

Adaptation of the virtual fields method for the identification of biphasic hyperelastic model parameters in soft biological tissues with osmotic swelling

Ruike Shi, Stéphane Avril, Haitian Yang, Víctor Acosta Santamaría, Yue Mei,

Yiqian He

► To cite this version:

Ruike Shi, Stéphane Avril, Haitian Yang, Víctor Acosta Santamaría, Yue Mei, et al.. Adaptation of the virtual fields method for the identification of biphasic hyperelastic model parameters in soft biological tissues with osmotic swelling. Strain, In press, 10.1111/str.12435. hal-04035036

HAL Id: hal-04035036 https://cnrs.hal.science/hal-04035036

Submitted on 17 Mar 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Adaptation of the virtual fields method for the identification of biphasic hyperelastic model parameters in soft biological tissues with osmotic swelling

Ruike Shi^a, Stéphane Avril^b, Haitian Yang^a, Víctor A. Acosta Santamaría^b, Yue Mei^a,
 Yiqian He^{a,c,*}

 ^aState Key Lab of Structural Analysis for Industrial Equipment, Department of Engineering Mechanics, Dalian University of Technology, Dalian 116024, P.R. China

 ^bMines Saint-Etienne, University of Lyon, University Jean Monnet, INSERM, SAINBIOSE U1059, F-42023 Saint-Etienne, France

^cMechanik–Materialtheorie, Ruhr-Universität Bochum, Bochum, Germany

11 Abstract

10

Biphasic hyperelastic models have become popular for soft hydrated tissues and there is a pressing need for appropriate identification methods using full-field measurement techniques such as digital volume correlation. This paper proposes to address this need with the virtual fields method (VFM). The main asset of the proposed approach is that it avoids the repeated resolution of complex nonlinear finite-element models. By choosing special virtual fields, the VFM approach can extract hyperelastic parameters of the solid part of the biphasic medium without resorting to identifying the model parameters driving the osmotic effects in the interstitial fluid. The proposed approach is verified and validated through three different examples: first using simulated data and then using experimental data obtained from porcine descending thoracic aortas samples in osmotically active solution.

Keywords: Biphasic hyperelasticity, parameter identification, chemoelasticity, virtual fields
 method, osmotic effects

14 **1. Introduction**

The mechanical behavior of soft biological tissues, such as cartilage or arteries, can be commonly modeled as biphasic, accounting for the intertwined contributions of fiber networks and interstitial fluid^[1]. This biphasic behavior implies chemomechanical couplings regulating the osmotic pressure of the interstitial fluid^[2]. An example of such coupling is the swelling effect that occurs when there is a gradient of osmotic pressure, which can be attributed to the presence of proteoglycans^[3, 4] within collagenous fiber networks in the tissues^[2].

 $[*] Corresponding \ author: heyiqian@dlut.edu.cn$

The biphasic hyperelastic constitutive models of hydrated biological tissues were originally introduced by Eringen and Ingram in the 1960s^[5, 6]. These biphasic models have been widely used in the biomechanics research of soft biological tissues. For instance,

— In the field of the biomechanics of cartilage, Mow et al.^[7] and Lai et al.^[8] estab-24 lished a biphasic model and a triphasic model. These models include mainly the collagen-25 proteoglycan matrix and the interstitial fluid, respectively. Then Ateshian et al.^[9–11] applied 26 for the models to articular cartilage with osmotic loading. Besides, Wilson et al.^[12] studied 27 the links between the biphasic swelling theories and mechano-electrochemical models. Wayne 28 et al.^[13] developed a u-p finite element model for biphasic theory by coupling deformation and 29 fluid pressure. Recently, Kandil et al.^[14] proposed a microstructure-based chemo-viscoelastic 30 model considering the osmo-induced deformation and internal fluid variation. 31

³²— In the field of biomechanics of arteries, Lanir^[1, 4] developed the biocomponent theory ³³ in 1987. He applied this theory to residual stresses of cardiovascular tissues. Then Azeloglu ³⁴ et al.^[15] proposed a numerical model combining the triphasic theory of Lai et al.^[8] with ³⁵ the FEM to study the influence of proteoglycans on residual stresses in the aorta. Their ³⁶ model was eventually validated experimentally. Recently Santamaría et al.^[16, 17] developed ³⁷ experimental methods for quantifying the chemoelastic effects in arterial tissues.

All those studies showed the importance of biphasic or triphasic theory to investigate the biomechanics of hydrated biological tissues, such as cartilage and arteries.

The identification of constitutive model parameters for soft biological tissues is still 40 flourishing^[18, 19]. Among the available identification approaches, the virtual fields method 41 (VFM) has been widely used in solid mechanics^[20–26], given its advantages, such as its insen-42 sitivity to the uncertainty of boundary conditions^[27], robustness^[28], and fast convergence^[29]. 43 Avril et al.^[30] first applied VFM to arterial tissues to identify anisotropic hyperelastic ma-44 terial parameters. Kim et al.^[31] used VFM to identify the material properties of human 45 aneurysmal aortas. Bersi et al.^[32] collected full-field biaxial data and used the VFM to 46 estimate regional variations in material parameters for a microstructurally motivated con-47 stitutive model. 48

The identification of biphasic hyperelastic model parameters has been rather challenging. 49 Nadeen et al.^[33] designed an apparatus for measuring osmotic pressure and estimated how 50 it contributes to cartilage stiffness. More recently, Santamaría et al.^[17] measured 3D strain 51 fields induced by chemoelastic effects in arteries. These studies showed the significance of 52 the biphasic constitutive model in arteries. Nevertheless, to the best of the authors' knowl-53 edge, there has been no existing work dedicated to the identification of biphasic constitutive 54 models using VFM. Besides, although soft tissues (e.g. arterial wall) could be multi-layered 55 structures, there is no work on the identification of layer-specific parameters for soft tissues 56 in the framework of biphasic hyperelastic model. 57

The objective of this work is to address this lack. Considering that biphasic constitutive models are computationally challenging due to coupling between elasticity and osmotic effects, we designed a novel specific VFM framework for chemoelasticity.

⁶¹ The paper is organized as follows: Section 2 introduces the materials and methods, ⁶² including constitutive relationship for biphasic hyperelasticity, general principle for VFM and the application of VFM for biphasic hyperelasticity. Section 3 reports the results of the
verification of the proposed model and its application into practical problems, first conducted
with simulated data and then with experimental data. Finally, the conclusion and discussion
are given in Section 4.

⁶⁷ 2. Materials and methods

⁶⁸ 2.1. The introduction for biphasic hyperelasticity in arterial tissues

⁶⁹ The framework of the biphasic constitutive relationship used in this paper is proposed ⁷⁰ by Azeloglu et al.^[15], in which the residual stresses are hypothesized to arise from the fixed-⁷¹ charge density of the proteoglycans present in the arterial tissues, inducing the Donnan ⁷² osmotic pressure relative to the external environment as only equilibrium conditions are ⁷³ considered for simplicity, when the solid and fluid velocities have reduced to zero. Therefore, ⁷⁴ the Cauchy stress T is assumed to have two components:

$$\boldsymbol{T} = \boldsymbol{T}_i + \boldsymbol{T}_s \tag{1}$$

where T_i is the interstitial fluid component and T_s is the solid matrix component.

As illustrated in Fig. 1, the total deformation gradient \mathbf{F} is realized by the multiplying a prestretched part \mathbf{F}_{pre} and a mechanical part $\mathbf{F}_m^{[14]}$. The \mathbf{F}_{pre} is a stress-free swelling due to osmotic effects without any mechanical loading. Therefore, the total deformation gradient



Figure 1: Multiplicative decomposition of the deformation gradient F

78

⁷⁹ F can be decomposed into two parts

$$\boldsymbol{F} = \boldsymbol{F}_m \boldsymbol{F}_{pre}.$$
 (2)

The interstitial fluid component of Cauchy stress T_i takes the form based on the Donnan equilibrium theory^[15, 34, 35], that is,

$$\boldsymbol{T}_{i} = -(p - p^{*})\boldsymbol{I} = -R\theta \left[\sqrt{\left(\mathbf{c}^{\mathrm{F}}\right)^{2} + \left(\overline{\mathbf{c}}^{*}\right)^{2}} - \overline{\mathbf{c}}^{*}\right]\boldsymbol{I}$$
(3)

where I is the identity tensor, p^* is the ambient pressure in the external bath (henceforth considered to be zero, $p^* = 0$), \bar{c}^* is the external bath salt osmolarity, R is the universal gas constant, and θ is absolute temperature, c^F is the proteoglycan fixed-charge density relating to the solid matrix relative volume change ($J = \det F$) via

$$\mathbf{c}^{\mathrm{F}} = \frac{\varphi_0^{\mathrm{w}} \mathbf{c}_0^{\mathrm{F}}}{J - 1 + \varphi_0^{\mathrm{w}}} \tag{4}$$

where c_0^F and φ_0^w are the fixed-charge density and water content, respectively. Note that in this paper the above chemical parameters are assumed to be homogeneous.

For the solid matrix component, consider an isotropic neo-Hookean constitutive relationship^[36]:

$$\boldsymbol{T}_{s} = \mu J^{-5/3} \left(\boldsymbol{B} - \frac{1}{3} I_{1} \boldsymbol{I} \right) + \kappa (J - 1) \boldsymbol{I}$$
(5)

⁹⁰ where $\boldsymbol{B} = \boldsymbol{F}\boldsymbol{F}^{\mathrm{T}}$ is the left Cauchy-Green deformation tensor, I_1 is the first invariant of ⁹¹ the right Cauchy-Green deformation tensor \boldsymbol{C} , and μ and κ are the shear modulus and bulk ⁹² modulus of the solid matrix, respectively.

In this paper, the reference configuration is set as the zero osmotic loading stage, and a conceptual sequence of configurations is assumed as shown in Fig. 1 according to Kandil et al.^[14], in which the stress-free chemical configuration is an intermediate virtual state due to osmotic swelling. In this way, the total deformation is divided into chemical-induced and mechanical-induced parts. Note that a non-zero prestretched deformation F_{pre} is produced to maintain the Cauchy stress T = 0 in the stress-free chemical configuration. By combining the Eq.(3), Eq.(4) and Eq.(5), and note that the prestretch F_{pre} has the form

$$\boldsymbol{F}_{pre} = \lambda_{pre} \boldsymbol{I} = (J_{pre})^{1/3} \boldsymbol{I}$$
(6)

where $J_{pre} = \det F_{pre}$, we have the equation for solving F_{pre} as

$$-p + \kappa (J_{pre} - 1) = 0 . (7)$$

¹⁰¹ Finally, the prestretch can be calculated as

$$\boldsymbol{F}_{pre} = \left(1 + \frac{p}{\kappa}\right)^{1/3} \boldsymbol{I} .$$
(8)

102 2.2. General principle for VFM

The virtual fields method (VFM) is based on the principle of virtual work, for quasi-static conditions with neglecting the acceleration and the body force^[20], which may be written such as,

$$-\int_{\Omega} \boldsymbol{T} : \boldsymbol{\varepsilon}^* \mathrm{d}\Omega + \int_{\partial\Omega_t} \boldsymbol{t} \cdot \boldsymbol{u}^* \mathrm{d}\partial\Omega_t = 0$$
(9)

where T is the actual Cauchy stress tensor across the domain of interest Ω and is related to the strains through constitutive equations, strains are derived from gradients of the measured displacement field $\boldsymbol{u}, \boldsymbol{t}$ are the tractions applied on a part of the boundary $\partial \Omega_t, \boldsymbol{u}^*$ is a kinematically admissible virtual displacement field, and $\boldsymbol{\varepsilon}^*$ is the associated virtual strain field.

First, a constitutive model is chosen, one can write in the general case

$$\boldsymbol{T} = \mathscr{G}(\boldsymbol{\varepsilon}),\tag{10}$$

where \mathscr{G} is a given function of the actual strain components. The constitutive parameters are also involved in \mathscr{G} . Therefore Eq.(9) can be written as

$$-\int_{\Omega} \mathscr{G}(\boldsymbol{\varepsilon}) : \boldsymbol{\varepsilon}^* \mathrm{d}\Omega + \int_{\partial\Omega_t} \boldsymbol{t} \cdot \boldsymbol{u}^* \mathrm{d}\partial\Omega_t = 0 .$$
 (11)

Then, virtual fields are chosen. Each virtual field introduced into equation Eq.(26) yields 114 one scalar equation. The constitutive parameters are then sought as solutions to a set of such 115 equations. The construction of the virtual fields is a key issue of the method. An important 116 feature is the fact that the above equation is verified for any kinematically admissible virtual 117 field $u^{*[20]}$. Kinematically admissible means that u^* must be continuous across the whole 118 volume and it must be equal to the prescribed displacement on the boundary. The virtual 119 field u^* can be constructed analytically or solving equations created automatically^[37]. The 120 detailed procedure of constructing virtual field for the proposed model in this paper will be 121 introduced later. 122

¹²³ 2.3. The adaptation of the VFM to biphasic hyperelasticity

124 2.3.1. Governing equations

Assuming quasi-static conditions and no body forces^[30], the VFM equations for the biphasic hyperelasticity can be written as

$$-\int_{\Omega} (\boldsymbol{T}_{i} + \boldsymbol{T}_{s}) : \boldsymbol{\varepsilon}^{*} \mathrm{d}\Omega + \int_{\partial \Omega_{t}} \boldsymbol{t} \cdot \boldsymbol{u}^{*} \mathrm{d}\partial \Omega_{t} = 0$$
(12)

where T_s and T_i denotes the solid matrix component and interstitial fluid component for actual Cauchy stress across the domain of interest, respectively, t are the tractions applied on the boundary $\partial \Omega_t$, Ω is the domain volume in the current configuration, and ε^* is the virtual strain field.

¹³¹ 2.3.2. Definition of virtual strain fields

In this paper, the virtual displacement fields are defined by solving a serial of equations. Firstly it is assumed that the osmotic pressure terms vanish under the principle of virtual work with the use of some special virtual fields, i.e. a special virtual field with $tr(\boldsymbol{\varepsilon}^*) = 0$, yielding

$$\int_{\Omega} -p(J)\boldsymbol{I} : \boldsymbol{\varepsilon}^* \mathrm{d}\Omega = 0 .$$
(13)

Further if we consider a three-layers arterial wall tissue for example, including intima, media and adventitia. Then, Eq.(12) can be written as

$$-\sum_{i=1}^{3} \int_{\Omega_{i}} \boldsymbol{T}(\mu_{i}) : \boldsymbol{\varepsilon}^{*} d\Omega + \int_{\partial \Omega_{t}} \boldsymbol{t} \cdot \boldsymbol{u}^{*} d\partial \Omega_{t} = 0$$
(14)

where Ω_i is the domain of the *i*-th layer, and μ_i is the shear modulus in Ω_i .

In order to construct the virtual strain fields ε^* in Eq.(13) a simple approach is to construct a linear virtual displacement field to produce a constant virtual strain field,

$$\boldsymbol{u}^* = (\eta_1 x) \boldsymbol{e}_x + (\eta_2 y) \boldsymbol{e}_y + (\eta_3 z) \boldsymbol{e}_z .$$
(15)

where it is required to make $\eta_1 + \eta_2 + \eta_3 = 0$ for meeting Eq.(13).

For the case of identification for multi-layers in Eq.(14), the virtual fields are required to be linearly independent when there are many unknown parameters to be identified simultaneously, and in order to realize $tr(\boldsymbol{\varepsilon}^*) = 0$, these more complicate virtual fields can be constructed by defining the curl of a potential vector field $\boldsymbol{H}^{[37]}$

$$\boldsymbol{u}^* = \boldsymbol{\nabla} \times \boldsymbol{H} , \qquad (16)$$

¹⁴⁶ since the divergence of the curl above always yields

$$div(\mathbf{\nabla} \times \mathbf{H}) = 0 , \qquad (17)$$

where $H = H_i e_i (i = x, y, z)$ and e_i is the unit vector in the global cartesian coordinate system.

¹⁴⁹ Thus, we can reconstruct virtual displacement fields such as

$$\boldsymbol{u}^* = \boldsymbol{\nabla} \times \boldsymbol{H} = \left(\frac{\partial H_z}{\partial y} - \frac{\partial H_y}{\partial z}\right) \boldsymbol{e}_x + \left(\frac{\partial H_x}{\partial z} - \frac{\partial H_z}{\partial x}\right) \boldsymbol{e}_y + \left(\frac{\partial H_y}{\partial x} - \frac{\partial H_x}{\partial y}\right) \boldsymbol{e}_z .$$
(18)

In this work, under the kinematically admissible condition, we calculated the virtual displacement field by assuming a special case in an uniaxial tension for a plate as shown in Fig. 2, where displacement boundary condition is $u_x|_{x=0} = u_y|_{y=0} = u_z|_{z=0} = 0$ which is usually used, and define the virtual fields as follow:

$$u_x^* = \frac{\partial H_z}{\partial y} - \frac{\partial H_y}{\partial z} = \eta x^k f(z) \tag{19}$$

$$u_y^* = \frac{\partial H_x}{\partial z} - \frac{\partial H_z}{\partial x} = 0 \tag{20}$$

$$u_z^* = \frac{\partial H_y}{\partial x} - \frac{\partial H_x}{\partial y} = \eta z^{k+1} g(x) , \qquad (21)$$

where k is the exponent parameter of the power functions, η is a scaling factor parameter which is usually set to a small value to prevent excessive virtual deformation, f(z) and g(x)are undetermined functions. Note that power functions are used for the virtual displacements in Eq.(19) and (21). The purpose of power functions is to generate virtual strain fields that avoid linear dependent VFM equations.

In order to simply the problem, we set $H_y = 0$, in this way H_x and H_z can be directly calculated by integration leading to

$$H_x = -\eta y \cdot z^{k+1} g(x) \tag{22}$$

$$H_z = \eta y \cdot x^k f(z) . \tag{23}$$

Then, by substituting Eqs.(22) and (23) into Eq.(20), we can deduce that

$$g(x) = -kx^{(k-1)}$$
(24)

$$f(z) = (k+1)z^k . (25)$$

Finally, combining Eqs.(19), (20), (21), (24) and (25), we obtain the general equations for these special virtual fields such as

$$\begin{cases} u_x^* = \eta(k+1)x^k z^k \\ u_y^* = 0 \\ u_z^* = -\eta k x^{k-1} z^{k+1}. \end{cases}$$
(26)

164 2.3.3. Summary of the complete procedure

Based on the theory and methods introduced above, the procedure for the implementation of the proposed model is briefly shown in Table 1, mainly including three parts in the following:

(1)**Full-field measurements:** The displacement fields used in this study have three 168 different possible origins, namely analytical solutions, FE analyses and real experiments. For 169 simple homogeneous (monolayer) uniaxial tension, the analytical solution can be used. For a 170 multilayered structure, the FEM is used to provide simulated measurements and derive the 171 deformation gradient F and the Jacobian J. For the experimental case, the displacements 172 are measured using optical coherence tomography (OCT) and digital volume correlation 173 (DVC) before deducing F, E and J, The traction t for the VFM in numerical verifications is 174 pointwise on the boundary, but as for experiment case, the measured stress is an average force 175 by assuming cross-section perfectly homogeneous as it is hard to measure the heterogeneous 176 traction distribution on the boundary in the experiment. 177

(2)**Building the VFM equations:** From the obtained F fields, we determine T according to Eq.(1),(3),(4),(5). Special virtual fields based on Eq.(26) are applied to Eq.(14). Since T depends on the unknown parameters μ , the virtual work equation Eq.(14) is not satisfied exactly, and a residual can be derived, as defined in Eq.(27). This residual is then minimized. The cost function can be written such as

$$f(\mu_1, \mu_2, \mu_3) = \sum_{v=1}^3 \left[-\sum_{i=1}^3 \int_{\Omega_i} \boldsymbol{T} : \boldsymbol{\varepsilon}^{*v} d\Omega + \int_{\partial\Omega} \boldsymbol{t} \cdot \boldsymbol{u}^{*v} d\partial\Omega \right]^2,$$
(27)

which represents the quadratic deviation between the internal virtual work (IVW) and the external virtual work (EVW), with v labelling the virtual field and i labelling the layer.

Using Gauss quadrature in isoparametric elements, integrals in the cost function can be changed into discrete sums, such as

$$f(\mu_1, \mu_2, \mu_3) = \sum_{v=1}^3 \left[-\sum_{i=1}^3 \sum_{ele} \boldsymbol{T}(\mu_i, \boldsymbol{X}^{ele}) : \boldsymbol{\varepsilon}^{*v}(\boldsymbol{X}^{ele}) + \sum_{eleb} t^{eleb} A^{eleb} \boldsymbol{u}^{*v}(\boldsymbol{X}^{eleb}) \right]^2$$
(28)

where *ele* and *eleb* denote the element and boundary element respectively, A^{eleb} is the area of one element side, $\boldsymbol{\varepsilon}^{*v}(\boldsymbol{X}^{ele})$ denotes the value of the *v*-th virtual strain field at the Gauss integration point in the *ele-th* element, and $\boldsymbol{\varepsilon}^{*v}$ is derived from \boldsymbol{u}^{*v} .

(3) Solving the VFM equations: We use the MATLAB *fminsearch* function to search for μ , in which the derivative-free Nelder-Mead simplex optimization algorithm is used for finding the minimum value of an unconstrained multivariable scalar function^[38]. The problem can be expressed as

$$\begin{cases} \text{Find } \boldsymbol{\mu} = \{\mu_1, \mu_2, \mu_3\} \\ f(\boldsymbol{\mu}) = \sum_{v=1}^3 \left(-W_v^{\text{In}} + W_v^{\text{Ex}}\right)^2 \\ \min f(\boldsymbol{\mu}) \end{cases}$$
(29)

¹⁹⁴ where W_v^{In} represents the IVW under the *v*-th virtual field, and W_v^{Ex} represents the EVW ¹⁹⁵ under the *v*-th virtual field. Eventually, the cost function is minimized through an iterative ¹⁹⁶ scheme and yields the unknown material parameters.

Table 1: Steps of the identification procedure for biphasic hyperelastic parameters

- 1. Forward simulation of chemomechanically coupled model and experimental tests in osmotically active solution.
- 2. Acquisition of full-field displacement from numerical calculation or experimental measurements.
- 3. Computation of deformation gradient F through the given full-field displacement.
- 4. Computation Cauchy stress T from chemomechanically constitutive equations.
- 5. Choice of virtual field that satisfies $tr(\boldsymbol{\varepsilon}^*) = 0$ to avoid the effect of the chemo term.
- 6. Computation of the value of cost function f.
- 7. Use of the optimization method to find a vector of $\boldsymbol{\mu}$ by minimizing f.
- 8. Checking whether the identified value equals the parameters used in forward simulation.

197 2.3.4. Noise investigation method

¹⁹⁸ In order to study the influence of measurement errors on identifications, we add artificial ¹⁹⁹ white noise to the displacement data obtained by the FEM simulations as^[39].

$$u_i^{a,n} = u_i^{a,\text{fem}} + \delta_i^{a,\Delta n} \quad \text{with} \quad \delta_i^{a,\Delta n} \sim \mathcal{N}(0, \ \sigma_u^2) \quad \forall (a,i) \in \Omega$$
(30)

where $u_i^{a,\text{fem}}$ and $\delta_i^{a,\Delta n}$ denote the true displacement from FEM and random noise, respectively, the $\delta_i^{a,\Delta n}$ represents the Gaussian white noise with zero mean and standard deviation σ_u ranging between 10^{-4} (low noise) to 10^{-3} (high noise)^[40]. For each fixed value of σ_u , 40 realizations of Gaussian white noise $\delta_i^{a,\Delta n}$ were generated. The average identified results for all the samples were calculated^[41].

²⁰⁵ The relative error was defined as

$$error = \frac{\widehat{\mu}_i - \overline{\mu}_i}{\widehat{\mu}_i} \times 100\%$$
(31)

where $\hat{\mu}_i$ and $\bar{\mu}_i$ are the exact shear modulus and average identified value under 40 samples respectively.

208 2.4. Verification of the new VFM based on analytical or FEM simulations

In this paper, the proposed model as established in previous section will be verified in theoretical cases before applying into the practical problems. In the verification, the fullfiled measurements are assumed to be 'virtually' obtained from analytical solutions or FE analyses. The derivations for analytical solutions and FE schemes are introduced in the following, respectively.

214 2.4.1. Analytical solution for a plate under tension

For some cases with simple geometry and boundary conditions, the analytical solution of the biphasic hyperelastic problem can be derived. For example, consider a plate under uniaxial tension, as shown in Fig. 2, where λ is the prescribed axial stretch, and α and β are



Figure 2: The geometry and boundary conditions for a plate under tension

217

²¹⁸ unknowns, then total deformation is

$$\boldsymbol{F} = \begin{bmatrix} \lambda & 0 & 0 \\ 0 & \alpha & 0 \\ 0 & 0 & \beta \end{bmatrix}.$$
 (32)

Under this uniaxial tension case, according to Eq.(1),(3),(4),(5), we could have

$$T_{22} = -p + \mu(\lambda\alpha\beta)^{-5/3} \left(\frac{2}{3}\alpha^2 - \frac{1}{3}\lambda^2 - \frac{1}{3}\beta^2\right) + \kappa(\lambda\alpha\beta - 1) = 0$$
(33)

$$T_{33} = -p + \mu(\lambda\alpha\beta)^{-5/3} \left(\frac{2}{3}\beta^2 - \frac{1}{3}\lambda^2 - \frac{1}{3}\alpha^2\right) + \kappa(\lambda\alpha\beta - 1) = 0.$$
(34)

The relationship $\alpha = \beta$ can be derived from the equations above, therefore the Eq.(34) can be rewritten as

$$T_{33} = -p + \mu(\lambda\beta^2)^{-5/3} \left(\frac{1}{3}\beta^2 - \frac{1}{3}\lambda^2\right) + \kappa(\lambda\beta^2 - 1) = 0$$
(35)

where p based on Eq.(3) and (4), can be elaborated as

$$p = \left(R\theta \left[\sqrt{\left(\frac{\varphi_0^{\mathrm{w}} c_0^{\mathrm{F}}}{\lambda \beta^2 - 1 + \varphi_0^{\mathrm{w}}} \right)^2 + \left(\overline{c}^*\right)^2} - \overline{c}^* \right] + p^* \right) .$$
(36)

²²³ To solve this nonlinear problem, the Newton method is applied, yielding

$$\beta_{n+1} = \beta_n - \frac{T_{33}(\beta_n)}{T_{33}'(\beta_n)}$$
(37)

where β_n is the *n*-th iteration of β . For any λ , β and α , hence **F** can be deduced with this iterative resolution.

226 2.4.2. FEM solutions

The FEM is an alternative approach when analytical resolution is not possible. Let $\mathbf{x} = \boldsymbol{\varphi}(\mathbf{X}, t)$ describe the motion of a body from the initial reference configuration $\mathbf{X} \in k(0)$ to its current configuration $\mathbf{x} \in k(t)$. Define the deformation gradient as

$$\boldsymbol{F} = \nabla_{\boldsymbol{X}} \boldsymbol{\varphi} \;. \tag{38}$$

²³⁰ The general total potential energy for the biphasic hyperelasticity model is

$$\Pi = \int_{\Omega} \Psi d\Omega - \int_{\Omega} \bar{\boldsymbol{b}} \cdot \boldsymbol{\varphi} d\Omega - \int_{\partial \Omega} \bar{\boldsymbol{t}} \cdot \boldsymbol{\varphi} d\partial \Omega .$$
(39)

where Ψ is the strain energy, $\bar{\boldsymbol{b}}$ is the body force vector per unit current volume of Ω and $\bar{\boldsymbol{t}}$ are the tractions on the boundary $\partial\Omega$.

Minimization of the potential energy with respect to φ results in a nonlinear system of equation, and the variational forms in the spatial description are

$$\int_{\Omega} \boldsymbol{T} : \nabla_{\boldsymbol{x}} \delta \boldsymbol{\varphi} \mathrm{d}\Omega - \int_{\Omega} \bar{\boldsymbol{b}} \cdot \delta \boldsymbol{\varphi} \mathrm{d}\Omega - \int_{\partial \Omega} \bar{\boldsymbol{t}} \cdot \delta \boldsymbol{\varphi} \mathrm{d}\partial\Omega = 0 .$$
(40)

In finite element discretization, isoparametric interpolations of geometry variables X and field variables φ are written as

$$\boldsymbol{X} = \sum_{I=1}^{n} N_{I}(\xi) \boldsymbol{X}_{I}, \quad \boldsymbol{\varphi} = \sum_{I=1}^{n} N_{I}(\xi) \boldsymbol{\varphi}_{I}$$
(41)

where ξ denotes the coordinates in the reference element, and n is the number of nodes in each element.

Substituting Eq.(41) into Eq.(40) and using the Newton-Raphson scheme to solve the nonlinear system of equations^[42], we obtain

$$\boldsymbol{R}_{\boldsymbol{\varphi}}^{i} + \boldsymbol{K}_{\boldsymbol{\varphi}}^{i} \cdot \Delta \boldsymbol{\varphi}^{i+1} = \boldsymbol{0}$$
(42)

241 where R_{φ} is the global residual vector, and the element stiffness matrix is given by

$$\boldsymbol{K}_{\boldsymbol{\varphi}} = \int_{\Omega} \nabla_{\boldsymbol{x}}^{T} \boldsymbol{N} \cdot \mathbb{C} \cdot \nabla_{\boldsymbol{x}} \boldsymbol{N} d\Omega + \int_{\Omega} [\nabla_{\boldsymbol{x}}^{T} \boldsymbol{N} \cdot \boldsymbol{T} \cdot \nabla_{\boldsymbol{x}} \boldsymbol{N}] \boldsymbol{I} d\Omega$$
(43)

²⁴² and the spatial elasticity tensor \mathbb{C} is given by

$$C = C^{osmo} + C^e \tag{44}$$

where \mathbb{C}^{e} is the spatial elasticity tensor for the solid matrix, which can be written as

$$\mathbb{C}^{e} = 4J^{-1}(\mathbf{F}\underline{\otimes}\mathbf{F}) : \frac{\partial^{2}W}{\partial \mathbf{C}^{2}} : \left(\mathbf{F}^{\mathrm{T}}\underline{\otimes}\mathbf{F}^{\mathrm{T}}\right)$$
(45)

in which $C = F^{T}F$ is the right Cauchy-Green deformation tensor, and W is a strain energy density function for the solid. Here, a hyperelastic constitutive model^[36] is used and is expressed as

$$W = \frac{1}{2}\mu \left(J^{-2/3} I_1 - 3 \right) + \frac{1}{2}\kappa (J - 1)^2$$
(46)

and \mathbb{C}^{osmo} in Eq.(44) is the tensor of the osmotic modulus,

$$\mathbb{C}^{osmo} = J^{-1}(\mathbf{F} \underline{\otimes} \mathbf{F}) : 2 \frac{\partial (-Jp\mathbf{C}^{-1})}{\partial \mathbf{C}} : (\mathbf{F}^{\mathrm{T}} \underline{\otimes} \mathbf{F}^{\mathrm{T}})$$
$$= -J \frac{\partial p}{\partial J} \mathbf{I} \otimes \mathbf{I} + p(2\mathbf{I} \underline{\overline{\otimes}} \mathbf{I} - \mathbf{I} \otimes \mathbf{I})$$
(47)

which results in part from the change in osmotic pressure with a change in tissue volume^[9, 33]. The detailed expression of \mathbb{C}^{osmo} can be found in Azeloglu et al.^[15].

250 2.5. Application to experimental data

Uniaxial tensile tests were previously performed to stimulate and investigate chemoelastic effects in arteries^[17]. In these experiments, porcine descending thoracic aortas were excised and used as test samples. Propylene glycol (PG) and phosphate-buffered saline (PBS) were mixed to prepare an osmotically active solution.

Three rectangular samples $(10 \times 58 \text{mm})$, named Aorta1, Aorta2 and Aorta3, were immersed in an 80% PG solution and tested in uniaxial tension (stepwise stress-relaxation tests). The loading direction was aligned with (Y-axis). Strain fields were measured during these tests using optical coherence tomography (OCT) and digital volume correlation $(\text{DVC})^{[16]}$.

Before carrying out the stepwise uniaxial tensile tests, a 1.15 stretch preconditioning (displacement increment of 5.2mm) was applied. The initial distance between clamps was 35.1mm, as shown in Fig. 3.



Figure 3: Setup used for obtaining experimental datasets and acquisitions of experimental data using the OCT-DVC technique^[17]

OCT volumetric images were acquired under a field of view (FOV) of $2 \times 4 \times 2.51$ mm (X-, Y- and Z-axes, respectively, see Fig. 3(b)). Four specific regions of interest were defined on the OCT volume images: the entire aortic wall (global), the intima region, the media region and the adventitia region. 3D displacement fields were measured using DVC with a local correlation algorithm (LA-DVC). Ideal correlation conditions were reached directly in the 3D OCT image sequence acquisition and using a rigid body translation test. The implemented correlation parameters can be found in our previous work^[16, 17]. The voxel size was a key parameter to reduce the uncertainties of the full-field measurements. The subset size parameter was used as (128, 96, 64, 32, 16, and 8 voxels). After measuring and fitting the displacement fields with tricubic functions, the components of the Green–Lagrange strain tensor were calculated with MatLab[®]. Here the error of measurement is not considered and only the mean value of displacement measurement is used for simplification.

275 3. Results

276 3.1. First case study: homogeneous material with uniaxial tension

This first case study is a plate under uniaxial tension, as introduced in Section 2.4.1, where an analytical solution is derived. A stretch λ is applied in the x direction at the right-side of the plate, and the constraints are applied as shown in Fig. 2. The parameters of the chemo-mechanically coupled constitutive model are reported in Table 2.

Table 2: Parameters of the chemo-mechanically coupled constitutive model						
Type	Description	Symbol	Values	Units		
	Universal gas constant	R	8.314	$\mathrm{kg} \cdot \mathrm{m}^2 \mathrm{s}^{-2} \mathrm{K}^{-1} \mathrm{mol}^{-1}$		
	Absolute temperature	θ	298	Κ		
Chemoelastic	Ambient pressure	p^*	0	kPa		
term	Initial fixed-charge density	c_0^F	[20, 40, 100, 200]	m meq/L		
	Water content	$arphi_0^w$	70	%		
	External bath salt osmolarity	\overline{C}^*	[10-2000]	mosM		
Hyperelastic	Shear modulus	μ	140	kPa		
	Bulk modulus	κ	1400	kPa		

280

281 3.1.1. Analytical solutions

Based on the analytical solutions of Section 2.4.1, the solutions of deformation and stress 282 are shown in Fig. 4. The deformation in Y-direction β is shown in Fig. 4(a) with different 283 external bath salt osmolarity \overline{c}^* , and it is seen that a larger β is produced with a lower \overline{c}^* 284 because of a larger swelling. The influences of \overline{c}^* on total Cauchy stress T_{11} is illustrated in 285 Fig. 4(b), in which a smaller external bath salt osmolarity produces a relatively smaller total 286 T_{11} , it can be explained that as the T_{11} has hyperelastic part and chemo part according to 287 Eqs.(3),(5), while the chemo part p will be increased with a lower \overline{c}^* as shown in Fig. 4(c), 288 therefore, a lower total Cauchy stress is produced. 289

290 3.1.2. Parameter identification

Since the material considered in the first case study is homogeneous, there is only one unknown parameter, μ . Thus, one single linear virtual displacement field based on Eq.(15) is considered, that is,

$$\boldsymbol{u}^* = \begin{bmatrix} 0.2x \\ -0.1y \\ -0.1z \end{bmatrix} \quad \boldsymbol{\varepsilon}^* = \begin{bmatrix} 0.2 \\ -0.1 \\ -0.1 \end{bmatrix}. \tag{48}$$



Figure 4: Analytical results under different external bath salt osmolarities and initial fixed-charge density $c_0^F = 200 \text{ meq/L}$ in uniaxial tension. (a) Stretch β variation in the Z-direction.(b) Cauchy stress T_{11} . (c) Osmotic pressure p.



Figure 5: The process of convergence for the shear modulus in the identification.

The shear modulus in solid part is well identified as shown in Table 3 regardless the 294 existence of chemo term and the convergence process of μ is shown in Fig. 5. Due to many 295 uncertainties in the chemo terms, we verify that the proposed VFM is immune to chemoelas-296 tic coupling effects by simulating three different external bath salt osmolarity environments 297 and one different initial fixed-charge density in the tissue. The identified results of shear 298 modulus $\tilde{\mu}$ are always equal to the exact shear modulus $\hat{\mu}$, as reported in Table 3. Moreover, 299 we also test the identification results under different initial guess values and target(exact) 300 values, the results are provided in Table 4, in which the initial values of shear modulus μ 301 changes from 1000 Pa to 10000 Pa and the exact value of shear modulus $\hat{\mu}$ changes from 302 130000 Pa to 150000 Pa, it is shown that the identified results maintain a good accuracy 303 with relative error less than 1%. 304

Based on the noise analysis method in Section 2.3.4, we tested the influence of noise on the identification, as reported in Table 5. It is shown that the error remains less than 3%, with a relatively low noise level ($\sigma_u \leq 5 \times 10^{-4}$) but gradually increases with the increase of noise level.

Туре	Symbol	Value			
External bath salt osmolarity	\overline{c}^*	50 mosM	100 mosM	500 mosM	500 mosM
Initial fixed-charge density	c_0^F	40 meq/L	40 meq/L	40 meq/L	60 meq/L
Tensile stretch	λ	1.2	1.2	1.2	1.2
Initial guess shear modulus	μ	1000 Pa	1000 Pa	1000 Pa	1000 Pa
Identified modulus	$\widetilde{\mu}$	140000 Pa	140000Pa	140000 Pa	140000 Pa
Exact modulus	$\widehat{\mu}$	140000Pa	140000Pa	140000Pa	140000Pa

Table 3: Identified shear modulus in the first case study

308

Identified $\widetilde{\mu}(\mathrm{Pa})$		Exact values $\widehat{\mu}(Pa)$					
and relative e	$\operatorname{error}(\%)$	130000	135000	140000	145000	150000	
	1000	129533	135192	140000	144500	150123	
	1000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	2000	129533	135192	140000	144500	150123	
	2000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	3000	129533	135192	140000	144500	150123	
	3000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	4000	129533	135192	140000	144500	150123	
Initial guoss	4000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
$\mu(P_2)$	5000	129533	135192	140000	144500	150123	
$\mu(\mathbf{I} \mathbf{a})$		(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	6000	129533	135192	140000	144500	150123	
		(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	7000	129533	135192	140000	144500	150123	
	1000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	8000	129533	135192	140000	144500	150123	
	8000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	0000	129533	135192	140000	144500	150123	
	9000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	
	10000	129533	135192	140000	144500	150123	
	10000	(-0.36)	(0.14)	(0)	(-0.34)	(0.08)	

Table 4: The identification results of different exact values and initial guesses

Table 5: The relative error of identified results under different levels of measurement noises

Standard deviation σ_u	1×10^{-4}	3×10^{-4}	5×10^{-4}	7×10^{-4}	1×10^{-3}
Exact $\widehat{\mu}(Pa)$	140000	140000	140000	140000	140000
Mean identified $\bar{\mu}(Pa)$	140349	139443	136382	131183	119993
error	-0.25%	0.4%	2.58%	6.30%	14.29%

309 3.2. Second case study: multilayer material under biaxial tension

The second case study relies on the chemomechanically coupled constitutive model that we implemented within the Abaqus/Standard commercial FE software by means of the user subroutine UMAT, based on Eqs.(3),(4),(5),(45) and (47).

The geometry of this second case study is shown in Fig. 6 to simulate the layered structure 313 of arteries. The size of the three-layers plate is $0.08 \times 0.04 \times 0.03$ (i.e. l=0.08, m=0.04314 and n=0.03) and the geometry is discretized with a fine mesh of 768 cubic elements. The 315 boundary conditions are the same as the first case study in Fig. 2, with $u_x|_{x=0} = u_y|_{y=0} =$ 316 $u_z|_{z=0} = 0$. Biaxial displacement loading is applied to the X and Y directions, including 317 1.30 stretch in X direction and 1.05 stretch in Y direction. Accordingly, the hyperelasitic 318 constitutive model is used for arterial tissues in this example. The values for the constitutive 319 parameters are those reported in Tables 6. Note here we use a large bulk modulus to make 320 the material nearly incompressible. 321



Figure 6: The geometry and displacement loading for the three-layers plate

322 3.2.1. FEM solutions

Firstly, we verify the accuracy of the FE solutions for direct analysis. As it is not possible to find the analytical solution for the three-layers examples, so we use the FE model introduced in Section 2.4.2 to solve the previous first case example with different bath osmolarity values. As it is shown in Fig. 7, it is found that there is a very good agreement between FE and analytical solutions.

328 3.2.2. Parameter identification

For the second case study, we can still consider the constant virtual strain field of Eq.(48), which is represented in Fig. 8(a). However, two other heterogeneous virtual fields are required to be introduced to ensure independence among the three VFM equations. Based on Eq.(26), two other virtual fields can be determined as illustrated in Fig. 8(b) and 8(c) where parameters setting details can also be found.

Table 6: Parameters of the chemo-mechanically coupled constitutive model for second case study						
Type	Description		Symbol	Values	Units	
	Universal g	as constant	R	8.314	$\mathrm{kg} \cdot \mathrm{m}^2 \mathrm{s}^{-2} \mathrm{K}^{-1} \mathrm{mol}^{-1}$	
	Absolute te	mperature	heta	298	Κ	
Chemoelastic	Ambient pr	essure	p^*	0	kPa	
term	Initial fixed-charge density		c_0^F	40	m meq/L	
	Water content		φ_0^w	70	%	
	External bath salt osmolarity		\overline{C}^*	[20, 100, 500, 1000]	mosM	
	Material 1	Shear modulus	μ_1	100	kPa	
		Bulk modulus	κ	6000	kPa	
Hyperelastic	Matarial 9	Shear modulus	μ_2	200	kPa	
term	Material 2	Bulk modulus	κ	6000	kPa	
	Matarial 2	Shear modulus	μ_3	300	kPa	
	material 5	Bulk modulus	κ	6000	kPa	



Figure 7: Verification of the FE model against the analytical model on the results of Cauchy stress



(c)

Figure 8: The three virtual displacement fields used in the second case study. (a) Constant strain field. (b) Heterogeneous virtual strain with k = 1 and $\eta = -0.01$. (c) Heterogeneous virtual strain with k = 2 and $\eta = 0.001$.



Figure 9: The identification process in the second case study. (a) Convergence plot of the novel VFM approach for each layer parameter. (b) Cost function f descent.

$\overline{\mathrm{C}}^*$	Туре	Material 1	Material 2	Material 3
	Initial value of $\boldsymbol{\mu}(\mathrm{Pa})$	150000	150000	150000
20 mosM	Identified $\widetilde{\boldsymbol{\mu}}(\mathrm{Pa})$	100000.95	199999.81	299998.34
100 mosM	Identified $\widetilde{\boldsymbol{\mu}}(\mathrm{Pa})$	99999.02	200001.77	300003.04
500 mosM	Identified $\widetilde{\boldsymbol{\mu}}(\operatorname{Pa})$	100001.24	199998.37	299996.26
1000 mosM	Identified $\widetilde{\boldsymbol{\mu}}(\mathrm{Pa})$	99997.62	200001.04	300006.52
	Exact $\widehat{\boldsymbol{\mu}}(\operatorname{Pa})$	100000	200000	300000
	Average relative error $(\%)$	1.388×10^{-3}	5.788×10^{-4}	1.247×10^{-3}

Table 7: The identified results for different layers under different external bath osmolarities

The identification for each layer is verified by considering with different chemoelastic terms. As reported in Table 7, the same values of μ are identified even if we assume different values for the external salt osmolarity. This demonstrates that the proposed model is effective in shear modulus identification without knowing the chemoelatic terms. The process of minimizing the cost function $f(\mu_1, \mu_2, \mu_3)$ is provided in Fig. 9.

The influence of noise on the identification is reported in Table 8. The material parameters are well recovered with a low noise level ($\sigma_u < 5 \times 10^{-4}$) and it is also shown that the error of identified results is increased with the increase of noise level. However, when the noise level reaches a higher level ($\sigma_u = 1 \times 10^{-3}$), the results deviate significantly from the reference values.

σ	Mean identified values $\bar{\mu}(\text{Pa})$							
\circ_u	$ar{\mu}_1$	error(%)	$ar{\mu}_2$	error(%)	$ar{\mu}_3$	error(%)		
1×10^{-4}	99622.35	0.38	200185.09	-0.09	298801.27	0.40		
3×10^{-4}	97340.56	2.66	197091.80	1.45	292038.95	2.65		
5×10^{-4}	92950.14	7.05	189106.28	5.45	279275.88	6.91		
7×10^{-4}	86117.79	13.88	175407.71	12.30	259909.74	13.36		
1×10^{-3}	63550.56	36.45	142574.59	28.71	209265.30	30.24		
Exact values	100000.00		200000.00		300000.00			

Table 8: The relative error of identified results in three layers under different levels of measurement noises

3.4. 3.3. Third case study: uniaxial tensile test of descending thoracic aortas

The third example consider the parameter identification based on the measurements in a uniaxial tensile test of descending thoracic aortas described in Section 2.5. Three samples with the same size were tested. The thickness ratio of each layer was set as 1:8:1 (adventitia:media:intima), according to the size of experimental sample^[17]. The geometry for the sample is shown in Fig. 10. The measurement for the average Green-Lagrange strains with the standard deviation(std) and the corresponding applied force in the different deformed configurations for each sample are reported in Tables 9, 10, and 11.



Figure 10: Geometry and boundary condition for an aortic sample immersed in a PG solution and subjected to a uniaxial tensile test

Tupo	Loading	Measured	F (%) (atd)	F (%) (atd)	F(%) (atd)	
Type	step	$\operatorname{stress}(MPa)$	$E_{xx}(70)$ (std)	$E_{yy}(70)$ (std)	$E_{zz}(70)$ (Std)	
	3rd	0.07	-1.510 (0.42)	4.352(0.26)	0.145 (1.18)	
Clobal	5th	0.1	-2.124(0.52)	$7.090\ (0.37)$	0.108(2.32)	
Global	$7 \mathrm{th}$	0.13	-2.596(0.71)	$9.554\ (0.51)$	0.170(3.56)	
	9th	0.17	-3.125(0.76)	11.847(0.60)	0.353(4.76)	
	3rd	0.07	-1.558(0.42)	4.381(0.35)	-2.271 (4.57)	
Intimo	5th	0.1	-2.136(0.44)	7.157(0.49)	-3.528(6.99)	
muma	$7 \mathrm{th}$	0.13	-2.600(0.65)	$9.603\ (0.67)$	-4.768(8.89)	
	9th	0.17	-3.129(0.74)	11.949(0.79)	-5.566(10.18)	
	3rd	0.07	-1.489(0.39)	4.376(0.23)	1.003(0.34)	
Madia	5th	0.1	-2.127(0.54)	7.112(0.32)	1.603(0.63)	
media	$7 \mathrm{th}$	0.13	-2.582(0.68)	$9.570\ (0.45)$	2.263(1.60)	
	9th	0.17	-3.114(0.72)	11.857 (0.52)	3.007(2.67)	
	3rd	0.07	-1.434(0.67)	4.273(0.35)	-2.120 (3.15)	
Adventitie	5th	0.1	-2.043(0.85)	6.958(0.46)	-3.525 (4.96)	
Auventitia	$7 \mathrm{th}$	0.13	-2.509(1.05)	$9.407 \ (0.65)$	-4.795(6.73)	
	$9 \mathrm{th}$	0.17	-3.020(1.24)	11.630(0.79)	-6.137(7.91)	

Table 9: The average Green-Lagrange strains from the experiment for Aorta1

Tupo	Loading	Measured	$F_{(\infty)}$ (gtd)	F (%) (atd)	$F(\mathcal{O}_{a})$ (atd)
туре	step	$\operatorname{stress}(MPa)$	$E_{xx}(70)$ (stu)	$E_{yy}(70)$ (stu)	$E_{zz}(70)$ (stu)
	3rd	0.06	-0.628(2.26)	3.572(1.80)	$0.493\ (0.38)$
Clobal	5th	0.1	-1.135(3.60)	6.412(2.92)	$0.627 \ (0.87)$
GIUDAI	$7 \mathrm{th}$	0.13	-1.475(4.93)	9.210(4.10)	$0.523\ (1.63)$
	$9 \mathrm{th}$	0.17	-1.982(6.02)	11.708(5.09)	0.385~(2.53)
	3rd	0.06	-0.370(2.93)	4.134(2.01)	-2.470 (10.00)
Intimo	5th	0.1	-0.717(4.68)	7.579(3.22)	-3.383(14.95)
muma	$7 \mathrm{th}$	0.13	-1.105(6.36)	10.846(4.38)	-4.156(18.03)
	$9 \mathrm{th}$	0.17	-1.688(8.12)	13.758(5.22)	-5.080(19.37)
	3rd	0.06	-0.733(2.55)	3.658(1.88)	0.645(1.60)
Modia	5th	0.1	-1.309(3.94)	6.555(2.96)	$0.919\ (1.50)$
meula	$7 \mathrm{th}$	0.13	-1.726(5.18)	9.555(3.94)	1.152(1.22)
	$9 \mathrm{th}$	0.17	-2.225(6.19)	12.021(5.15)	$1.627\ (1.19)$
Adventitie	3rd	0.06	-1.233(2.91)	1.912(1.63)	-2.372(5.29)
	5th	0.1	-1.511(4.12)	3.393(2.70)	-3.950(7.58)
Auventitia	$7 \mathrm{th}$	0.13	-1.775(4.62)	4.830(4.07)	-4.421 (8.65)
	9th	0.17	-2.076(5.25)	5.926(5.35)	-4.922(11.71)

Table 10: The average Green-Lagrange strains from the experiment for Aorta2

Table 11: The average Green-Lagrange strains from the experiment for Aorta3

Type	Loading	Measured	F (%) (std)	F (%) (std)	F (%) (std)
туре	step	stress(MPa)	$E_{xx}(70)$ (stu)	$E_{yy}(70)$ (stu)	$E_{zz}(70)$ (Stu)
	3rd	0.08	-1.315(1.28)	4.940(0.62)	-0.269 (0.71)
Clobal	5th	0.12	-2.303(1.92)	8.377(1.03)	-0.708(1.66)
Giobai	$7 \mathrm{th}$	0.17	-2.923(2.54)	11.174(1.24)	-1.020(2.76)
	$9 \mathrm{th}$	0.21	-3.368(3.08)	14.004(1.60)	-1.124(3.80)
	3rd	0.08	-1.506 (1.13)	4.929(0.58)	-1.655(3.22)
Intimo	5th	0.12	-2.642(1.82)	8.405(0.89)	-3.531(5.50)
III0IIIIa	$7 \mathrm{th}$	0.17	-3.432(2.45)	11.206(1.02)	-5.844(8.40)
	$9 \mathrm{th}$	0.21	-3.978(2.94)	14.030(1.34)	-7.480(10.18)
	3rd	0.08	-1.235(1.55)	4.909(0.69)	0.156(0.71)
Modia	5th	0.12	-2.210(2.17)	8.342(1.06)	$0.279\ (0.70)$
Media	$7 \mathrm{th}$	0.17	-2.836(2.88)	11.166(1.26)	$0.687 \ (1.08)$
	$9 \mathrm{th}$	0.21	-3.287(3.36)	14.069(1.57)	1.472(1.40)
	3rd	0.08	-1.367(1.47)	4.860(1.01)	-2.878(4.84)
Adventitio	5th	0.12	-2.213(2.13)	8.097(1.66)	-5.542(7.43)
Auvennua	$7 \mathrm{th}$	0.17	-2.590(2.60)	10.910(2.24)	-8.711 (11.56)
	$9 \mathrm{th}$	0.21	-3.072(3.25)	13.703(2.71)	-11.764 (13.49)

Initia	Initial guesses $\boldsymbol{\mu}(MPa)$			Identified values $\tilde{\boldsymbol{\mu}}(MPa)$		
Intima	Media	Adventitia	-	Intima	Media	Adventitia
0.1	0.1	0.1	-	0.4746	1.0040	0.4831
0.2	0.2	0.2	-	0.4746	1.0040	0.4831
0.3	0.3	0.3	-	0.4746	1.0040	0.4831
0.4	0.4	0.4	-	0.4746	1.0040	0.4831
1.0	1.0	1.0	-	0.4746	1.0040	0.4831
2.0	2.0	2.0	-	0.4746	1.0040	0.4831
3.0	3.0	3.0	-	0.4746	1.0040	0.4831
10.0	10.0	10.0	-	0.4746	1.0040	0.4831
1.0	2.0	3.0	_	0.4746	1.0040	0.4831
3.0	2.0	1.0	-	0.4746	1.0040	0.4831

Table 12: The identification results for experimental measurements from different initial guesses

Firstly, we investigate the influence of initial guess on the identification results as shown 352 in Table 12, in which one group of experimental measurements in Table 9 is used, it is shown 353 that the identified results for different layers of arteries are the same for ten different group of 354 initial guesses. Then, the identification results for three samples using the proposed VFM are 355 shown in Figs. 11(a), 11(b), and 11(c), respectively. Despite the chemoelastic effects going 356 on in these samples, the identified shear moduli show only marginal variations with the 357 applied force, indicating that the identification method is able to eliminate the non-purely 358 elastic effects. Indeed, it is assumed that these effects only affect the hydrostatic pressure 359 to which the VFM is immune. This assumption permitted the separate identification of the 360 shear modulus of each layer, as shown in Fig. 12. The largest shear moduli are obtained in 361 the media layer, with values of approximately 1 MPa. The intima and adventitia appeared 362 to be at least half as stiff. Although the obtained absolute values may be specific to the 363 conditions used for the purpose of this study (immersion in PG to induce osmotic effects), 364 the relative values between each layer indicate a significant gradient of elastic properties 365 across the thickness. 366

³⁶⁷ 4. Conclusion and discussion

The mechanical behavior of soft biological tissues can be significantly affected by the chemical potentials of interstitial fluids, resulting in obvious chemomechanical coupling phenomena. And chemoelastic constitutive models are widely used for soft biological tissues. However, to the best of our knowledge, there have been very few studies related to parameter identification of these models using VFM combined with optical full-field measurement techniques.

In this paper, VFM was applied for the parameter identification of biphasic chemoelastic models using full-field measurements. The proposed approach avoids iterative resolutions



Figure 11: Identified shear moduli of tested samples with average strains across each layer and the entire aortic wall. (a) Aorta1, (b)Aorta2,(c)Aorta3.



Figure 12: Identified shear moduli (mean value and deviation bar) for each layer of the tested samples

of direct problems with complex nonlinear chemomechanical interaction relationships. An advantage of the proposed approach is that it can extract hyperelastic parameters without knowing the chemoelastic parameters of the model simply by designing a special virtual field. Therefore, the proposed approach will be useful for further parameter identifications in soft biological tissues.

The proposed approach was assessed to identify the layer-specific stiffness properties of 381 arteries based on strain fields. The identification results based on numerical simulations 382 showed that the proposed model is immune to chemoelastic effects. Moreover, the shear 383 modulus of the media of porcine descending thoracic aortas immersed in an 80% PG so-384 lution was identified with values of approximately 1 MPa, however, the identified shear 385 modulus is smaller than the instantaneous shear modulus, which was identified to be ap-386 proximately 3 MPa in a previous work^[17]. Indeed, the instantaneous shear modulus was 387 found by assuming incompressibility of the solid part in the biphasic hyperelastic model, 388 whereas identification with the VFM did not require such an assumption. Another interest-389 ing finding is that the layer-specific shear moduli can be identified simultaneously with the 390 VFM. The shear modulus of the media layer is nearly twice as stiff as that of the intima and 391 adventitia. The ratios between interlayer shear moduli of arteries are similar to those ob-392 tained in the work of Peña et al.^[43], in which they were approximately 1:2:1. Therefore, the 393 proposed approach was proved to provide an effective identification of interlayer gradients of 394 shear moduli despite the chemoelastic effects. A potential application of the proposed VFM 395 approach is the multiscale mechanical characterization of biological tissues since both the 396 effective material parameters at the macroscopic scale (as in Example 1) and the multilayer 397

or heterogeneous material parameters at the mesoscopic scale (as in Examples 2 and 3) can be well identified, showing effective multiscale identification for soft biological tissues even in the presence of chemoelastic effects.

A main limitation in this study is that the VFM was only tested with the neo-Hookean 401 model. Due to the obvious anisotropic behavior of a ortic wall, future work will extend 402 the approach to anisotropic hyperelastic models, such as the Holzapfel^[44] and MA-HGO^[45] 403 models which are used extensively to model collagen fibre reinforced biological materials, 404 and the use of neo-Hookean model could produce a systematic variation of the apparent 405 shear modulus with increasing strain. Another limitation of the work is that we ignore the 406 stiffening effect of soft tissues for the sake of simplicity, a more complex VFM for nonlinear 407 problems is required to consider this case. 408

In summary, we have presented a new VFM approach for parameter identification that is well suited for chemoelastic constitutive relationships in soft biological tissues. The identification of layer-specific elastic parameters can be applied even in the presence of complex chemoelastic effects. Important developments will extend the approach to multiscale constitutive models, and anisotropic effects of soft tissues.

⁴¹⁴ Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

417 Acknowledgements

The research leading to this paper was funded by the NSFC Grants [12072063, 11972109], the Fundamental Research Funds for the Central Universities [DUT21YG129], the Grants of State Key Laboratory of Structural Analysis for Industrial Equipment [S22403, GZ21104], and the Alexander von Humboldt Foundation [1217594].

422 Data availability statement

⁴²³ The data discussed in this work are available on request.

424 References

- [1] Y. Lanir, Osmotic swelling and residual stress in cardiovascular tissues, Journal of Biomechanics 45 (2012) 780-789. doi:10.1016/j.jbiomech.2011.11.018.
- [2] A. E. Ehret, K. Bircher, A. Stracuzzi, V. Marina, M. Zündel, E. Mazza, Inverse poroelasticity as a fundamental mechanism in biomechanics and mechanobiology, Nature Communications & (2017) 1002, doi:10.1028/c41167.017.00801.2
- 429 Communications 8 (2017) 1002. doi:10.1038/s41467-017-00801-3.

- [3] I. S. Kovach, The importance of polysaccharide configurational entropy in determining the osmotic swelling pressure of concentrated proteoglycan solution and the bulk compressive modulus of articular cartilage, Biophysical Chemistry 53 (1995) 181–187. doi:10.1016/0301-4622(94)00100-X.
- [4] Y. Lanir, Biorheology and fluid flux in swelling tissues. I. Bicomponent theory for
 small deformations, including concentration effects, Biorheology 24 (1987) 173–187.
 doi:10.3233/BIR-1987-24210.
- [5] A. Eringen, J. D. Ingram, A continuum theory of chemically reacting media—I, International Journal of Engineering Science 3 (1965) 197–212. doi:10.1016/0020-7225(65)
 90044-3.
- [6] J. D. Ingram, A. Eringen, A continuum theory of chemically reacting media—II Constitutive equations of reacting fluid mixtures, International Journal of Engineering Science
 5 (1967) 289–322. doi:10.1016/0020-7225(67)90040-7.
- [7] V. C. Mow, S. C. Kuei, W. M. Lai, C. G. Armstrong, Biphasic creep and stress relaxation
 of articular cartilage in compression: theory and experiments, Journal of Biomechanical
 Engineering 102 (1980) 73–84. doi:10.1115/1.3138202.
- [8] W. M. Lai, J. S. Hou, V. C. Mow, A triphasic theory for the swelling and deformation
 behaviors of articular cartilage, Journal of Biomechanical Engineering 113 (1991) 245–
 258. doi:10.1115/1.2894880.
- [9] G. A. Ateshian, N. O. Chahine, I. M. Basalo, C. T. Hung, The correspondence between equilibrium biphasic and triphasic material properties in mixture models of articular cartilage, Journal of Biomechanics 37 (2004) 391–400. doi:10.1016/S0021-9290(03) 00252-5.
- [10] G. A. Ateshian, B. J. Ellis, J. A. Weiss, Equivalence between short-time biphasic and
 incompressible elastic material responses, Journal of Biomechanical Engineering 129
 (2007) 405-412. doi:10.1115/1.2720918.
- [11] G. A. Ateshian, V. Rajan, N. O. Chahine, C. E. Canal, C. T. Hung, Modeling the matrix of articular cartilage using a continuous fiber angular distribution predicts many observed phenomena, Journal of Biomechanical Engineering 131 (2009) 061003. doi:10.
 1115/1.3118773.
- [12] W. Wilson, C. C. van Donkelaar, J. M. Huyghe, A comparison between mechanoelectrochemical and biphasic swelling theories for soft hydrated tissues, Journal of
 Biomechanical Engineering 127 (2005) 158–165. doi:10.1115/1.1835361.
- [13] J. S. Wayne, S. L.-Y. Woo, M. K. Kwan, Application of the u-p finite element method
 to the study of articular cartilage, Journal of Biomechanical Engineering 113 (1991)
 397-403. doi:10.1115/1.2895418.

- [14] K. Kandil, F. Zaïri, A. Derrouiche, T. Messager, F. Zaïri, Interlamellar-induced timedependent response of intervertebral disc annulus: A microstructure-based chemoviscoelastic model, Acta Biomaterialia 100 (2019) 75–91. doi:10.1016/j.actbio.2019.
 10.005.
- E. U. Azeloglu, M. B. Albro, V. A. Thimmappa, G. A. Ateshian, K. D. Costa, Heterogeneous transmural proteoglycan distribution provides a mechanism for regulating residual stresses in the aorta, American Journal of Physiology-Heart and Circulatory Physiology 294 (2008) H1197–H1205. doi:10.1152/ajpheart.01027.2007.
- ⁴⁷⁴ [16] V. A. Acosta Santamaría, M. F. García, J. Molimard, S. Avril, Three-dimensional
 ⁴⁷⁵ full-field strain measurements across a whole porcine aorta subjected to tensile loading
 ⁴⁷⁶ using optical coherence tomography-digital volume correlation, Frontiers in Mechanical
 ⁴⁷⁷ Engineering 4 (2018) 3. doi:10.3389/fmech.2018.00003.
- [17] V. A. Acosta Santamaría, M. F. García, J. Molimard, S. Avril, Characterization of chemoelastic effects in arteries using digital volume correlation and optical coherence tomography, Acta Biomaterialia 102 (2020) 127–137. doi:10.1016/j.actbio.2019.11.
 049.
- [18] S. Avril, S. Evans (Eds.), Material Parameter Identification and Inverse Problems in Soft
 Tissue Biomechanics, volume 573 of *CISM International Centre for Mechanical Sciences*,
 Springer International Publishing, Cham, 2017. doi:10.1007/978-3-319-45071-1.
- [19] S. Avril, M. Bonnet, A.-S. Bretelle, M. Grédiac, F. Hild, P. Ienny, F. Latourte,
 D. Lemosse, S. Pagano, E. Pagnacco, F. Pierron, Overview of identification methods of
 mechanical parameters based on full-field measurements, Experimental Mechanics 48
 (2008) 381–402. doi:10.1007/s11340-008-9148-y.
- ⁴⁸⁹ [20] F. Pierron, M. Grédiac, The Virtual Fields Method, Springer New York, New York, NY, 2012. doi:10.1007/978-1-4614-1824-5.
- ⁴⁹¹ [21] M. Grédiac, Principe des travaux virtuels et identification. (Principle of virtual work
 ⁴⁹² and identification), Comptes Rendus de l'Académie des Sciences. Série II 309 (1989).
- ⁴⁹³ [22] M. Grédiac, E. Toussaint, F. Pierron, Special virtual fields for the direct determination of material parameters with the virtual fields method. 1—Principle and definition, International Journal of Solids and Structures 39 (2002) 2691–2705.
 ⁴⁹⁶ doi:10.1016/S0020-7683(02)00127-0.
- M. Grédiac, E. Toussaint, F. Pierron, Special virtual fields for the direct determination of material parameters with the virtual fields method. 2—Application to inplane properties, International Journal of Solids and Structures 39 (2002) 2707–2730. doi:10.1016/S0020-7683(02)00128-2.

- [24] M. Grédiac, E. Toussaint, F. Pierron, Special virtual fields for the direct determination of material parameters with the virtual fields method. 3. Application to the bending
 rigidities of anisotropic plates, International Journal of Solids and Structures 40 (2003)
 2401–2419. doi:10.1016/S0020-7683(03)00030-1.
- [25] Y. Mei, J. Liu, X. Guo, B. Zimmerman, T. D. Nguyen, S. Avril, General finite-element
 framework of the virtual fields method in nonlinear elasticity, Journal of Elasticity
 (2021). doi:10.1007/s10659-021-09842-8.
- Y. Mei, J. Deng, X. Guo, S. Goenezen, S. Avril, Introducing regularization into the virtual fields method (VFM) to identify nonhomogeneous elastic property distributions, Computational Mechanics 67 (2021) 1581–1599. doi:10.1007/s00466-021-02007-3.
- [27] M. Grédiac, F. Pierron, S. Avril, E. Toussaint, The virtual fields method for extracting
 constitutive parameters from full-field measurements: a review, Strain 42 (2008) 233–
 253. doi:10.1111/j.1475-1305.2006.tb01504.x.
- [28] S. Avril, M. Grédiac, F. Pierron, Sensitivity of the virtual fields method to noisy data,
 Computational Mechanics 34 (2004) 439–452. doi:10.1007/s00466-004-0589-6.
- [29] S. Avril, F. Pierron, General framework for the identification of constitutive parameters
 from full-field measurements in linear elasticity, International Journal of Solids and
 Structures 44 (2007) 4978–5002. doi:10.1016/j.ijsolstr.2006.12.018.
- [30] S. Avril, P. Badel, A. Duprey, Anisotropic and hyperelastic identification of in vitro
 human arteries from full-field optical measurements, Journal of Biomechanics 43 (2010)
 2978-2985. doi:10.1016/j.jbiomech.2010.07.004.
- J.-H. Kim, S. Avril, A. Duprey, J.-P. Favre, Experimental characterization of rupture in human aortic aneurysms using a full-field measurement technique, Biomechanics and Modeling in Mechanobiology 11 (2012) 841–853. doi:10.1007/s10237-011-0356-5.
- [32] M. R. Bersi, C. Bellini, P. Di Achille, J. D. Humphrey, K. Genovese, S. Avril, Novel methodology for characterizing regional variations in the material properties of murine aortas, Journal of Biomechanical Engineering 138 (2016) 071005. doi:10.1115/1.
 4033674.
- [33] N. O. Chahine, F. H. Chen, C. T. Hung, G. A. Ateshian, Direct measurement of osmotic
 pressure of glycosaminoglycan solutions by membrane osmometry at room temperature,
 Biophysical Journal 89 (2005) 1543–1550. doi:10.1529/biophysj.104.057315.
- [34] F. G. Donnan, The theory of membrane equilibria., Chemical Reviews 1 (1924) 73–90.
 doi:10.1021/cr60001a003.
- ⁵³⁴ [35] J. Overbeek, The donnan equilibrium, Progress in Biophysics and Biophysical Chem-⁵³⁵ istry 6 (1956) 57–84. doi:10.1016/S0096-4174(18)30104-5.

- ⁵³⁶ [36] A. F. Bower, Applied Mechanics of Solids, CRC Press, 2009. doi:10.1201/ 9781439802489.
- [37] Y. Mei, S. Avril, On improving the accuracy of nonhomogeneous shear modulus identification in incompressible elasticity using the virtual fields method, International Journal of Solids and Structures 178-179 (2019) 136-144. doi:10.1016/j.ijsolstr.2019.06.
 025.
- [38] J. C. Lagarias, J. A. Reeds, M. H. Wright, P. E. Wright, Convergence Properties of the
 Nelder-Mead Simplex Method in Low Dimensions, SIAM Journal on Optimization 9
 (1998) 112-147. doi:10.1137/S1052623496303470.
- [39] P. Thakolkaran, A. Joshi, Y. Zheng, M. Flaschel, L. De Lorenzis, S. Kumar, NNEUCLID: Deep-learning hyperelasticity without stress data, Journal of the Mechanics
 and Physics of Solids 169 (2022) 105076. doi:10.1016/j.jmps.2022.105076.
- [40] M. Flaschel, S. Kumar, L. De Lorenzis, Unsupervised discovery of interpretable hypere lastic constitutive laws, Computer Methods in Applied Mechanics and Engineering 381
 (2021) 113852. doi:10.1016/j.cma.2021.113852.
- [41] H. Yiqian, Y. Haitian, Solving inverse couple-stress problems via an element-free
 Galerkin (EFG) method and Gauss-Newton algorithm, Finite Elements in Analysis
 and Design 46 (2010) 257-264. doi:10.1016/j.finel.2009.09.009.
- ⁵⁵⁴ [42] T. Waffenschmidt, C. Polindara, A. Menzel, S. Blanco, A gradient-enhanced largedeformation continuum damage model for fibre-reinforced materials, Computer Methods in Applied Mechanics and Engineering 268 (2014) 801–842. doi:10.1016/j.cma.2013.
 ⁵⁵⁷ 10.013.
- [43] J. A. Peña, M. A. Martínez, E. Peña, Layer-specific residual deformations and uniaxial and biaxial mechanical properties of thoracic porcine aorta, Journal of the Mechanical Behavior of Biomedical Materials 50 (2015) 55–69. doi:10.1016/j.jmbbm.2015.05.024.
- [44] G. A. Holzapfel, T. C. Gasser, R. W. Ogden, A new constitutive framework for arterial
 wall mechanics and a comparative study of material models, in: S. C. Cowin, J. D.
 Humphrey (Eds.), Cardiovascular Soft Tissue Mechanics, Kluwer Academic Publishers,
 Dordrecht, 2004, pp. 1–48. doi:10.1007/0-306-48389-0_1.
- [45] D. Nolan, A. Gower, M. Destrade, R. Ogden, J. McGarry, A robust anisotropic hyper elastic formulation for the modelling of soft tissue, Journal of the Mechanical Behavior
 of Biomedical Materials 39 (2014) 48–60. doi:10.1016/j.jmbbm.2014.06.016.