval, M., Updegrove, T. B., Anantharaman, V., Aravind, L., Waters, L. S., et al. (2015). The Ubiquitous yybP-ykoY Riboswitch Is a Manganese-Responsive Regulatory Element. *Mol. Cell* 57 (6), 1099–1109. doi: 10.1016/j.molcel.2015.01.035

Djoko, K. Y., Ong, C. L., Walker, M. J., and McEwan, A. G. (2015). The Role of Copper and Zinc Toxicity in Innate Immune Defense Against Bacterial Pathogens. *J. Biol. Chem.* 290 (31), 18954–18961. doi: 10.1074/jbc.R115.647099

Eisenbart, S. K., Alzheimer, M., Pernitzsch, S. R., Dietrich, S., Stahl, S., and Sharma, C. M. (2020). A Repeat-Associated Small RNA Controls the Major Virulence Factors of Helicobacter Pylori. *Mol. Cell* 80 (2), 210–226.e217. doi: 10.1016/j.molcel.2020.09.009

Fee, J. A. (1991). Regulation of Sod Genes in Escherichia Coli: Relevance to Superoxide Dismutase Function. *Mol. Microbiol.* 5 (11), 2599–2610. doi: 10.1111/j.1365-2958.1991.tb01968.x

Furukawa, K., Ramesh, A., Zhou, Z., Weinberg, Z., Vallery, T., Winkler, W. C., et al. (2015). Bacterial Riboswitches Cooperatively Bind Ni(2+) or Co(2+) Ions and Control Expression of Heavy Metal Transporters. *Mol. Cell* 57 (6), 1088–1098. doi: 10.1016/j.molcel.2015.02.009

Gaballa, A., Antelmann, H., Aguilar, C., Khakh, S. K., Song, K. B., Smaldone, G. T., et al. (2008). The Bacillus Subtilis Iron-Sparing Response is Mediated by a Fur-Regulated Small RNA and Three Small, Basic Proteins. *Proc. Natl. Acad. Sci. U.S.A.* 105 (33), 11927–11932. doi: 10.1073/pnas.0711752105

Garcia, Y. M., Barwinska-Sendra, A., Tarrant, E., Skaar, E. P., Waldron, K. J., and Kehl-Fie, T. E. (2017). A Superoxide Dismutase Capable of Functioning With Iron or Manganese Promotes the Resistance of Staphylococcus Aureus to Calprotectin and Nutritional Immunity. *PloS Pathog.* 13 (1), e1006125. doi: 10.1371/journal.ppat.1006125

Grim, K. P., San Francisco, B., Radin, J. N., Brazel, E. B., Kelliher, J. L., Párraga Solórzano, P. K., et al. (2017). The Metallophore Staphylopine Enables Staphylococcus Aureus To Compete With the Host for Zinc and Overcome Nutritional Immunity. *mBio* 8 (5), e01281–17. doi: 10.1128/mBio.01281-17

Grunenwald, C. M., Choby, J. E., Juttukonda, L. J., Beavers, W. N., Weiss, A., Torres, V. J., et al. (2019). Manganese Detoxification by MntE Is Critical for Resistance to Oxidative Stress and Virulence of Staphylococcus Aureus. *mBio* 10 (1), e02915–18. doi: 10.1128/mBio.02915-18

Han, K., Tjaden, B., and Lory, S. (2016). GRIL-Seq Provides a Method for Identifying Direct Targets of Bacterial Small Regulatory RNA by *In Vivo* Proximity Ligation. *Nat. Microbiol.* 2, 16239–16239. doi: 10.1038/nmicrobiol.2016.239

Harper, L., Balasubramanian, D., Ohneck, E. A., Sause, W. E., Chapman, J., Mejia-Sosa, B., et al. (2018). Staphylococcus Aureus Responds to the Central Metabolite Pyruvate To Regulate Virulence. *mBio* 9 (1), e02272–17. doi: 10.1128/mBio.02272-17

Hood, M. I., and Skaar, E. P. (2012). Nutritional Immunity: Transition Metals at the Pathogen-Host Interface. *Nat. Rev. Microbiol.* 10 (8), 525–537. doi: 10.1038/nrmicro2836

Huang, X., Shin, J. H., Pinochet-Barros, A., Su, T. T., and Helmann, J. D. (2017). Bacillus Subtilis MntR Coordinates the Transcriptional Regulation of Manganese Uptake and Efflux Systems. *Mol. Microbiol.* 103 (2), 253–268. doi: 10.1111/mmi.13554

Imlay, J. A. (2003). Pathways of Oxidative Damage. *Annu. Rev. Microbiol.* 57, 395–418. doi: 10.1146/annurev.micro.57.030502.090938

Imlay, J. A. (2014). The Mismetallation of Enzymes During Oxidative Stress. *J. Biol. Chem.* 289 (41), 28121–28128. doi: 10.1074/jbc.R114.588814

Jørgensen, M. G., Pettersen, J. S., and Kallipolitis, B. H. (2020). sRNA-Mediated Control in Bacteria: An Increasing Diversity of Regulatory Mechanisms. *Biochim. Biophys. Acta Gene Regul. Mech.* 1863 (5), 194504. doi: 10.1016/j.bbagrm.2020.194504

Jacques, J. F., Jang, S., Prévost, K., Desnoyers, G., Desmarais, M., Imlay, J., et al. (2006). RyhB Small RNA Modulates the Free Intracellular Iron Pool and is Essential for Normal Growth During Iron Limitation in Escherichia Coli. *Mol. Microbiol.* 62 (4), 1181–1190. doi: 10.1111/j.1365-2958.2006.05439.x

Khasheii, B., Mahmoodi, P., and Mohammadzadeh, A. (2021). Siderophores: Importance in Bacterial Pathogenesis and Applications in Medicine and Industry. *Microbiol. Res.* 250, 126790. doi: 10.1016/j.micres.2021.126790

Kim, H. M., Shin, J. H., Cho, Y. B., and Roe, J. H. (2014). Inverse Regulation of Fe- and Ni-Containing SOD Genes by a Fur Family Regulator Nur Through Small RNA Processed From 3'UTR of the sodF mRNA. *Nucleic Acids Res.* 42 (3), 2003–2014. doi: 10.1093/nar/gkt1071

Kinoshita-Daitoku, R., Kiga, K., Miyakoshi, M., Otsubo, R., Ogura, Y., Sanada, T., et al. (2021). A Bacterial Small RNA Regulates the Adaptation of Helicobacter Pylori to the Host Environment. *Nat. Commun.* 12 (1), 2085–2085. doi: 10.1038/s41467-021-22317-7

Kramer, J., Özkaya, Ö., and Kümmerli, R. (2020). Bacterial Siderophores in Community and Host Interactions. *Nat. Rev. Microbiol.* 18 (3), 152–163. doi: 10.1038/s41579-019-0284-4

Lalaouna, D., Baude, J., Wu, Z., Tomasini, A., Chicher, J., Marzi, S., et al. (2019). RsaC sRNA Modulates the Oxidative Stress Response of Staphylococcus Aureus During Manganese Starvation. *Nucleic Acids Res.* 47 (1), 9871–9887. doi: 10.1093/nar/gkz728

Lalaouna, D., Carrier, M. C., Semsey, S., Brouard, J. S., Wang, J., Wade, J. T., et al. (2015). A 3' External Transcribed Spacer in a tRNA Transcript Acts as a Sponge for Small RNAs to Prevent Transcriptional Noise. *Mol. Cell* 58 (3), 393–405. doi: 10.1016/j.molcel.2015.03.013

Lee, J. W., and Helmann, J. D. (2006). The PerR Transcription Factor Senses H2O2 by Metal-Catalysed Histidine Oxidation. *Nature* 440 (7082), 363–367. doi: 10.1038/nature04537

Lhospice, S., Gomez, N. O., Ouerdane, L., Brutesco, C., Ghssein, G., Hajjar, C., et al. (2017). Pseudomonas Aeruginosa Zinc Uptake in Chelating Environment is Primarily Mediated by the Metallophore Pseudopaline. *Sci. Rep.* 7 (1), 17132. doi: 10.1038/s41598-017-16765-9

Li, Z., and Stanton, B. A. (2021). Transfer RNA-Derived Fragments, the Underappreciated Regulatory Small RNAs in Microbial Pathogenesis. *Front. Microbiol.* 12. doi: 10.3389/fmicb.2021.687632

Liu, M. Y., Gui, G., Wei, B., Preston, J. F., Oakford, L., Yüksel, U., et al. (1997). The RNA Molecule CsrB Binds to the Global Regulatory Protein CsrA and Antagonizes its Activity in Escherichia Coli. *J. Biol. Chem.* 272 (28), 17502–17510. doi: 10.1074/jbc.272.28.17502

Li, Y., and Zamble, D. B. (2009). Nickel Homeostasis and Nickel Regulation: An Overview. *Chem. Rev.* 109 (10), 4617–4643. doi: 10.1021/cr900010n

Lonergan, Z. R., Palmer, L. D., and Skaar, E. P. (2020). Histidine Utilization Is a Critical Determinant of Acinetobacter Pathogenesis. *Infect. Immun.* 88 (7), e00118–20. doi: 10.1128/iai.00118-20

Masse, E., and Gottesman, S. (2002). A Small RNA Regulates the Expression of Genes Involved in Iron Metabolism in Escherichia Coli. *Proc. Natl. Acad. Sci. U.S.A.* 99 (7), 4620–4625. doi: 10.1073/pnas.032066599

Mediati, D. G., Wu, S., Wu, W., and Tree, J. J. (2021). Networks of Resistance: Small RNA Control of Antibiotic Resistance. *Trends Genet.* 37 (1), 35–45. doi: 10.1016/j.tig.2020.08.016

Melamed, S., Peer, A., Faigenbaum-Romm, R., Gatt, Y. E., Reiss, N., Bar, A., et al. (2016). Global Mapping of Small RNA-Target Interactions in Bacteria. *Mol. Cell* 63 (5), 884–897. doi: 10.1016/j.molcel.2016.07.026

Merchant, S. S., and Helmann, J. D. (2012). Elemental Economy: Microbial Strategies for Optimizing Growth in the Face of Nutrient Limitation. *Adv. Microb. Physiol.* 60, 91–210. doi: 10.1016/b978-0-12-398264-3.00002-4

Merchant, A. T., and Spatafora, G. A. (2014). A Role for the DtxR Family of Metalloregulators in Gram-Positive Pathogenesis. *Mol. Oral. Microbiol.* 29 (1), 1–10. doi: 10.1111/omi.12039

Nairn, B. L., Lonergan, Z. R., Wang, J., Braymer, J. J., Zhang, Y., Calcutt, M. W., et al. (2016). The Response of Acinetobacter Baumannii to Zinc Starvation. *Cell Host Microbe* 19 (6), 826–836. doi: 10.1016/j.chom.2016.05.007

Nairz, M., Dichtl, S., Schroll, A., Haschka, D., Tymoszuk, P., Theurl, I., et al. (2018). Iron and Innate Antimicrobial Immunity-Depriving the Pathogen, Defending the Host. *J. Trace Elem. Med. Biol.* 48, 118–133. doi: 10.1016/j.jtemb.2018.03.007

Niederhoffer, E. C., Naranjo, C. M., Bradley, K. L., and Fee, J. A. (1990). Control of Escherichia Coli Superoxide Dismutase (sodA and Sodb) Genes by the Ferric Uptake Regulation (Fur) Locus. *J. Bacteriol.* 172 (4), 1930–1938. doi: 10.1128/jb.172.4.1930-1938.1990

Nitzan, M., Rehani, R., and Margalit, H. (2017). Integration of Bacterial Small RNAs in Regulatory Networks. *Annu. Rev. Biophysics.* 46 (1), 131–148. doi: 10.1146/annurev-biophys-070816-034058

Núñez, G., Sakamoto, K., and Soares, M. P. (2018). Innate Nutritional Immunity. *J. Immunol.* 201 (1), 11–18. doi: 10.4049/jimmunol.1800325

Pannuri, A., Vakulskas, C. A., Zere, T., McGibbon, L. C., Edwards, A. N., Georgellis, D., et al. (2016). Circuitry Linking the Catabolite Repression and Csr Global Regulatory Systems of Escherichia Coli. *J. Bacteriol.* 198 (21), 3000–3015. doi: 10.1128/jb.00454-16

Párraga Solórzano, P. K., Yao, J., Rock, C. O., and Kehl-Fie, T. E. (2019). Disruption of Glycolysis by Nutritional Immunity Activates a Two-Component System That Coordinates a Metabolic and Antihost Response by Staphylococcus Aureus. *mBio* 10 (4), e01321–19. doi: 10.1128/mBio.01321-19

Peñaloza, D., Acuña, L. G., Barros, M. J., Núñez, P., Montt, F., Gil, F., et al. (2021). The Small RNA RyhB Homologs From Salmonella Typhimurium Restrain the Intracellular Growth and Modulate the SPI-1 Gene Expression Within RAW264.7 Macrophages. *Microorganisms* 9 (3), 635. doi: 10.3390/microorganisms9030635

Pinochet-Barros, A., and Helmann, J. D. (2018). Redox Sensing by Fe(2+) in Bacterial Fur Family Metalloregulators. *Antioxid. Redox Signal* 29 (18), 1858–1871. doi: 10.1089/ars.2017.7359

Porcheron, G., and Dozois, C. M. (2015). Interplay Between Iron Homeostasis and Virulence: Fur and RyhB as Major Regulators of Bacterial Pathogenicity. *Vet. Microbiol.* 179 (1-2), 2–14. doi: 10.1016/j.vetmic.2015.03.024

Porcheron, G., Habib, R., Houle, S., Caza, M., Lépine, F., Daigle, F., et al. (2014). The Small RNA RyhB Contributes to Siderophore Production and Virulence of Uropathogenic Escherichia Coli. *Infect. Immun.* 82 (12), 5056–5068. doi: 10.1128/iai.02287-14

Potts, A. H., Vakulskas, C. A., Pannuri, A., Yakhnin, H., Babitzke, P. , and Romeo, T. (2017). Global Role of the Bacterial Post-Transcriptional Regulator CsrA Revealed by Integrated Transcriptomics. *Nat. Commun.* 8 (1), 1596. doi: 10.1038/s41467-017-01613-1

Pourciau, C., Lai, Y. J., Gorelik, M., Babitzke, P., and Romeo, T. (2020). Diverse Mechanisms and Circuitry for Global Regulation by the RNA-Binding Protein CsrA. *Front. Microbiol.* 11, e01034–19. doi: 10.3389/fmicb.2020.601352

Pourciau, C., Pannuri, A., Potts, A., Yakhnin, H., Babitzke, P., and Romeo, T. (2019). Regulation of Iron Storage by CsrA Supports Exponential Growth of Escherichia Coli. *mBio* 10 (4). doi: 10.1128/mBio.01034-19

Quendera, A. P., Seixas, A. F., dos Santos, R. F., Santos, I., Silva, J. P. N., Arraiano, C. M., et al. (2020). RNA-Binding Proteins Driving the Regulatory Activity of Small Non-Coding RNAs in Bacteria. *Front. Mol. Biosci.* 7. doi: 10.3389/fmolb.2020.00078

Radin, J. N., Kelliher, J. L., Parraga Solorzano, P. K., and Kehl-Fie, T. E. (2016). The Two-Component System ArlRS and Alterations in Metabolism Enable Staphylococcus Aureus to Resist Calprotectin-Induced Manganese Starvation. *PloS Pathog.* 12 (11), e1006040. doi: 10.1371/journal.ppat.1006040

Radin, J. N., Zhu, J., Brazel, E. B., McDevitt, C. A., and Kehl-Fie, T. E. (2019). Synergy Between Nutritional Immunity and Independent Host Defenses Contributes to the Importance of the MntABC Manganese Transporter During Staphylococcus Aureus Infection. *Infect. Immun.* 87 (1), e00642–18. doi: 10.1128/iai.00642-18

Romeo, T., Gong, M., Liu, M. Y., and Brun-Zinkernagel, A. M. (1993). Identification and Molecular Characterization of Csra, a Pleiotropic Gene From Escherichia Coli That Affects Glycogen Biosynthesis, Gluconeogenesis, Cell Size, and Surface Properties. *J. Bacteriol.* 175 (15), 4744–4755. doi: 10.1128/jb.175.15.4744-4755.1993

Sarpong, D. D., and Murphy, E. R. (2021). RNA Regulated Toxin-Antitoxin Systems in Pathogenic Bacteria. *Front. Cell Infect. Microbiol.* 11. doi: 10.3389/fcimb.2021.661026

Semsey, S. (2014). A Mixed Incoherent Feed-Forward Loop Allows Conditional Regulation of Response Dynamics. *PloS One* 9 (3), e91243. doi: 10.1371/journal.pone.0091243

Sevilla, E., Bes, M. T., Peleato, M. L., and Fillat, M. F. (2021). Fur-Like Proteins: Beyond the Ferric Uptake Regulator (Fur) Paralog. *Arch. Biochem. Biophys.* 701, 108770. doi: 10.1016/j.abb.2021.108770

Stork, M., Grijpstra, J., Bos, M. P., Mañas Torres, C., Devos, N., Poolman, J. T., et al. (2013). Zinc Piracy as a Mechanism of Neisseria Meningitidis for Evasion of Nutritional Immunity. *PloS Pathog.* 9 (10), e1003733. doi: 10.1371/journal.ppat.1003733

Sy, B. M., and Tree, J. J. (2022). The Small RNA CyaR Activates Translation of the Outer Membrane Haem Receptor chuA in Enterohemorrhagic Escherichia Coli. *Front. Microbiol.* 13. doi: 10.3389/fmicb.2022.821196

Vannini, A., Pinatel, E., Costantini, P. E., Pelliciari, S., Roncarati, D., Puccio, S., et al. (2017). Comprehensive Mapping of the Helicobacter Pylori NikR Regulon Provides New Insights in Bacterial Nickel Responses. *Sci. Rep.* 7, 45458–45458. doi: 10.1038/srep45458

Varela, P. F., Velours, C., Aumont-Niçaise, M., Pineau, B., Legrand, P., and Poquet, I. (2019). Biophysical and Structural Characterization of a Zinc-Responsive Repressor of the MarR Superfamily. *PloS One* 14 (2), e0210123. doi: 10.1371/journal.pone.0210123

Waldron, K. J., and Robinson, N. J. (2009). How do Bacterial Cells Ensure That Metalloproteins Get the Correct Metal? *Nat. Rev. Microbiol.* 7 (1), 25–35. doi: 10.1038/nrmicro2057

Waters, L. S. (2020). Bacterial Manganese Sensing and Homeostasis. *Curr. Opin. Chem. Biol.* 55, 96–102. doi: 10.1016/j.cbpa.2020.01.003

Weilbacher, T., Suzuki, K., Dubey, A. K., Wang, X., Gudapaty, S., Morozov, I., et al. (2003). A Novel sRNA Component of the Carbon Storage Regulatory System of Escherichia Coli. *Mol. Microbiol.* 48 (3), 657–670. doi: 10.1046/j.1365-2958.2003.03459.x

Wen, Y., Feng, J., and Sachs, G. (2013). Helicobacter Pylori 5'ureb-sRNA, a Cis-Encoded Antisense Small RNA, Negatively Regulates ureAB Expression by Transcription Termination. *J. bacteriol.* 195 (3), 444–452. doi: 10.1128/JB.01022-12

Wen, Y., Feng, J., Scott, D. R., Marcus, E. A., and Sachs, G. (2011). A Cis-Encoded Antisense Small RNA Regulated by the HP0165-HP0166 Two-Component System Controls Expression of ureB in Helicobacter Pylori. *J. bacteriol.* 193 (1), 40–51. doi: 10.1128/JB.00800-10

Wilderman, P. J., Sowa, N. A., FitzGerald, D. J., FitzGerald, P. C., Gottesman, S., Ochsner, U. A., et al. (2004). Identification of Tandem Duplicate Regulatory Small RNAs in Pseudomonas Aeruginosa Involved in Iron Homeostasis. *Proc. Natl. Acad. Sci. U.S.A.* 101 (26), 9792–9797. doi: 10.1073/pnas.0403423101

Xu, J., and Cotruvo, J. A.Jr. (2020). The czcD (NiCo) Riboswitch Responds to Iron (Ii). *Biochemistry* 59 (15), 1508–1516. doi: 10.1021/acs.biochem.0c00074

Zeinert, R., Martinez, E., Schmitz, J., Senn, K., Usman, B., Anantharaman, V., et al. (2018). Structure-Function Analysis of Manganese Exporter Proteins Across Bacteria. *J. Biol. Chem.* 293 (15), 5715–5730. doi: 10.1074/jbc.M117.790717

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