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## Cancer and immunotherapy: A role for microbiota composition

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## Abstract (127 words)

Human microbiota plays a key role in preserving homeostasis; therefore, alteration in its composition is associated with susceptibility to various diseases. Recent findings suggest that gut microbiota may influence response to cancer treatment, especially immune checkpoint blockers (ICB). The development of ICB has changed cancer patients' outcomes and has allowed sustained recovery. Unfortunately, some patients do not respond to ICB and microbiota may be a promising new biomarker to identify patients that will have benefit from ICB. This review presents relationship between microbiome composition or microbiota-derived metabolites and response to ICB or immune-related adverse events. Furthermore, we will present different strategies to modulate microbiota composition in patients in order to enhance ICB efficacy or dampen their toxicities which could lead to the emergence of interesting complementary treatments.

## Introduction

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The human microbiota is composed of around  $3 \times 10^{13}$  microorganisms (bacteria, viruses, fungi, and protozoa) in the epithelial barrier surfaces and has a commensal relationship with the host. Individual's microbes composition changes over time due to host genetics [1], diet [2], medication [3], lifestyle, geographical localization [4] or age, leading to a significant interindividual variation. Gut microbiome has several physiologic functions and participates in compounds hydrolysis [5], vitamin production [6], gut integrity and control of pathogen colonization [7], protection against systemic infections [8] and immune system development [9]. It is now well established that intestinal dysbiosis, defined as an imbalance of the bacterial ecosystem, and digestive and even extra-digestive diseases are linked [10]. Commensal microbiota contributes to individual's immunity, affecting overall health. Disruption in the homeostasis of microbial communities induces local chronic inflammation and activation of the immune system, both mechanisms that could play a role in tumorigenesis [11–13]. Several evidences support the role of some bacteria, as viruses, in carcinogenesis and cellular dysplasia. Salmonella typhimurium might be correlated with the onset of gallbladder cancer [11], Helicobacter pylori influences the pathogenesis of gastric cancer [14] through chronic inflammation and influence over the local microenvironment to promote carcinogenesis. Growing evidence shows that gut microbial dysbiosis contributes to the development of colorectal cancer (CRC). A study identifies different compositions between tumoral tissue and neighboring tumor healthy tissue. Firmicutes, Fusobacteria, Lactococcus and Fusobacterium were found in abundance in CRC tissue compared to healthy one while Pseudomonas and Escherichia-Shigella were under-represented in tumoral tissue compared to healthy one [15]. As evidences suggest that microbiota composition could confers susceptibility to certain cancers, it may also influence cancer outcomes [16]. Over the years, gut microbiota composition has emerged as a new target to shape efficacy and toxicity of cancer therapy [17] and nowadays, data support, even in patients, its role as a predictive biomarker of response to chemotherapy or immunotherapy or for susceptibility to treatment-related toxicities [18–26]. Consequently, there is a growing interest in targeting these microbes to improve therapeutic anticancer strategies.

## Gut microbiota modulation of ICB treatment efficacy and toxicity

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In the last decade, ICB have changed the therapeutic strategies of several solid cancers by enhancing antitumor immune response [27]. These FDA- and EMA-approved monoclonal antibodies target co-inhibitory molecules to block pathways of negative regulation of the immune system: Cytotoxic T-lymphocyte-associated protein-4 (CTLA-4), programmed-cell death protein-1 receptor (PD-1) or its ligand (PD-L1) down-regulate the immune response when its activation become harmful to the host. Tumor cells and immune cells in the tumor microenvironment overexpress these molecules to escape antitumor immune surveillance [28]. Blocking these receptors or ligands can restore a patient's immune anti-tumoral response. ICBs have had a major impact on cancer treatment, but a non-negligible proportion of patients do not respond to these immunotherapies, some cancers show primary or secondary resistance to ICBs and response rate differs between patients and cancer localizations. As gut microorganisms play a role in the balanced ecosystem and in shaping normal and pathological immune response, contributing to carcinogenesis, microbiome might play a role in anti-tumor immunosurveillance and has then emerged as a potential biomarker for response to ICBs. Some evidences indicate that it could be part of the underlying reasons for non-responsiveness, as particular taxa have been identified in ICB responders' gut microbiota or stools in preclinical models [18,29,30] and patients [19,20,22–26].

#### • Evidence in preclinical mouse models

One of the first study demonstrating a relationship between gut microbiota and response to immunotherapy was conducted in mice injected subcutaneously with cancer cells and treated

with CpG-oligonucleotide, a ligand of Toll-like receptor 9 and an antibody directed against the immunosuppressive interleukin-10 receptor. This association of immunotherapies induces tumor necrosis factor (TNF) secretion by tumor-associated myeloid cells followed by a CD8+ T cell response leading to tumor hemorrhagic necrosis. Antibiotic (ATB)-treated or germ-free mice showed an impaired production of proinflammatory cytokines by immune cells. Gavage with Alistipes species restore TNF-dependent tumor response to immunotherapy whereas Lactobacillus species dampened this response (Figure 1) [18]. A second study explored the role of microbiota composition in tumor-bearing mice treated with anti-CTLA-4. In this study, anti-CTLA-4 activity was inhibited in germ-free or ATB-treated mice compared to controls. A single injection of anti-CTLA-4 significantly affect the microbiome composition with a relative enrichment of Bacteroides thetaiotaomicron, Bacteroides uniformis and to a less extent Bacteroides fragilis. Recolonization of germ-free or ATB-treated mice with these species restored the anticancer response to anti-CTLA4, and increased Th1 immune responses in the tumor-draining lymph nodes and maturation of intratumoral dendritic cells (Figure 1). In this study, authors also suggested that anti-CTLA-4-induced dysbiosis was linked to the severity of gut inflammation and intestinal lesions [29]. Sivan et al. observed that microbiota influences anti-PD-L1 response in a melanoma animal model. Mice harboring different microbiota had different outcomes regarding antitumor immunity and melanoma growth. Abundance of Bifidobacterium species in gut microbiota was found to be correlated with T-cell response and treatment with anti-PD-L1 in combination with oral feeding with these bacteria led to nearly abolishing tumor outgrowth by increasing dendritic cells maturation and antitumor CD8+ T cells activity (Figure 1) [30]. Altogether, these pioneering studies demonstrated the link between microbiota composition and response or IrAE to ICB in preclinical models.

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#### • Evidence in patients with cancer

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Numerous clinical studies investigated microbiota's influence in patients with solid cancer treated with ICBs. The first study in patients investigated microbiota composition and the onset of ipilimumab-induced colitis in metastatic melanoma patients. The composition of microbiota was assessed before ipilimumab initiation; microbial richness was found to be similar between patients who developed anti-CTLA-4-induced colitis and those who remained colitis free. However, Bacteroidetes phylum was underrepresented in the patients who developed immunemediated colitis [21]. Similar to this work, a study in patients with metastatic melanoma and treated with ipilimumab confirmed that patients with baseline microbiota enriched with Bacteroidetes phylum were protected from ipilimumab-induced colitis [20]. In mice, anti-CTLA-4 treatment induced a microbial dysbiosis after the first injection [29] that was not observed in patients except at the time of ipilimumab-induced colitis [20]. Patients with metastatic melanoma with baseline microbiota enriched with Faecalibacterium genus and other Firmicutes had longer progression-free survival (PFS) and overall survival (OS) compared to patients whose baseline microbiota was driven by *Bacteroides* genus. At the species level, Faecalibacterium prausnitzii was more represented in stools at baseline in patients that will benefit from ipilimumab treatment. Moreover, a lower proportion of peripheral blood regulatory T cells was observed in patients with microbiota enriched with Faecalibacterium. Finally, ipilimumab treatment led to a higher ICOS induction on CD4+ T cells in patients whose baseline microbiota was enriched with Faecalibacterium showing a link between gut microbiota composition and immunodynamics of ipilimumab (Figure 1) [20]. Another study realized in 39 patients with metastatic melanoma and treated with ipilimumab, nivolumab, ipilimumab plus nivolumab, or pembrolizumab found a number of bacteria in the Firmicutes phylum (Faecalibacterium prauznitzii, Holdemania filiformis, and Dorea formicigenerans) associated with anti-CTLA-4 plus anti-PD-1 clinical efficacy [19]. Gopalakrishnan et al.

investigated the composition of gut microbiome of patients with metastatic melanoma treated with anti-PD-1. Higher germs diversity and abundance of Ruminococcaceae were found in responding patients, and high diversity in the stool microbiome was correlated with significantly prolonged PFS contrasting with low stool diversity and abundance of Bacteroidales in non-responder patients (Figure 1). At the genus level, patients with high Faecalibacterium abundance had significantly prolonged PFS while patients with high abundance of Bacteroidales had shortened PFS [22]. All these studies were conducted in patients with metastatic melanoma however ICB are also indicated in a wide range of tumors and recent studies suggested that good or hostile microbiota composition may vary from a tumor location to another. In patients with non-small cell lung cancer (NSCLC) or urothelial carcinoma and treated with anti-PD-1, ATB treatment was associated with shortened PFS and OS. Baseline microbiota analysis showed a higher richness in patients with clinical benefit. Overrepresentation of unclassified and classified Firmicutes, as well as Akkermansia and Alistipes was found in responding patients. At the species level, Akkermansia muciniphila was the most significantly associated with favorable clinical outcome in these epithelial tumors [23]. A recent work on advanced renal cell carcinoma patients treated with nivolumab, demonstrated that the alpha-diversity (measure of microbiome diversity applicable to a single sample) and beta-diversity (measure of similarity or dissimilarity of two communities i.e. between samples) of the patient's stool composition might be used as biomarkers to differentiate responder from non-responder population, and to predict which patient could have a PFS longer than a year [24]. The use of ATB treatments can affect the gut microbiome composition thus it is very difficult to differentiate the chicken from the egg. In patients with NSCLC treated with anti-PD-1/PD-L1 monotherapy, ATB treatment was associated with decreased alpha diversity at baseline and patients who did not receive ATB had feces enriched with Clostridia, specifically Ruminococcaceae UCG 13, Clostridiales, and Agathobactera. To determine whether the

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composition of the microbiota influences response to ICB even in the absence of ATB, its composition was explored in eubiotic patients by excluding those who received ATB [25]. In ATB-free patients, decreased alpha diversity associated with shorter OS, *Ruminococcaceae* UCG 13 and *Agathobacter* were overrepresented in patients with clinical benefit. *Akkermansia* species was not associated with improved clinical outcomes but was associated with less severe irAE profile (Figure 1) [25]. Interestingly, *Agathobacter*, that was associated with a favorable outcome, was also associated with more severe irAE [25].

## Microbiota-derived metabolites and immunotherapy-induced response or

#### **IrAE**

As described above, gut microbiome composition influences outcomes and IrAE of ICB, but how it could have an impact on a distant tumor lesion remains unclear. Gut microbiota influences immunity mainly through its metabolites, such as short chain fatty acids (SCFA) as acetate, propionate and butyrate, indoles derivatives, polyamines, water-soluble vitamins and others, produced in very large amounts in the colon through bacterial fermentation of dietary fibers [31,32]. Some studies have therefore investigated whether these metabolites could be related to the composition of the microbiota and to response or IrAE to ICB. Shotgun metagenomic sequencing for metabolic pathway reconstruction on patients treated with ipilimumab with and without colitis revealed that spermidine/putrescine polyamine transport system and three modules involved in the biosynthesis of B vitamins ((riboflavin (B2), pantothenate (B5) and thiamine (B1)) were more abundant in colitis-free patients [21]. This observation seems reminiscent of several data in inflammatory bowel disease were blood B vitamins (thiamine and riboflavin) concentration was significantly lower [33]. Thus, determination of these vitamins in blood could be an interesting tool for the early detection of anti-CTLA-4-induced colitis. To date, no data have been provided to support this hypothesis,

but since validated clinical routine tests exist, it seems interesting to include these measurements in the biological assessment of patients before the start of ICB. A recent study explored the role of systemic SCFA (acetate, propionate and butyrate) and ipilimumab outcomes [34]. As described before, gut microbiota enriched in Faecalibacterium is linked to clinical benefit [20] and these microorganisms are SCFA producers. Coutzac and colleagues showed that Faecalibacterium were inversely correlated with serum concentration of butyrate, and high serum butyrate (and propionate) concentration was associated with shorter PFS and OS in two independent cohorts of patients with metastatic melanoma and treated with ipilimumab [34]. In mice, butyrate gavage was able to limit anti-CTLA-4 anti-tumor efficacy through the inhibition of dendritic cell maturation and restrained anti-CTLA-4-induced tumorspecific T cells. In patients, high blood concentration of butyrate (and propionate) was associated with higher concentration of regulatory T cells before ipilimumab initiation, and patients with low butyrate concentrations at baseline had higher increase of memory CD4+ T cells as well as ICOS induction following ipilimumab, compared to patients with high serum butyrate [34]. Interestingly, a work from Ferrere et al. [35] described that ketogenic diet is associated with increase of ketone bodies such as  $\beta$ -hydroxybutyrate and acetoacetate. Ketogenic diet or treatment with  $\beta$ -hydroxybutyrate could synergized with anti-CTLA-4 + anti-PD-1 therapy in mice. It is described that ketogenic diet affect microbiome composition with an increase in the abundance of Firmicutes and Actinobacteria and a decrease in the abundance of *Bacteroidetes*. Moreover, under these ketogenic conditions, plasma ketone bodies levels increased markedly, whereas plasma SCFA such as propionate and butyrate levels decreased dramatically [36]. Finally Mager et al. showed that inosine could modulate response to anti-CTLA-4 associated to the Toll-like receptor 9 agonist CpG [37]. In this study, authors demonstrated that Bifidobacterium pseudolongum could significantly enhance efficacy of ICB through production of the metabolite inosine. Inosine could activate Th1 in an adenosine A2A

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receptor-dependent fashion and required a costimulus (CpG agonist) along with anti-CTLA-4 treatment. In this work, it has to be stressed out that in the absence of CpG or other costimulus, inosine increased tumor weight and reduced anti-CTLA-4-induced antitumor immunity demonstrating that the effect of inosine was context-dependent [37]. Indeed and as demonstrated by authors through *in vitro* experiments, the effect of inosine in terms of induction or inhibition Th1 differentiation was context dependent; in the presence of exogenous IFN-γ, inosine strongly boosted Th1 differentiation of naïve T cells while in the absence of IFN-γ, inosine inhibited Th1 differentiation [37]. Altogether, these studies provided a vision on the relationship between composition of the gut-microbiota and clinical response to ICB via microbiota-derived metabolites.

## Strategies to modulate the microbial composition of the gut microbiota in

## patients with cancer and treated with ICB

Human microbiome is investigated as a novel therapeutic target to support treatment with ICB as a personalized treatment to improve prognosis of cancer patients. Main approaches of gut microbiota's modulation to potentiate immunotherapy response in cancer patients are presented in Figure 2.

#### • Fecal microbiota transplantation (FMT)

FMT is the most direct method to change the composition of gut microbiota. FMT is a therapy that restores intestinal microbial diversity with the transfer of fecal material from a healthy individual into the gastrointestinal tract of a patient. FMT has demonstrated remarkable clinical efficacy against *Clostridium difficile* infection and it is now recommended for the treatment of this infection for recurrent or refractory stage [38–40]. Interest is growing in the therapeutic potential of FMT for other diseases, e.g., inflammatory bowel diseases or hematologic

malignancies. Several studies have highlighted microbiome's role in the pathogenesis of autoimmune and inflammatory diseases. The pathogenesis is not fully understood, but may implicate environmental factors (life-style, diet, drugs, and infections) and certain genetic backgrounds. Several preclinical studies have experimented FMT, as already discussed in the first part of this review, and regarding patients treated with ICB, some data are starting to indicate that FMT could have a potential interest. Wang et al. reported a first case of refractory ICB-associated colitis successfully treated with FMT from healthy donors, in two patients with different solid tumor [41]. Clinical trials combining FMT and ICB treatment are ongoing to assess its role on limiting ICB-induced toxicities or improving clinical outcomes (NCT04038619, NCT04130763, NCT03353402, NCT03341143). Results from two of these clinical trials have been recently published. These phase I trials have assessed safety and efficacy of FMT, where stools from patients who have previously responded to ICB, were transferred to ICB-resistant patients with metastatic melanoma. In the first study, stool from two donors (donor 1 and donor 2) with complete response (CR) for at least 1 year following anti-PD-1 treatment were used. FMT was preceded with microbiota depletion with oral ATB treatment. FMT was then performed with a colonoscopy followed by the administration of oral stool capsules before reinduction of anti-PD-1 therapy. Maintenance FMT with oral stool capsules was realized along with anti-PD-1 every 14 days with a total of six combined treatment cycles. None of the 10 patients who had FMT developed any moderate-to-severe irAEs (grade 2 or greater) even if five patients did developed irAEs during their first ICB-line. Objective response to treatment were demonstrated in three recipients, one CR and two partial response (PR). All patients who experienced an objective response received FMT from the donor number one in this study. No specific response-inducer microbiota characteristics could be evidenced [42]. In a second study, stool from 7 donors, 4 with CR and 3 with PR following anti-PD-1 treatment were used [43]. Sixteen melanoma patients, all refractory to anti-PD-1 therapy, were

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included and received a single FMT performed with a colonoscopy followed by anti-PD-1 therapy every 3 weeks until disease progression or intolerable toxicity. Among the fifteen patients evaluable for clinical response, one patient experienced a CR and 2 patients experienced a PR. This study showed that FMT successfully colonized the gut of responding patients and could reprogrammed the tumor microenvironment to overcome primary resistance to anti-PD-1 in some patients with advanced melanoma. These results suggest that FMT in melanoma patients seems to be safe and to have a clinical benefit in ICB-resistant patients. However, the number of patients treated remains very low and these clinical trials were not randomized; thus, it cannot be ruled out that the resumption of anti-PD-1 therapy is responsible for objective clinical responses independent of FMT. Further controlled studies should allow us to answer this question. Moreover, we have to remain cautious about security of FMT, as this therapy can increase the risks of transmitting other pathogens, although transplant origin is from healthy subjects with a supposedly well-balanced microbiome [40]. Recently, in a pilot FMT study treating patients with steroid-refractory intestinal acute graft-versus-host-disease [44], a patient died of cerebral hemorrhage caused by thrombocytopenia. No adverse events were observed during and after FMT, but this event is to be pointed out. More recently, one immunocompetent patient died, and a second was infected, after receiving the same donors' stools preparations containing extended-spectrum β-lactamase-producing Escherichia coli [45]. These events stressed out the necessity to constantly improve donor screening to limit transmission of microorganisms that could lead to adverse infectious events especially in fragile patients such as cancer patients.

#### Antibiotics

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As ATB inhibit bacteria growth or induce certain strains death, they can alter microbiota diversity and composition, which may play a role on tumorigenesis and on tumor's response to

treatment. Presence of F. nucleatum had a close relationship with the poor prognosis of CRC patients and probably protected tumor against immune cell attack [46,47]. Using ATB would be a way to decrease F.nucleatum in CRC patients as generally, clinically isolated F. nucleatum strains are sensitive to several ATB, such as metronidazole, clindamycin, and some β-lactamin ATB [48]. A study has shown that metronidazole treatment effectively decreased tumor volumes in CRC patient-derived xenograft mice model [49]. Several studies have investigated ATB role on ICB efficacy, as presented in the first part of our work. They presented a correlation between ATB-pretreatment, short relapse, and survival with shorter PFS and OS [23,50]. Routy et al. observed that ATB compromised efficacy of anti-PD-1 treatment in patients with advanced NSCLC, renal cell carcinoma or urothelial carcinoma. They studied fecal samples' microbiome, segregating responders from non-responders and demonstrated that an overrepresentation of Firmicutes, as well as Akkermansia and Alistipes were associated with favorable clinical outcome in patients with NSCLC and renal cell carcinoma [23]. A recent paper demonstrated that ATB treatment in NSCLC patients could reduce the alpha-diversity (within sample diversity) of the patient's stool composition thus, it remains difficult to determine if microbiota composition at baseline or ATB-use could be a biomarker of ICB efficacy. A meta-analysis of observational studies with 826 patients treated with ATB (28,6%) out of 2889 patients with solid cancers and treated with ICB showed negative outcomes in ATB-treated patients, with shorter PFS and OS [51]. Broad spectrum ATB can have detrimental effect on ICB efficacy; research on targeted ATB to selectively increase immune-potentiating bacteria might be a potential lead to improve current ICB therapies.

#### • Prebiotics, Probiotics, and postbiotics

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Intervention strategies on microbiome are currently investigated, using probiotics, prebiotics, symbiotics and postbiotics to favorably modulate patient's gut microbiota. Various clinical and experimental studies show the role of these microbiota manipulations in cancer prophylaxis and

treatment, throughout mechanisms like apoptosis, antioxidant activity, immune response and epigenetics [52,53]. Prebiotics are nondigestible, oligosaccharides food ingredients that affect the host by selectively stimulating the growth and activity of bacteria in the colon that have the potential to improve host health. Probiotics refer to live microorganisms that show beneficial effects on the host health by modifying microflora of the host. Postbiotics are non-viable bacterial products or functional bioactive compounds, produced during fermentation from probiotics microorganisms, which may be used to promote health. They include many compounds as metabolites, short-chain fatty acids (SCFAs), microbial cell fractions, functional proteins, extracellular polysaccharides, cell lysates, teichoic acid, peptidoglycan-derived muropeptides and pili-type structures [54,55]. The effect of these microbe-targeted therapies has been investigated with anti-cancer therapeutic strategies such as chemotherapy, radiotherapy or surgery, but studies with ICB remain scarce, although many encouraging results in vitro reported effects of these therapies on the immune system [56]. Concerning prebiotics, Liu et al. recently presented a case of murine colon cancer treated with anti-PD-L1 therapy and prebiotic bilberry anthocyanin combo, this association was accompanied with improved tumor CD8+ T-cell infiltration and control of the tumor growth. Their results also showed that prebiotic might play a role in potentiating ICB treatment by restoring microbiota diversity [57]. The immunomodulating role of probiotics [58,59] has led to increased interests toward their potential role in enhancing antitumor immunity. Traditional probiotics generally show few improvements, but next generation probiotics could be interesting therapeutic tool. They have an immunomodulatory effect and certain bacterial strains can modulate dendritic cells maturation stage and increase IL-10 release [60,61]. They may also induce a change in the macrophage phenotype, from pro-inflammatory M1 to anti-inflammatory M2 [62]. Certain strains, in particular Lactobacillus acidophilus [63] and Lactobacillus casei BL23 [64], have shown antitumor properties in mouse models. Other strains such as Enterococcus hirae and

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Barnesiella intestinihominis increase tumor infiltration of CD8+ T lymphocytes [65]. Several studies in mice have shown that the use of immunotherapies combined with probiotics such as Bifidobacterium spp, Akkermansia muciniphila or Enterococcus hirae increases clinical efficacy in mice as already discussed [23,30]. However, we must be vigilant and data observed in mice must be validated in patients before encouraging the use of probiotics. Indeed, preliminary results presented a 70% lower chance of response to ICB for patients who took commercially available probiotics. These commercial probiotics were associated to low diversity of gut microbiome, previously found to be linked to poorer response to ICB [66]. Postbiotics have been shown to allow modulation of inflammatory and anti-inflammatory activities by affecting immune cell migration, adhesion, cytokine expression, cell proliferation and activation, and apoptosis. As already discussed, postbiotics such as butyrate, propionate [34] or inosine [37,67] could modify ICB clinical efficacy. These recent findings support the possible utilization of specific bacterial therapies or microbiota modulation in addition to ICB to enhance their efficacy but few studies presented their link with immune-mediated side effects, such as diarrhea, colitis, or dermatologic events. More studies are needed to confirm the therapeutic potential of these approaches and to clarify the mechanisms. Identification of molecular pathways implicated in the activity of such microbiota-derived metabolites could help in the identification of new targets that could enhance ICB activity in patients.

#### Bacteriophages

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Bacteriophages are viruses that infect and replicate only in bacterial cells. They play an important role in regulating bacterial populations and are naturally present in the microbiome to insure equilibrium. They could be engineered for therapeutic purposes [68] but no clinical trials on human have started yet to our knowledge and preclinical studies remain scarce. A prophage is a bacteriophage genome inserted and integrated into the circular bacterial DNA chromosome. Recently, one dominant epitope of bacteriophage tail length tape measure protein

(TMP1) encoded by the genome of Siphoviridae bacteriophages, was found in *Enterococcus hirae* microbial genome thus an *Enterococcus hirae* prophage. TMP1 coded by *Enterococcus hirae* prophage microbial genome has been found to be an epitope which could induce a memory CD8+ T cell response which cross-reacts with cancer antigen (Figure 2). The clinical relevance of this molecular mimicry theory has then been investigated in patients with lung or renal cancer. In patients, presence of bacteriophage was associated with better survival after anti-PD-1 immunotherapy. Cancer patients in whom the TMP phage sequence was found in the microbiota at baseline, showed a better response to PD-1 blockade [69]. This study highlights the importance of microbiota-derived peptides, which can cross-react with tumor antigens. Furthermore, the characterization of antigens and bacteriophages that transmit these cross-reactive antigenic specificities to commensal bacteria may allow the design of new and specific vaccine candidates to boost ICB efficacy.

## Conclusion

Results from preclinical and clinical studies did not identify abundance of one specific bacteria that could be a common biomarker of ICB response. Different bacteria species can have opposite effects on anticancer treatment, and high inter-individual variation in its composition, diet, medications, and geographical localization have to be considered. However, a common point seems to be found in most studies, the richness and diversity of the gut microbiome appeared to be associated with better clinical outcome. Several hypotheses were raised about microbiota composition and response to ICB. First, antigens from gut microorganisms could stimulate anticancer immune response on their own through cross-reactivity (i.e. antigen mimicry between gut bacteria and tumor antigens), or provide a synergistic effect with ICB through the modulation of the immune system and/or inflammation at distant site. More results are needed to better understand microbiota mechanisms and its role in ICB efficacy. Precision

medicine in cancer treatment would involve modulation of the microbiota through different strategies to improve patients' outcomes. Current means available for modulating the response to immunotherapy using the microbiota seem promising but have also various limits, such as immune-mediated side effects and lack of safety data.

## Figure Legends

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Figure 1. Gut microbiota modulation of ICB treatment efficacy and toxicity. A-C. Data from mouse preclinical models. Main phylum genus or species that have been described as associated with response or resistance to CpG combined with a blocking IL-10 receptor (IL-10R) antibody (A) or blocking CTLA-4 antibodies (B) or blocking anti-PD-L1 antibodies (C) are presented. **D-G**. Data from patients treated with ICB. Main phylum genus or species that have been described as associated with response/resistance/IrAE, in patients with metastatic melanoma and treated with ipilimumab (αCTLA-4, **D**), blocking anti-PD-1 antibodies (αPD-1, E) or combo treatment ipilimumab and nivolumab (αCTLA-4+αPD-1, G) are presented. Main phylum genus or species that have been described as associated with response or resistance, in patients with epithelial tumors as NSCLC and renal cell carcinoma and treated with  $\alpha PD-1$  or  $\alpha PD-L1$  (**F**) are also presented. Figure 2: Strategies to modulate microbiota composition in cancer patients treated with **ICB.** A. FMT using stools from patients with complete or partial response (CR or PR) to another non-responder or who has ICB-induced colitis. **B.** All possible strategies. Antibiotics could be used if certain cocktails are validated to specifically deplete bacteria associated with resistance or toxicities. The value of FMT needs to be validated in controlled clinical trials. Prebiotics such as bilberry anthocyanins could enhanced ICB anti-tumor effect; probiotics such as Faecalibacterium, Bifidobacterium, Enterococcus hirae, Akkermansia muciniphila, Lactobacillus or Burkholderia could modulate the diversity of the gut microbiota and enhance anti-tumor immunity; postbiotics with some having a beneficial impact like inosine and others having a detrimental impact, such as butyrate or propionate, targeting specific pathways associated with these detrimental effects could enhance ICB efficacy. C. Prophage can activate immune response through molecular mimicry between phage-encoded and cancer antigens and

- improve survival of ICB-treated patients. Discovery of these phage-encoded antigens may allow the discovery of new targetable tumor antigens.

## **DECLARATION**

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#### 430 **BIBLIOGRAPHY**

- Ussar S, Fujisaka S, Kahn CR. Interactions between host genetics and gut microbiome in diabetes and metabolic syndrome. Mol Metab 2016;5:795–803. https://doi.org/10.1016/j.molmet.2016.07.004.
- 434 [2] Zmora N, Zeevi D, Korem T, Segal E, Elinav E. Taking it Personally: Personalized 435 Utilization of the Human Microbiome in Health and Disease. Cell Host Microbe 436 2016;19:12–20. https://doi.org/10.1016/j.chom.2015.12.016.
- Tralau T, Sowada J, Luch A. Insights on the human microbiome and its xenobiotic metabolism: what is known about its effects on human physiology? Expert Opin Drug Metab Toxicol 2015;11:411–25. https://doi.org/10.1517/17425255.2015.990437.
- 440 [4] Donaldson GP, Lee SM, Mazmanian SK. Gut biogeography of the bacterial microbiota. 441 Nat Rev Microbiol 2016;14:20–32. https://doi.org/10.1038/nrmicro3552.
- Flint HJ, Scott KP, Duncan SH, Louis P, Forano E. Microbial degradation of complex carbohydrates in the gut. Gut Microbes 2012;3:289–306. https://doi.org/10.4161/gmic.19897.
- Kau AL, Ahern PP, Griffin NW, Goodman AL, Gordon JI. Human nutrition, the gut microbiome, and immune system: envisioning the future. Nature 2011;474:327–36. https://doi.org/10.1038/nature10213.
- Kamada N, Chen GY, Inohara N, Núñez G. Control of Pathogens and Pathobionts by the Gut Microbiota. Nat Immunol 2013;14:685–90. https://doi.org/10.1038/ni.2608.
- 450 [8] Zeng MY, Cisalpino D, Varadarajan S, Hellman J, Warren HS, Cascalho M, et al. Gut
  451 Microbiota-Induced Immunoglobulin G Controls Systemic Infection by Symbiotic
  452 Bacteria and Pathogens. Immunity 2016;44:647–58.
  453 https://doi.org/10.1016/j.immuni.2016.02.006.
- Chung H, Pamp SJ, Hill JA, Surana NK, Edelman SM, Troy EB, et al. Gut immune maturation depends on colonization with a host-specific microbiota. Cell 2012;149:1578–93. https://doi.org/10.1016/j.cell.2012.04.037.
- 457 [10] Helmink BA, Khan MAW, Hermann A, Gopalakrishnan V, Wargo JA. The microbiome, 458 cancer, and cancer therapy. Nat Med 2019;25:377–88. https://doi.org/10.1038/s41591-459 019-0377-7.
- [11] Di Domenico EG, Cavallo I, Pontone M, Toma L, Ensoli F. Biofilm Producing
   Salmonella Typhi: Chronic Colonization and Development of Gallbladder Cancer. Int J
   Mol Sci 2017;18. https://doi.org/10.3390/ijms18091887.
- 463 [12] Gong J, Chehrazi-Raffle A, Placencio-Hickok V, Guan M, Hendifar A, Salgia R. The 464 gut microbiome and response to immune checkpoint inhibitors: preclinical and clinical 465 strategies. Clin Transl Med 2019;8:9. https://doi.org/10.1186/s40169-019-0225-x.
- 466 [13] Grivennikov SI, Wang K, Mucida D, Stewart CA, Schnabl B, Jauch D, et al. Adenoma-467 linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour 468 growth. Nature 2012;491:254–8. https://doi.org/10.1038/nature11465.
- 469 [14] Ishaq S, Nunn L. Helicobacter pylori and gastric cancer: a state of the art review.
  470 Gastroenterol Hepatol Bed Bench 2015;8:S6–14.
- 471 [15] Gao Z, Guo B, Gao R, Zhu Q, Qin H. Microbiota disbiosis is associated with colorectal cancer. Front Microbiol 2015;6:20. https://doi.org/10.3389/fmicb.2015.00020.
- Hard [16] Bhatt AP, Redinbo MR, Bultman SJ. The role of the microbiome in cancer development and therapy. CA Cancer J Clin 2017;67:326–44. https://doi.org/10.3322/caac.21398.
- 475 [17] Karin M, Jobin C, Balkwill F. Chemotherapy, immunity and microbiota--a new triumvirate? Nat Med 2014;20:126–7. https://doi.org/10.1038/nm.3473.

- [18] Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, et al.
   Commensal bacteria control cancer response to therapy by modulating the tumor
   microenvironment. Science 2013;342:967–70. https://doi.org/10.1126/science.1240527.
- 480 [19] Frankel AE, Coughlin LA, Kim J, Froehlich TW, Xie Y, Frenkel EP, et al. Metagenomic 481 Shotgun Sequencing and Unbiased Metabolomic Profiling Identify Specific Human Gut 482 Microbiota and Metabolites Associated with Immune Checkpoint Therapy Efficacy in 483 Melanoma Patients. Neoplasia 2017;19:848–55. 484 https://doi.org/10.1016/j.neo.2017.08.004.
- 485 [20] Chaput N, Lepage P, Coutzac C, Soularue E, Le Roux K, Monot C, et al. Baseline gut 486 microbiota predicts clinical response and colitis in metastatic melanoma patients treated 487 with ipilimumab. Ann Oncol 2017;28:1368–79. https://doi.org/10.1093/annonc/mdx108.
- 488 [21] Dubin K, Callahan MK, Ren B, Khanin R, Viale A, Ling L, et al. Intestinal microbiome 489 analyses identify melanoma patients at risk for checkpoint-blockade-induced colitis. Nat 490 Commun 2016;7:10391. https://doi.org/10.1038/ncomms10391.
- [22] Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, et al.
   Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients.
   Science 2018;359:97–103. https://doi.org/10.1126/science.aan4236.
- 494 [23] Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, et al. Gut 495 microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. 496 Science 2018;359:91–7. https://doi.org/10.1126/science.aan3706.
- [24] Derosa L, Routy B, Fidelle M, Iebba V, Alla L, Pasolli E, et al. Gut Bacteria
   Composition Drives Primary Resistance to Cancer Immunotherapy in Renal Cell
   Carcinoma Patients. Eur Urol 2020;78:195–206.
   https://doi.org/10.1016/j.eururo.2020.04.044.
- [25] Hakozaki T, Richard C, Elkrief A, Hosomi Y, Benlaïfaoui M, Mimpen I, et al. The Gut
   Microbiome Associates with Immune Checkpoint Inhibition Outcomes in Patients with
   Advanced Non-Small Cell Lung Cancer. Cancer Immunol Res 2020;8:1243–50.
   https://doi.org/10.1158/2326-6066.CIR-20-0196.
- 505 [26] Matson V, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre M-L, et al. The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients.
  507 Science 2018;359:104–8. https://doi.org/10.1126/science.aao3290.
- 508 [27] Farkona S, Diamandis EP, Blasutig IM. Cancer immunotherapy: the beginning of the end of cancer? BMC Med 2016;14:73. https://doi.org/10.1186/s12916-016-0623-5.
- 510 [28] Chen DS, Mellman I. Oncology meets immunology: the cancer-immunity cycle. 511 Immunity 2013;39:1–10. https://doi.org/10.1016/j.immuni.2013.07.012.
- [29] Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament C, et al. Anticancer
   immunotherapy by CTLA-4 blockade relies on the gut microbiota. Science
   2015;350:1079–84. https://doi.org/10.1126/science.aad1329.
- [30] Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, Earley ZM, et al.
   Commensal Bifidobacterium promotes antitumor immunity and facilitates anti-PD-L1
   efficacy. Science 2015;350:1084–9. https://doi.org/10.1126/science.aac4255.
- 518 [31] Thorburn AN, Macia L, Mackay CR. Diet, metabolites, and "western-lifestyle" inflammatory diseases. Immunity 2014;40:833–42. 520 https://doi.org/10.1016/j.immuni.2014.05.014.
- 521 [32] Maslowski KM, Mackay CR. Diet, gut microbiota and immune responses. Nat Immunol 2011;12:5–9. https://doi.org/10.1038/ni0111-5.
- 523 [33] Pan Y, Liu Y, Guo H, Jabir MS, Liu X, Cui W, et al. Associations between Folate and Vitamin B12 Levels and Inflammatory Bowel Disease: A Meta-Analysis. Nutrients 2017;9. https://doi.org/10.3390/nu9040382.

- 526 [34] Coutzac C, Jouniaux J-M, Paci A, Schmidt J, Mallardo D, Seck A, et al. Systemic short 527 chain fatty acids limit antitumor effect of CTLA-4 blockade in hosts with cancer. Nat 528 Commun 2020;11:2168. https://doi.org/10.1038/s41467-020-16079-x.
- 529 [35] Ferrere G, Tidjani Alou M, Liu P, Goubet A-G, Fidelle M, Kepp O, et al. Ketogenic diet 530 and ketone bodies enhance the anticancer effects of PD-1 blockade. JCI Insight 2021;6. 531 https://doi.org/10.1172/jci.insight.145207.
- [36] Miyamoto J, Ohue-Kitano R, Mukouyama H, Nishida A, Watanabe K, Igarashi M, et al.
   Ketone body receptor GPR43 regulates lipid metabolism under ketogenic conditions.
   Proc Natl Acad Sci U S A 2019;116:23813–21.
   https://doi.org/10.1073/pnas.1912573116.
- 536 [37] Mager LF, Burkhard R, Pett N, Cooke NCA, Brown K, Ramay H, et al. Microbiome-537 derived inosine modulates response to checkpoint inhibitor immunotherapy. Science 538 2020;369:1481–9. https://doi.org/10.1126/science.abc3421.
- 539 [38] Trang C, Scanzi J, Galperine T, Mosca A, Batista R, Sokol H. infection: guidelines actualisation for clinical practice 2021;24:7.
- [39] Sokol H, Galperine T, Kapel N, Bourlioux P, Seksik P, Barbut F, et al. Faecal
   microbiota transplantation in recurrent Clostridium difficile infection: Recommendations
   from the French Group of Faecal microbiota Transplantation. Dig Liver Dis
   2016;48:242–7. https://doi.org/10.1016/j.dld.2015.08.017.
- [40] Brandt LJ, Reddy SS. Fecal microbiota transplantation for recurrent clostridium difficile
   infection. J Clin Gastroenterol 2011;45 Suppl:S159-167.
   https://doi.org/10.1097/MCG.0b013e318222e603.
- 548 [41] Wang Y, Wiesnoski DH, Helmink BA, Gopalakrishnan V, Choi K, DuPont HL, et al. 549 Fecal microbiota transplantation for refractory immune checkpoint inhibitor-associated 550 colitis. Nat Med 2018;24:1804–8. https://doi.org/10.1038/s41591-018-0238-9.
- 551 [42] Baruch EN, Youngster I, Ben-Betzalel G, Ortenberg R, Lahat A, Katz L, et al. Fecal microbiota transplant promotes response in immunotherapy-refractory melanoma patients. Science 2021;371:602–9. https://doi.org/10.1126/science.abb5920.
- 554 [43] Davar D, Dzutsev AK, McCulloch JA, Rodrigues RR, Chauvin J-M, Morrison RM, et al. 555 Fecal microbiota transplant overcomes resistance to anti-PD-1 therapy in melanoma 556 patients. Science 2021;371:595–602. https://doi.org/10.1126/science.abf3363.
- [44] Qi X, Li X, Zhao Y, Wu X, Chen F, Ma X, et al. Treating Steroid Refractory Intestinal
   Acute Graft-vs.-Host Disease With Fecal Microbiota Transplantation: A Pilot Study.
   Front Immunol 2018;9:2195. https://doi.org/10.3389/fimmu.2018.02195.
- [45] DeFilipp Z, Bloom PP, Torres Soto M, Mansour MK, Sater MRA, Huntley MH, et al.
   Drug-Resistant E. coli Bacteremia Transmitted by Fecal Microbiota Transplant. N Engl
   J Med 2019;381:2043–50. https://doi.org/10.1056/NEJMoa1910437.
- [46] Yu T, Guo F, Yu Y, Sun T, Ma D, Han J, et al. Fusobacterium nucleatum Promotes
   Chemoresistance to Colorectal Cancer by Modulating Autophagy. Cell 2017;170:548 563.e16. https://doi.org/10.1016/j.cell.2017.07.008.
- Zhang S, Yang Y, Weng W, Guo B, Cai G, Ma Y, et al. Fusobacterium nucleatum
   promotes chemoresistance to 5-fluorouracil by upregulation of BIRC3 expression in
   colorectal cancer. J Exp Clin Cancer Res 2019;38:14. https://doi.org/10.1186/s13046-018-0985-y.
- 570 [48] Mandell, Douglas, and Bennett's. Principles and Practice of Infectious Diseases. 8th Edition, Saunders; 2014.
- 572 [49] Bullman S, Pedamallu CS, Sicinska E, Clancy TE, Zhang X, Cai D, et al. Analysis of 573 Fusobacterium persistence and antibiotic response in colorectal cancer. Science 574 2017;358:1443–8. https://doi.org/10.1126/science.aal5240.

- 575 [50] Derosa L, Hellmann MD, Spaziano M, Halpenny D, Fidelle M, Rizvi H, et al. Negative 576 association of antibiotics on clinical activity of immune checkpoint inhibitors in patients 577 with advanced renal cell and non-small-cell lung cancer. Ann Oncol 2018;29:1437–44. 578 https://doi.org/10.1093/annonc/mdy103.
- 579 [51] Wilson BE, Routy B, Nagrial A, Chin VT. The effect of antibiotics on clinical outcomes 580 in immune-checkpoint blockade: a systematic review and meta-analysis of observational 581 studies. Cancer Immunol Immunother 2020;69:343–54. https://doi.org/10.1007/s00262-582 019-02453-2.
- 583 [52] Dasari S, Kathera C, Janardhan A, Praveen Kumar A, Viswanath B. Surfacing role of 584 probiotics in cancer prophylaxis and therapy: A systematic review. Clin Nutr 585 2017;36:1465–72. https://doi.org/10.1016/j.clnu.2016.11.017.
- 586 [53] Guarner F, Schaafsma GJ. Probiotics. Int J Food Microbiol 1998;39:237–8. https://doi.org/10.1016/s0168-1605(97)00136-0.
- 588 [54] Żółkiewicz J, Marzec A, Ruszczyński M, Feleszko W. Postbiotics-A Step Beyond Pre-589 and Probiotics. Nutrients 2020;12. https://doi.org/10.3390/nu12082189.
- [55] Akatsu H. Exploring the Effect of Probiotics, Prebiotics, and Postbiotics in
   Strengthening Immune Activity in the Elderly. Vaccines (Basel) 2021;9.
   https://doi.org/10.3390/vaccines9020136.
- 593 [56] Chang C-J, Lin T-L, Tsai Y-L, Wu T-R, Lai W-F, Lu C-C, et al. Next generation 594 probiotics in disease amelioration. J Food Drug Anal 2019;27:615–22. 595 https://doi.org/10.1016/j.jfda.2018.12.011.
- [57] Liu X, Wang L, Jing N, Jiang G, Liu Z. Biostimulating Gut Microbiome with Bilberry
   Anthocyanin Combo to Enhance Anti-PD-L1 Efficiency against Murine Colon Cancer.
   Microorganisms 2020;8. https://doi.org/10.3390/microorganisms8020175.
- 599 [58] Geier MS, Butler RN, Howarth GS. Probiotics, prebiotics and symbiotics: a role in 600 chemoprevention for colorectal cancer? Cancer Biol Ther 2006;5:1265–9. 601 https://doi.org/10.4161/cbt.5.10.3296.
- 602 [59] Mackowiak PA. Recycling metchnikoff: probiotics, the intestinal microbiome and the quest for long life. Front Public Health 2013;1:52. 604 https://doi.org/10.3389/fpubh.2013.00052.
- [60] Konieczna P, Groeger D, Ziegler M, Frei R, Ferstl R, Shanahan F, et al. Bifidobacterium
   infantis 35624 administration induces Foxp3 T regulatory cells in human peripheral
   blood: potential role for myeloid and plasmacytoid dendritic cells. Gut 2012;61:354–66.
   https://doi.org/10.1136/gutjnl-2011-300936.
- [61] Jeon SG, Kayama H, Ueda Y, Takahashi T, Asahara T, Tsuji H, et al. Probiotic
   Bifidobacterium breve induces IL-10-producing Tr1 cells in the colon. PLoS Pathog
   2012;8:e1002714. https://doi.org/10.1371/journal.ppat.1002714.
- 612 [62] Sichetti M, De Marco S, Pagiotti R, Traina G, Pietrella D. Anti-inflammatory effect of 613 multistrain probiotic formulation (L. rhamnosus, B. lactis, and B. longum). Nutrition 614 2018;53:95–102. https://doi.org/10.1016/j.nut.2018.02.005.
- [63] Chen C-C, Lin W-C, Kong M-S, Shi HN, Walker WA, Lin C-Y, et al. Oral inoculation
   of probiotics Lactobacillus acidophilus NCFM suppresses tumour growth both in
   segmental orthotopic colon cancer and extra-intestinal tissue. Br J Nutr 2012;107:1623–
   34. https://doi.org/10.1017/S0007114511004934.
- [64] Jacouton E, Michel M-L, Torres-Maravilla E, Chain F, Langella P, Bermúdez-Humarán
   LG. Elucidating the Immune-Related Mechanisms by Which Probiotic Strain
   Lactobacillus casei BL23 Displays Anti-tumoral Properties. Front Microbiol
   2018;9:3281. https://doi.org/10.3389/fmicb.2018.03281.
- [65] Daillère R, Vétizou M, Waldschmitt N, Yamazaki T, Isnard C, Poirier-Colame V, et al.
   Enterococcus hirae and Barnesiella intestinihominis Facilitate Cyclophosphamide-

- Induced Therapeutic Immunomodulatory Effects. Immunity 2016;45:931–43. https://doi.org/10.1016/j.immuni.2016.09.009.
- [66] Christine N S, Gopalakrishnan V, McQuade J, Wargo JA. Abstract 2838: The gut
   microbiome (GM) and immunotherapy response are influenced by host lifestyle factors
   March 29-April 3.
- 630 [67] Kroemer G, Zitvogel L. Inosine: novel microbiota-derived immunostimulatory 631 metabolite. Cell Res 2020;30:942–3. https://doi.org/10.1038/s41422-020-00417-1.
- [68] Schmidt C. Phage therapy's latest makeover. Nat Biotechnol 2019;37:581–6.
   https://doi.org/10.1038/s41587-019-0133-z.

637

634 [69] Fluckiger A, Daillère R, Sassi M, Sixt BS, Liu P, Loos F, et al. Cross-reactivity between 635 tumor MHC class I-restricted antigens and an enterococcal bacteriophage. Science 636 2020;369:936–42. https://doi.org/10.1126/science.aax0701.













