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1 **Title Page**

2 **Historical dispersal and host-switching formed the evolutionary history of a**
3 **globally distributed multi-host parasite - the *Ligula intestinalis* species complex**

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34 **Abstract**

35 Studies on parasite biogeography and host spectrum provide insights into the processes
36 driving parasite diversification. Global geographical distribution and a multi-host spectrum
37 make the tapeworm *Ligula intestinalis* a promising model for studying both the vicariant
38 and ecological modes of speciation in parasites. To understand the relative importance of
39 host association and biogeography in the evolutionary history of this tapeworm, we
40 analysed mtDNA and reduced-represented genomic SNP data for a total of 139 specimens
41 collected from 18 fish-host genera across a distribution range representing 21 countries.
42 Our results strongly supported the existence of at least 10 evolutionary lineages and
43 estimated the deepest divergence at approximately 4.99-5.05 Mya, which is much younger
44 than the diversification of the fish host genera and orders. Historical biogeography analyses
45 revealed that the ancestor of the parasite diversified following multiple vicariance events
46 and was widespread throughout the Palearctic, Afrotropical, and Nearctic between the late
47 Miocene and early Pliocene. Cyprinoids were inferred as the ancestral hosts for the parasite.
48 Later, from the late Pliocene to Pleistocene, new lineages emerged following a series of
49 biogeographic dispersal and host-switching events. Although only a few of the current
50 *Ligula* lineages show narrow host-specificity (to a single host genus), almost no host
51 genera, even those that live in sympatry, overlapped between different *Ligula* lineages. Our
52 analyses uncovered the impact of historical distribution shifts on host switching and the
53 evolution of host specificity without parallel host-parasite co-speciation. Historical
54 biogeography reconstructions also found that the parasite colonized several areas
55 (Afrotropical and Australasian) much earlier than was suggested by only recent faunistic
56 data.

57 *Keywords:* Historical biogeography, demographic history, *Ligula* species complex, host
58 specificity

59

60 **Introduction**

61 Host specificity is a key feature of parasitic and symbiotic organisms. It defines their ability to
62 survive on different hosts (Poulin, 2011), disperse (Máková et al., 2018) and maintain the
63 genetic diversity of populations (Martinu et al., 2018; Wacker et al., 2019). The evolution of
64 host specificity in multi-host parasites is a prospective candidate for the mechanism of
65 sympatric adaptive speciation (Kalbe et al., 2016). For long, the evolution of host-specific
66 lineages has been viewed mostly from the co-speciation (or co-diversification) angle, where
67 the evolution of the parasite is directly associated with that of the host(s) (Hafner & Nadler,
68 2017; Štefka et al., 2011). However, the origin of host-specific lineages is often linked to a
69 series of historical biogeographical events (Ding et al., 2022; Perrot-Minnot et al., 2018).
70 Therefore, incongruence may appear between the parasite and host phylogenies for various
71 reasons, including founder events, bottlenecks and extinctions either in the host or parasite,
72 and, most importantly, due to host-switching that could have occurred over large-scale episodic
73 periods of palaeo-environmental and climatic fluctuations, such as Quaternary glaciations
74 (Hoberg et al., 2012; Johnson et al., 2002; Zarlenga et al., 2006). Under stable environmental
75 conditions, co-divergence between closely associated organisms may occur (Haukialmi et al.,
76 2016), whereas under changing conditions, hosts possibly move in search of suitable habitats
77 and find themselves in contact with species they did not previously overlap with. These
78 fluctuations provide new opportunities for parasites to switch between host species (Hoberg et
79 al., 2012; Hoberg & Brooks, 2008, 2013).

80 Following a successful host switch, gene flow may be maintained or ceased between the
81 ancestral and newly established parasite population, depending on the duration of the
82 distribution overlap between the host species (e. g., Techer et al., 2022; Wang et al., 2016).
83 Phylogenomic and historical biogeography methods allow for reconstructing past distribution
84 changes and associated gene flow events in parasite lineages, providing tools to study the
85 emergence and diversification of parasite species, identification of cryptic lineages and
86 prediction of disease transmission (Angst et al., 2022; Doyle et al., 2022). Yet, although
87 metazoan parasites represent 40% of all living metazoan species, relatively few studies have
88 investigated the historical biogeography of parasitic species (Dobson et al., 2008). Instead,
89 these studies often investigate the host–parasite co-phylogeny. Whilst the co-phylogenetic
90 approach has demonstrated significant co-evolutionary signals in parasites with a direct life
91 cycle (Rahmouni et al., 2022; Šimková et al., 2004, 2022), it often finds weak host–parasite
92 co-evolutionary signals in generalist multi-host parasites or parasites with complex life cycles
93 involving intermediate hosts, possessing more varied dispersal opportunities (Hoberg &
94 Brooks, 2008; Perrot-Minnot et al., 2018). Importantly, the co-phylogenetic framework does
95 not consider the evolution of host-specific lineages in parasites whose origin is much younger
96 than the age of the hosts, where the co-phylogenetic methods simply cannot be applied (Bouzid
97 et al., 2008) .

98

99 To study the contribution of past processes to the evolution of host-specific parasite lineages,
100 we applied historical biogeography methods to study the tapeworm *Ligula intestinalis* sensu
101 lato (Cestoda: Diphyllbothriidea). Its widespread geographical distribution and multi-host
102 spectrum make it a promising model for studying the vicariant and ecological modes of
103 speciation (Hoole et al., 2010; Nazarizadeh et al., 2022a; Štefka et al., 2009). It is an obligatory
104 endoparasite with a complex life cycle involving two intermediate hosts (crustacean copepod

105 as the first and planktivorous fish as the second intermediate host) and birds feeding on the
106 infected fish as the definitive host (Dubinina, 1980). *L. intestinalis* spends a maximum of only
107 five days in its final host, rapidly entering the sexual phase and dispersing eggs into the water
108 with host faeces, while the secondary larval stage, plerocercoid, represents the most distinct
109 phase of the parasite's life cycle (Dubinina, 1980). It lasts at least 10 months and occurs in the
110 peritoneal cavity of planktivorous fish, negatively impacting the health, fertility and behaviour
111 of individual fish (Gabagambi et al., 2019; Loot et al., 2002) as well as the ecology of the entire
112 fish population (Kennedy et al., 2001). *L. intestinalis* usually appears in cyprinoid fish
113 [previously the family Cyprinidae], but is capable of invading intermediate hosts from various
114 taxonomic groups such as Salmonidae, Galaxiidae and Catostomidae (Chapman et al., 2006;
115 Dubinina, 1980; Štefka et al., 2009).

116 Studies exploring the phylogenetic relationships and population structure of the *L. intestinalis*
117 species complex identified at least six major lineages with different levels of host and
118 geographic distribution: (1) clade A (European and north African populations), (2) clade B
119 (European, Chinese, Australian and North African populations), (3) *L. alternans* (Asia)
120 [previously *L. digramma*], (4) Canada, (5) Chinese and (6) Ethiopian populations (Bouزيد et
121 al., 2008; J. Štefka et al., 2009). The studies indicated that both geography and host specificity
122 may determine the genealogical relationships of the parasites by inducing genetic
123 differentiation. However, their sampling lacked several biogeographical areas, and the
124 historical demography of the lineages was not elucidated. Thus, the course of interaction
125 between the historical geographical distribution of the parasite and the evolution of host-
126 specific lineages has remained a contentious issue.

127 The main objective of the present study is to reveal the contribution of historical geographical
128 distribution and past host usage (specificity) in the formation of contemporary lineage diversity
129 in parasites with high dispersal capabilities. Using a geographically wide sampling of *Ligula*

130 tapeworms as a study system, we characterised it with SNP genotyping and mitochondrial
131 sequencing. We aimed to i) determine the true lineage diversity and levels of host specificity
132 of parasites forming a species complex which lacks distinctive morphological traits; ii) provide
133 dates for major diversification events in the evolutionary history of the parasite, and iii)
134 determine the factors and events influencing lineage diversity in space and time by the analyses
135 of historical areas, demographic history of lineages and past gene flow.

136 **Materials and Methods**

137 **Sample collection and DNA extraction**

138 We analysed 139 specimens of *Ligula* plerocercoids collected from 18 fish genera over a broad
139 geographic area representing 21 countries (Table S1). Integrity and quantity of newly isolated
140 samples (extracted with DNeasy blood and tissue kit [Qiagen] from specimens preserved
141 in 96% ethanol) and DNA extracts from previous studies (Bouzid et al., 2008; Štefka et al.,
142 2009) were verified on 0.8% agarose gel and with Qubit 2.0 Fluorometer (Invitrogen). The
143 final set of mitochondrial (86 new specimens were added to 76 sequences from Bouzid et al.,
144 2008 and Štefka et al., 2013 studies) and ddRAD (all new specimens) samples covered the
145 distribution of the major lineages found in previous studies, which was complemented with
146 several new areas to represent five terrestrial biogeographic realms: Nearctic, Palearctic,
147 Afrotropical, Oriental and Australasian (Fig. 1, Table S1a and S1b).

148 **Mitochondrial DNA sequencing**

149 Amplification of three fragments of the mitochondrial genome, including cytochrome b (Cyt
150 *b*), cytochrome oxidase subunit 1 (COI) and NADH dehydrogenase 1 (ND1) genes generated
151 a concatenated dataset with a total length of 1693 bp. All genes were amplified by polymerase
152 chain reaction (PCR; see Supplementary Material, section 1) and sequenced using primers
153 specifically designed for this study or previously published primers (Table S2). Sanger

154 sequences were assembled and edited using Geneious Prime v.2022.1 (Gene Codes) and
155 checked for possible stop codons based on the Alternative Flatworm Mitochondrial Code
156 (transl_table 14).

157 **ddRAD library preparation and reference genome**

158 ddRAD libraries were prepared using 350 ng of DNA per sample following a modified
159 version of the protocol proposed by Peterson et al. (2012) (Supplementary Material, section
160 2). Two samples of *Dibothriocephalus latus* (Diphyllbothriidea) were included as outgroups.
161 Libraries were sequenced at several lanes of Illumina NovaSeq 150 bp PE (Novogene UK)
162 yielding 6.8 million paired-end reads per sample on average. Steps for preparation of
163 multiplexed ddRAD-seq libraries for each barcoded individual sample (Table S1a) and
164 purification are outlined in supplementary material, section 2. We also generated a draft
165 genome of *Ligula* using Illumina platforms (OmniSeq), yielding an assembly comprising
166 49,218 scaffolds and covering 780 Mb of the genome; gene prediction and annotation for this
167 assembly are ongoing.

168 **ddRAD data assembly and SNP calling**

169 FastQC (Andrews, 2010) was applied for an initial quality check. The process_radtags
170 program in Stacks v.2.5.3 was used to demultiplex and filter the raw reads with low quality and
171 uncalled bases (Rochette et al., 2019). Also, cut sites, barcodes and adaptors were removed
172 from ddRAD outputs. Next, paired-end read alignment of each sample to the reference genome
173 was performed using the default mode of Bowtie 2 (Langmead & Salzberg, 2012). SNPs calling
174 and assembling loci were carried out from the alignment using the ref_map.pl wrapper
175 program. Data were then filtered for missingness, minor allele frequency (MAF) and linkage
176 disequilibrium (LD) in Stacks (see Supplementary Material, section 3 for details). In sum, we
177 generated three SNP matrices in variant call format (VCF) comprising (1) 44,298 SNPs with a
178 mean coverage per locus of ca. 20 \times , (2) second linkage disequilibrium filtered data set (0.2

179 squared coefficient of correlation) including 10,937 SNPs and a mean coverage per locus of
180 ca. 18×, and (3) two outgroups which were added to the third data set comprising 4471 unlinked
181 SNPs and a mean coverage per locus of ca. 25×.

182 **Analysis of sequence diversity**

183 Three gene fragments (*Cyt b*: 405 bp COI: 396 bp, and ND1: 893 bp) were added to an
184 alignment that included 76 sequences of *Cyt b* and COI from Bouzid et al. (2008), Bouzid et
185 al. (2013) and Bouzid et al. (2013) studies. All gene fragments were aligned separately using
186 MUSCLE performed in MEGA v.5 (Tamura et al., 2011). We used the three gene fragments
187 for phylogenetic analyses; however, all genetic distance-based analyses were carried out based
188 on *Cyt b* and COI sequences which had no missingness. Analysis of population genetic
189 diversity (sequence polymorphism, haplotype diversity and nucleotide diversity) was
190 performed in DnaSP v.5 (Librado & Rozas, 2009) on samples grouped according to their
191 phylogenetic relationships (see below).

192 For nuclear SNP data, we estimated the expected (HE) and observed (HO) heterozygosities,
193 private alleles, and inbreeding coefficients among the parasite populations using the R package
194 SambaR (de Jong et al., 2021). The first ddRAD dataset (44,298 SNPs) was applied for the
195 analysis as the calculation assumed that the input data were not reduced based on LD filtering.
196 We used Plink v.1.90 (Chang et al., 2015) to convert the VCF file to the bed format. The default
197 data filtering was applied with the “filterdata” function in the R package (see De Jong et al.,
198 2021).

199 **Phylogenetic analyses**

200 The best-fitting substitution models and partition schemes were estimated for the three mtDNA
201 genes using a greedy algorithm in PartitionFinder v.2.1.1 (Lanfear et al., 2017). Using Bayesian
202 inference (BI) and maximum likelihood (ML), the phylogenetic relationships of the *Ligula*
203 lineages were reconstructed. BI analysis was conducted using MrBayes v.3.1.2 (Ronquist &

204 Huelsenbeck, 2003) on the data partitioned based on codon positions and using the settings as
205 described in supplementary material, section 4. We inferred the ML tree using IQTREE v.2.1.2
206 (Minh et al., 2020) under the best-fitting model from PartitionFinder (Table S3) and 1,000
207 ultrafast bootstrap replicates. We also used the -bnni option to minimize the risks of
208 overestimating support values to a minimum. We included *Dibothriocephalus nihonkaiensis*
209 and *D. latus* (Li et al., 2018a), sister groups of *Ligula*, as the outgroups.
210 We utilised the third data set (4471 unlinked SNPs) to reconstruct the phylogenetic gene tree
211 and the species tree using three approaches. Using PartitionFinder, we first determined the
212 parameters applied as input into RAxML-ng to reconstruct an ML tree (Kozlov et al., 2019).
213 Second, the alignment was used for a Bayesian phylogenetic analysis. BI was completed using
214 ExaBayes v.1.5 (Aberer et al., 2014) under a model partitioning scheme analogous to the ML
215 analyses. Third, species trees were investigated using SVDquartets in PAUP v.4.0a147
216 (Chifman & Kubatko, 2014). Further details on the assessments of convergence and ML, BI
217 and species trees are provided in supplementary material, section 4.

218 **Population structure analyses**

219 Using the concatenated COI and Cyt *b* (801 bp), haplotype networks were constructed to
220 visualize relationships among the parasite populations via the Median-Joining algorithm in
221 PopArt v.1.7 (Leigh & Bryant, 2015). Moreover, ddRAD data set was applied to construct a
222 phylogenetic network using a neighbour-net algorithm and 1,000 bootstraps executed in
223 SplitsTree v.4.10 (Huson & Bryant, 2006).

224 Several clustering analyses were performed to discover the number of distinct genetic groups.
225 First, the model-based evolutionary clustering method was applied in ADMIXTURE
226 (Alexander et al., 2009) via its parallel processing capabilities (AdmixPiPe: Mussmann et al.,
227 2020). Second, a Principal Component Analysis (PCA) was performed using the glPCA
228 function from the adegenet R package (Jombart & Collins, 2015). Next, we applied the

229 TESS3R package in R to provide additional estimates of the role of geography in the genetic
230 structure (Caye et al., 2018). TESS3R explores the implications of genetic diversity in natural
231 populations using geographic and genotypic data simultaneously. Lastly, we applied the
232 programs fineRADstructure and RADpianter v.0.2 (Malinsky et al., 2018) to reveal population
233 genetic structure based on the nearest-neighbour haplotype (For details on ADMIXTURE and
234 fineRADstructure analyses, see Supplementary Material, section 5).

235 **Species delimitation analyses**

236 To detect species boundaries in the *Ligula* species complex using the mtDNA data, species
237 delimitation was performed using three independent approaches (genetic distance and tree-
238 based approaches): the Generalized Mixed Yule Coalescent model (GMYC by Pons et al.,
239 2006), Bayesian implementation of Poisson Tree Processes model (bPTP by Zhang et al.,
240 2013), and Assemble Species by Automatic Partitioning (ASAP by Puillandre et al., 2021
241 Supplementary Material, section 6).

242 To make comparisons among species delimitation models using SNP data, we conducted Bayes
243 Factor Delimitation (BFD, Leaché et al., 2014) implemented in SNAPP in BEAST2 (Bouckaert
244 et al., 2014; Bryant et al., 2012). Comparing models by marginal likelihood scores, this
245 approach assesses support for the alternative model using Bayes factors under the multispecies
246 coalescent (MSC) model. We tested a nested set of hypotheses up to the maximum potential
247 number of species, where every lineage was considered a different species (Supplementary
248 Material, section 6).

249 **Divergence time and phylogeographic structure**

250 For mtDNA, divergence dates were estimated using a coalescent-based model implemented in
251 BEAST v.1.8.2 (Drummond et al., 2012) Due to a lack of available fossil or geographic events,
252 we used two common rates of substitution for the mt-genes (Cyt $b = 0.0195$, COI= 0.0225
253 substitutions/site/Myr), which were previously estimated for species of *Taenia* tapeworms

254 (Cyclophyllidea). In addition, we applied the divergence time between *Spirometra* and
255 *Diphyllobothrium* genera (at the base of Diphyllbothriidea), which is normally distributed
256 with a mean of 11.47 Mya, a standard deviation of 1.14 Mya and a 95% interval of 9.58–13.36
257 Mya. The best-fit partitioning schemes were assigned based on the Akaike information
258 criterion (AICc) in PartitionFinder and the analysis was run using the settings reported in the
259 supplementary material, section 7.

260 To infer the divergence time among major tapeworm lineages using SNP data, we first
261 estimated the substitution rates for protein-coding sequences (CDS) of the *L. intestinalis*
262 genome based on the methods suggested by Wang et al. (2015). Protein sequences of 10
263 flatworms (*D. latus*, *Spirometra erinaceieuropaei*, *Taenia solium*, *T. asiatica*, *T. saginata*,
264 *Echinococcus granulosus*, *E. multilocularis*, *Hymenolepis microstoma*, *Schistosoma*
265 *japonicum*, *S. mansoni*) were obtained from WormBase (<https://wormbase.org/>). After
266 concatenating all CDS parts, they were aligned together using MAFFT v.7.4 ('L-INS-I'
267 algorithm) (Katoh & Standley, 2013) and their gaps were discarded by Trimal v.1.4 (Capella-
268 Gutiérrez et al., 2009). The best evolution models for the CDS alignments were calculated
269 using PartitionFinder (GTR+I+G). ML trees were reconstructed with 400 bootstrapping
270 replicates using RAxML under the best fit models. The substitution rate (branch-length) for
271 *Ligula* was estimated to be 0.0201 which was 1.02-fold higher than that of *T. solium* (0.0196
272 mutations per site) (Fig. S1). Wang et al. (2015) calculated the mutation rate/site/year in the *T.*
273 *solium* genome (2.82×10^{-9}). Therefore, we estimated 2.89×10^{-9} substitutions/site/year for the
274 *Ligula* genome. We used Beast v1.8 with 141 sequences, including two *D. latus* outgroups to
275 reconstruct the divergence dates of the sequences under a birth-death process as a tree prior
276 and the uncorrelated relaxed molecular clock model for the mutation rate (2.89×10^{-9}). The
277 analysis was run using the settings presented in the supplementary material, section 7.

278

279 **Historical biogeography and ancestral host reconstructions**

280 Inferring ancestral ranges and the historical biogeography of *Ligula* was done based on three
281 models of biogeographical range extension, including Bayesian inference (BAYAREA-like),
282 Dispersal-Vicariance (DIVA-like), and Dispersal-Extinction-Cladogenesis (DEC) models, all
283 estimated by the BioGeoBEARS R package (Matzke, 2016). As BioGeoBEARS requires an
284 ultrametric tree of species/population, we used the BEAST chronogram from both mtDNA and
285 ddRAD analyses, and all outgroups and specimens were pruned using Mesquite v.3.7
286 (Maddison & Maddison, 2021).

287 These models enable the exploration of different possibilities of vicariance, extinction, and
288 dispersal. Furthermore, by incorporating a founder-event parameter (+J) into the analysis, we
289 attained clastogenic dispersal outside of the parental areas (referred to as jump speciation). The
290 *Ligula* species complex was divided into five biogeographic realms: (1) Palearctic, (2)
291 Afrotropical, (3) Indomalayan, (4) Australasian and (5) Nearctic. Inferred ancestral ranges
292 were allowed to occupy up to five areas, thereby facilitating clear estimations. Likelihood-
293 based measures of three biogeographic models along with their modified J types were ranked
294 according to the AIC method to evaluate model quality.

295 We applied phylogenetic trees from both mtDNA and ddRAD data to reconstruct ancestral host
296 fish affiliations using a Bayesian framework implemented in sMap v.1.0.7 (Bianchini and
297 Sánchez-Baracaldo, 2021). The software uses stochastic mapping analysis to reconstruct the
298 evolutionary history of discrete characters at the branch and node of the tree. In the present
299 study, *Ligula* plerocercoids were collected from 41 fish species representing 18 genera in four
300 orders (Cypriniformes, Osmeriformes, Gobiiformes and Galaxiiformes). Character states were
301 defined based on the host taxonomy at the order level. Therefore, we defined four different
302 states as follows: 1) lineage China infects Osmeriformes, 2) *L. pavlovskii* was associated with

303 Gobiiformes, 3) lineages of Australia and New Zealand infect Gobiiformes and Galaxiiformes,
304 and 4) lineages of Ethiopia, EAST African Rift (EAR) and lineages A and B were associated
305 with Cypriniformes (Supplementary Material, section 8).

306 **Genome-wide analysis of hybridization and introgression**

307 Treemix v.1.12 (Pickrell & Pritchard, 2012) was applied to examine gene flow among
308 populations. Utilizing allelic frequency data, the analysis generated an ML tree, followed by
309 inferring the historical migratory events between populations. We assumed n migration events
310 and the calculation of each migration event was done separately. We used the third ddRAD
311 data set and grouped it into 11 populations based on phylogenetic analysis. The R package
312 ‘optM’ (Fitak, 2021) was then used to optimize the number of migration events according to
313 Evanno et al. (2005) (Supplementary Material, section 9). Visualization of the graph results
314 was accomplished by the plot function (plotting_funcs.R) in R. We also employed the ABBA-
315 BABA (D-statistics) and F statistics tests to further investigate patterns of gene flow among
316 populations (Patterson et al., 2012). This analysis provides a basic yet comprehensive
317 framework for deciphering deviation from a strictly bifurcating phylogeny. ABBA-BABA
318 statistics were calculated in DSUITE v.0.4 (Malinsky et al., 2021) using the DTRIOS command
319 for 139 specimens from 11 lineages and two specimens from *D. latus* as outgroup
320 (Supplementary Material, section 9).

321 **Demographic history**

322 The Extended Bayesian Skyline Plot (EBSP) (Heled & Drummond, 2008) was used to
323 separately analyse the demographic changes for each lineage based on the Cyt *b* and COI data
324 for inference of past population dynamics over time. EBSP analyses were implemented in
325 BEAST v.2 under the GTR + G + I (invariant sites) model and a strict molecular clock. We
326 employed the same mutation rate as in the divergence time analysis. We used the plotEBSP R

327 function to draw the skyline plot (Heled, 2015). The *X*-axes indicate time reported in units of
328 thousands of years before present (BP), while the *Y*-axes display mean effective population
329 size (*N_e*) in millions of individuals divided by generation time plotted on a log scale. Moreover,
330 The stairway plot2 (Liu & Fu, 2020) was used for estimating contemporary effective
331 population sizes and changes in effective population size of the *Ligula* species complex over
332 time. SambaR R package was used to convert the second ddRAD data (unlinked SNPs) file
333 into folded SFS files (see details in Supplementary Material, section 10).

334 **Results**

335 **Summary statistics**

336 We compared the genetic diversity of all parasite populations based on the mtDNA (162
337 specimens) and ddRAD (139 specimens) data. The concatenated mtDNA matrix with 162
338 *Ligula* individuals and 801 bp aligned positions (404 bp Cyt *b* and 397 bp COI) was
339 characterized by 535 monomorphic sites and 246 polymorphic sites (23 singletons and 223
340 parsimony informative sites). We also added 62 ND1 sequences (892 bp) to the concatenated
341 data set for all phylogenetic analyses based on ML and BI methods. Estimates of nucleotide
342 diversity for the two mitochondrial genes ranged from 0.0025 to 0.0196 and the haplotype
343 diversity was between 0.61 and 1. The mean observed and expected heterozygosity ranged
344 between 0.0005–0.044 and 0.0023–0.104, respectively. Additionally, populations from China,
345 Australia and New Zealand, and Central and South Africa (CSA) demonstrated the lowest
346 levels of haplotype diversity and observed and expected diversities (except *L. alternans*). The
347 *Ligula* population from Ethiopia (12.8) showed the highest number of private alleles; however,
348 populations in Lineage B, Australia and New Zealand, *L. alternans* and CSA had the fewest
349 private alleles (5.8) (Table 1).

350 **Phylogenetic relationships and species trees of *Ligula* spp**

351 BI and ML analyses of the three mitochondrial genes (1693 bp) generated consistent trees with
352 a considerably well-supported phylogenetic structure (Fig. 2A). Similarly, the reconstructed
353 phylogenies based on ddRAD data (4471 SNP) showed identical topologies in the BI and ML
354 methods (Fig. 2B). The phylogenetic structure revealed maximum support for all lineages in
355 both mtDNA and ddRAD data sets. In all reconstructions, the *Ligula* population from Ethiopia
356 was strongly supported by high posterior probability (pp=1) and bootstrap support (BS=100)
357 as the basal lineage to all the tapeworms. Parasite populations from the Nearctic separated from
358 other lineages with maximal statistical robustness. All phylogenetic trees from mtDNA and
359 ddRAD data disclosed a sister group relationship between populations from EAR (Tanzania
360 and Kenya) and CSA populations. The phylogenetic relationships of the lineages derived from
361 mtDNA data were in agreement with the phylogeny of the ddRAD data (except China and *L.*
362 *pavlovskii* lineages). Within mtDNA, *L. pavlovskii* diverged from Lineage A and *L. alternans*
363 with high statistical support. However, *L. pavlovskii* was distinctly separated from Lineage A,
364 China, *L. alternans*, Lineage B, Australia and New Zealand. To sum up, our results added five
365 new distinct lineages (*L. pavlovskii*, Australia, New Zealand, EAR and CSA) to the six
366 previously defined *Ligula* lineages (Bouzid et al., 2008; Štefka et al., 2009). The SVDquartet
367 species trees recovered the same topology as the ML and BI analyses but with variable posterior
368 probability and bootstrap values (Fig. 2).

369 **Population genetic structure and host usage**

370 A total of 133 haplotypes were found based on the Cyt *b* and COI sequences in 162 individuals.
371 The mitochondrial haplotype network based on TCS analysis with a 95% parsimony
372 connection limit revealed the same 11 distinct haplogroups as in the phylogenetic analysis (Fig.
373 3A, connection limit =12 mutations). Individual haplogroups showed a high level of host
374 specificity. Although many lineages were retrieved from Cypriniformes, the genera (and
375 families) of the fish rarely overlapped between lineages even in areas of sympatry (such as

376 Europe). Haplogroup A is widely distributed in Europe, Iran and Russia and included *Ligula*
377 populations from Cypriniformes (*A. brama*, *R. rutilus*, *B. bjoerkna*, *S. erythrophthalmus*, *S.*
378 *cephalus*, *A. alburnus*, *P. phoxinus* and *C. carassius*). The *L. alternans* haplogroup was
379 connected to haplogroup A with 39 mutational steps and comprised parasite populations from
380 Iran, China, and far east Russia, which were associated with *Hemiculter* (Cypriniformes:
381 Xenocyprididae) and *Neosalanx* (Osmeriformes: Salangidae). Haplogroup B was linked to
382 Australia and China with 30 and 36 mutational steps, respectively. Furthermore, haplotypes
383 from Australia and New Zealand diverged from each other with 41 mutational steps and
384 included parasite populations from Galaxiiformes (*Galaxias*) and Gobiiformes
385 (*Gobiomorphus*). Lineage B and the haplogroup in China were found in Cypriniformes
386 (*Rhodeus*, *Gobio* and *Pseudophoxinus*) and Osmeriformes (*Neosalanx*), respectively. The
387 haplogroup in EAR comprised parasite populations of *E. sardella* that diverged from the
388 haplogroup in CSA (*E. anoplus* and *E. paludinosus*) by 22 mutational steps. The *Ligula*
389 populations from Nearctic Cypriniformes (*S. atromaculatus* from Canada and *R. osculus* from
390 Oregon) split from the other lineages by 55 and 120 mutational steps. Furthermore,
391 haplogroups of *L. pavlovskii* from Gobiiformes (*N. fluviatilis*, *N. melanostomus*, *A. fluviatilis*,
392 *P. minus* and *P. microps*) and Ethiopia from Cypriniformes (*L. intermedius*, *L. tsanenis*, *L.*
393 *brevicephalus* and *E. humilis*) separated from each other by 110 mutational steps (Fig. 3A).
394 Contrary to the mitochondrial haplotype network, the phylogenetic network based on ddRAD
395 data clustered Australia and New Zealand together, revealing 10 well-supported clusters for
396 the *Ligula* species complex (Fig. 3B). Ethiopia was the first lineage branching at the base. EAR
397 and CSA split from each other with 98% bootstrap support value, diverging from *L. pavlovskii*
398 and Ethiopia with 100% bootstrap support value. Similarly, Australia and New Zealand
399 differed from Lineage B. Also, Lineage A demonstrated a divergence from all parasite
400 populations with strong bootstrap value (100%).

401 Population structure analyses showed the three new clusters (*L. pavlovskii*, EAR, CSA) were
402 clearly separated from the five previously identified lineages (Lineage A, *L. alternans*,
403 Australia, Lineage B and China) (Fig. 4). Cross-validation of the admixture analysis revealed
404 an optimal number of K=10 clusters (Fig S2). All lineages were separated from each other,
405 although some admixture was observed between EAR and CSA (Fig. 4A). PCA patterns were
406 mainly analogous to the clustering recognised by admixture. The first two PCs represented
407 22.6% and 18.1% of the total genetic variation, separated into 10 groups. EAR was close to
408 CSA and had an overlap on both PC1 and PC2 axes. The third PC (16%) clearly discriminated
409 Lineage A, Lineage B and Ethiopia from the remaining groups (Fig. 4B). Similarly, the cross-
410 entropy curve of the spatial population genetic structure recovered a likely number of K=10
411 genetic clusters (Fig. S3). We spatially interpolated the ancestry coefficients projected onto a
412 map to describe the global genetic structure in our data (Fig. 4C). The resulting spatial pattern
413 revealed 10 components of ancestry in the sampling space.

414 The analysis of shared ancestry in the fineRADstructure confirmed the results of the cluster
415 analysis with K=10, with significant support in the dendrogram. Although the recent co-
416 ancestry was greatest between EAR and CSA, the two lineages formed distinct groups in the
417 analysis. Moreover, a single sample from New Zealand showed a higher level of shared genetic
418 background with the cluster from Australia. These two clusters had relatively higher co-
419 ancestry with Lineage B and China. Moreover, lineages A and B exhibited a moderate common
420 ancestry with each other, whereas the lowest level of co-ancestry sharing was discovered
421 among Ethiopia, *L. pavlovskii*, Nearctic, EAR and CSA. This is consistent with the admixture
422 results where a level of admixed population was observed between EAR and CSA as well as
423 Australia and Lineage B (Fig. 5).

424 **Species delimitation analyses**

425 The species delimitation methods (ASAP, GMYC, bPTP and BFD) produced estimates that
426 ranged from 10 to 15 species (GMYC and bPTP) (Fig. 6). ASAP analysis with the best score
427 delimited 10 and 12 species based on *Cyt b* (lowest score = 2.5), COI (lowest score = 2.5) and
428 ND1 (lowest score = 4). In the ND1 fragment, out of the 10 top-scoring models, three showed
429 distance thresholds of 5.7%, 1.4% and 2.7% with seven, 13 and 12 species. The third was
430 selected as the most likely initial estimate of species diversity. Tree-based analyses identified
431 the same composition and number of species. The results of the BFD hypothesis testing are
432 shown in Fig. 4, Fig. S6 and Table S4. The ddRAD data set favoured 10 candidate species
433 based on marginal likelihood (3820.21, Fig. S4) and BF (32.5). Overall, all delimitation
434 analyses determined Lineage A, *L. alternans*, *L. pavlovskii*, China and Nearctic as different
435 species. However, GMYC and bPTP over-split Lineage B and Ethiopia into three candidate
436 species. In addition, all the methods (except ASAP based on *Cyt b* and BFD) distinguished a
437 species boundary between Australia and New Zealand.

438 **Divergence time and phylogeographic structure**

439 Estimate of the divergence times using mitochondrial genes was consistent with the time-
440 calibrated tree of ddRAD, strongly supporting the basal cladogenesis between lineages of the
441 *Ligula* species complex in late Miocene and early Pliocene (mtDNA= 4.99 Mya, 95% highest
442 posterior density intervals (HPD): 4.76–5.27 Mya and ddRAD =5.05 Mya, HPD: 4.56–5.49),
443 with the Ethiopian population emerging as the basal lineage to all *Ligula* populations (Fig. 7).
444 Subsequent cladogenetic events dating to mid-Pliocene at approximately 3.35–4.01 Mya
445 (mtDNA=4.01 Mya, HPD= 4.87–3.10 Mya and ddRAD=3.35 Mya, HPD= 3.04–3.78 Mya)
446 isolated the ancestor of Nearctic lineage from other populations. Both mtDNA and ddRAD
447 results revealed a sister relationship between EAR and CSA which diverged from each other
448 in early Pleistocene at 1.01–1.14 Mya (mtDNA= 1.14 Mya, HPD: 0.71–1.41 Mya and ddRAD
449 =1.01 Mya, HPD: 0.66–1.29 Mya). Furthermore, both dated trees strongly supported a sister

450 relationship between Australia and New Zealand which diverged from Lineage B at ~1.12–
451 1.23 Mya (mtDNA= 1.12 Mya, 95% HPD: 0.78–1.81 Mya and ddRAD =1.23 Mya, HPD: 1.11–
452 1.49 Mya). In mtDNA, *L. pavlovskii* diverged from *L. alternans* and Lineage A at ~2.41 Mya
453 (Fig. 7A, HPD: 0.78–2.96 Mya); however, ddRAD revealed that *L. pavlovskii* split from the
454 other six lineages at ~2.48 Mya (Fig. 7B, HPD: 2.29–2.88 Mya). Furthermore, in mtDNA,
455 China diverged from Australia, New Zealand and Lineage B at ~1.71 Mya (Fig. 7A, HPD:
456 1.11–2.17 Mya), while in ddRAD, China formed a sister relationship to *L. alternans*, diverging
457 at ~1.52 Mya (Fig. 7B, HPD: 1.09–1.85 Mya).

458 **Historical biogeography and gene flow analyses**

459 Ancestral range estimation analyses were applied to chronograms resulting from both the
460 mtDNA and ddRAD data sets (Fig. 8A). A comparison of the six biogeographic models with
461 the likelihood ratio test and AIC indicated that DIVALIKE+J was the top fitting model for both
462 the mtDNA and ddRAD data sets (Tables S4). The preferred hypothesis would be that the
463 common ancestor was widespread throughout the Palearctic, Afrotropical and Nearctic
464 biogeographic realms between the late Miocene and early Pliocene. Moreover, vicariance took
465 place between Nearctic and Palearctic in mid-Pliocene. With respect to anagenetic or gradual
466 speciation processes, our parameters indicate that range contraction made an equal contribution
467 as did range expansion in both data sets (Table S4). The ancestor of EAR and CSA emerged
468 through a rare jump dispersal event from the Palearctic to the Afrotropical in late Pliocene. In
469 addition, the China lineage was formed as a result of founder event speciation between
470 Palearctic and Indomalaya in mid-Pleistocene. Finally, Australia and New Zealand were
471 genetically isolated from their ancestor populations in the Palearctic (Fig 8A).

472 Of the three-character models evaluated using sMap, the ER model was the best supported
473 (BIC, Table S5) for both mtDNA and ddRAD data sets. The Bayesian stochastic character

474 mapping of ancestral host fish associations demonstrated an ancient affiliation with
475 cypriniforms at the root of all *Ligula* lineages (Fig 8B). In both data sets, mean marginal
476 posterior estimates showed that cypriniforms contributed as the most recent common ancestral
477 host for the parasite with 81.4% and 82.3% in mtDNA and ddRAD, respectively. Also, the
478 most recent common ancestor (MRCA) of *Ligula* had the highest host switching from
479 cypriniforms to the other fish orders. Furthermore, *Ligula*'s MRCA had the second highest
480 average time spent (mtDNA =9.1%, ddRAD= 12.1%) in Gobiiformes and Galaxiiformes, and
481 the lowest average time spent in Osmeriformes (Table S4, Supporting Material, mtDNA=5.5%
482 and ddRAD=6.1%). A host shift from cypriniforms towards Gobiiformes and Galaxiiformes
483 was discovered in both data sets for the MRCA of Lineage B, Australia and New Zealand. In
484 addition, we identified the occurrence of several host shifts for the MRCA of China and *L.*
485 *alternans*. On the other hand, the mtDNA chronogram revealed the MRCA of *L. pavlovskii*
486 experienced three host shifts from Cypriniformes to Gobiiformes and Osmeriformes, while
487 ddRAD stochastic map detected a host shift between Cypriniformes and Gobiiformes (Fig.
488 8B).

489 **Genome-wide hybridization and introgression**

490 The estimated species tree from Treemix recovered similar topology (Fig. 9A) and relative
491 branch lengths to the phylogenetic trees. The simplest model that described the data was the
492 model that assumed three migrations. No significant migration events were retrieved by more
493 complex models (Fig S4). The results revealed gene flow from Nearctic into *L. alternans* and
494 China populations. Moreover, two gene flow events were detected from Lineage B into China
495 and *L. pavlovskii*. This finding shows how introgression affects phylogenetic relationships in
496 closely related species. To determine possible introgression events, we performed ABBA-
497 BABA tests for 10 trios of closely related phylogroups. D statistics for 47 of the trios were
498 significantly different from zero ($Z > 3$, p-adjusted $<.000919$) and the values ranged between

499 0.244 and 0.492 (Table S6). Based on the sliding window analysis, SNPs demonstrating the
500 ABBA-BABA patterns for each trio were distributed across 25–29 different contigs (Table S3).
501 Nevertheless, it is possible that a single gene flow event leads to multiple elevated f_4 -ratio and
502 D results. When we calculated a branch-specific statistic $fb(C)$ that partially disentangled the
503 correlated f_4 -ratio results, we found that only two out of the eight branches exhibited significant
504 excess sharing of derived alleles at least with one other lineage (Fig. 10B). An internal branch
505 that represented Lineage A and China exhibited an excess sharing of derived alleles with
506 Australia, New Zealand, *L. pavlovskii* and Lineage B. Another $fb(C)$ signal involves an internal
507 branch as the common ancestor of Lineage B with *L. pavlovskii*, EAR and CSA (Fig. 9B).
508 These results could also be attributed to incomplete lineage sorting. It must be noted that even
509 a single introgression event may result in significant $fb(C)$ values in multiple related
510 phylogroups (Malinsky et al., 2018).

511 **Historical and contemporary demographics**

512 The demographic history based on two mtDNA genes (Cyt *b* and COI) and ddRAD data sets
513 is presented for nine parasite lineages in Fig. 10. Two lineages (*L. alternans* and New Zealand)
514 were not plotted due to their low effective sample sizes ($ESS < 200$) and low sample sizes. EBSP
515 based on the mtDNA suggested that demographic expansion took place at approximately 100
516 kya (thousand years ago) for lineages A and B (Fig. 10 A and B). Moreover, the stairway plot
517 based on the ddRAD data showed a clear sign of ancestral bottleneck between 1.5 kya and 2
518 kya followed by a range expansion for these lineages. China showed a population expansion
519 after experiencing a bottleneck between 4 kya and 8 kya. Australia demonstrated a trend
520 indicative of recent population shrinkage. Our mtDNA results uncovered a rapid population
521 expansion of approximately 100 kya for *L. pavlovskii*; however, ddRAD revealed a bottleneck
522 at ~50 kya with a decreased pattern from 6 kya onwards. A stable population size is indicated

523 for EAR and CSA during the last 3 kya. In addition, a recent population reduction and a strong
524 signal of demographic expansion were detected for the Nearctic and Ethiopian lineages.

525 **Discussion**

526

527 Host specificity, an inherent feature of parasitic and symbiotic organisms, can be a product of
528 co-diversification (Engelstädter & Fortuna, 2019), but it is often the result of a more convoluted
529 evolutionary history including host switches and dispersal events (Hoberg & Brooks, 2008;
530 Matthews et al., 2022). The latter should be peculiar to parasites where the host lineages are
531 much older than the parasite lineages, such as the *L. intestinalis* species complex. Using our
532 example of a speciating parasite lineage with a global distribution, we demonstrated how host
533 specificity of individual *Ligula* lineages was formed by different historical events involving
534 long-distance dispersals, host switches and occasional introgression. Our analyses based on a
535 dense SNP matrix and mtDNA data provided a clear picture of the cryptic diversity within the
536 *L. intestinalis* species complex and a robust time-calibrated phylogeography. By analysing
537 biogeographic and demographic patterns of individual lineages, we revealed patterns of the
538 parasite's speciation and dispersal throughout its distribution range. Below, we discuss the
539 results focusing on individual factors contributing to the parasite's diversification.

540 **Phylogenetic relationships and occasional mito-nuclear discordance**

541 *Ligula* phylogenies based on ddRAD and mtDNA data revealed 11 distinct lineages and
542 showed a generally well-supported phylogenetic structure for the *Ligula* species complex. We
543 characterized several new lineages from previously unexplored areas (CSA, EAR) or lineages
544 that were not analysed in the global context before (*L. pavlovskii*, New Zealand; see Lagrue et
545 al., 2018; Vitál et al., 2021). In addition, several differences in the phylogeny of the group were
546 recovered compared to previous studies based on fewer loci and less material (Bouزيد et al.,

547 2008; Štefka et al., 2009). Ethiopia formed a basal lineage to other populations in our analysis,
548 contrary to the findings of the previous study (Bouzid et al., 2008) where Nearctic samples
549 (from Canada and Mexico) were at the basal position. Our dataset also allowed for a more fine
550 scale distinction of the lineages previously clustered with Lineage B (Bouzid et al., 2008).

551 Only two cases of discordance between mitochondrial and nuclear-ddRAD phylogenies were
552 found, with *L. pavlovskii* and China showing different topologies in each data set. To exclude
553 a possibility that the incongruence is only due to the lack of signal in the smaller mtDNA
554 dataset, we performed a congruence test between mtDNA and ddRAD topologies and found
555 the topology differences were significant (AU and SH tests). Similar mito-nuclear phylogenetic
556 discordances have also been reported in several other metazoan taxa (e.g. Bouzid et al., 2008;
557 Shaw, 2002; Weigand et al., 2017), often credited to the molecular signature of selection
558 (Morales et al., 2015), incomplete lineage sorting (Degnan & Rosenberg, 2009) or introgressive
559 hybridization (Bisconti et al., 2018). Our analyses of historical gene flow indicated past
560 introgression events between multiple lineages (see the discussion below). Thus, we suggest
561 introgressions as the mechanism responsible for the emergence of mito-nuclear discordances
562 for China and *L. pavlovskii* lineages.

563 **Species delimitation reveals extensive cryptic diversity in *L. intestinalis* s.l. populations**

564 The taxonomy of *Ligula* tapeworms has always been complicated due to the general uniformity
565 of their plerocercoids and adults (Dubinina, 1980), resulting in a lack of suitable morphological
566 features to determine species boundaries using traditional methods, except for an already
567 known trait, the duplicated reproductive complexes in *L. alternans* (Luo et al., 2003). Indeed,
568 numerous species of *Ligula* have been proposed to accommodate plerocercoids from different
569 fish hosts (see Dubinina, 1980 for their extensive list), but almost all these taxa have been
570 synonymised with *L. intestinalis*. This species was then considered as a euryxenous parasite of

571 cyprinoid fishes, maturing in several fish-eating birds (Kuchta & Scholz, 2017; Li et al., 2018;
572 Petkevičiūtė, 1992).

573 Here, we used a range of species delimitation analyses to identify the putative species in the
574 *Ligula* complex. The results based on ddRAD data revealed a slightly lower number of
575 candidate species than the non-recombining mtDNA. Our findings also indicated that although
576 different methods of species delimitation produced highly consistent species boundaries, they
577 were not entirely similar. Carstens et al. (2013) argued that such incongruence either signifies
578 a difference in the power of one or more of the methods to detect cryptic lineages or that
579 assumptions of the species delimitation approach have been violated. They suggested that
580 species delimitation inferences should be drawn cautiously in order to avoid delimiting entities
581 that fail to accurately represent evolutionary lineages. Therefore, taking the conservative
582 approach, we identified at least 10 putative species for the *Ligula* species complex as follows:
583 (1) *L. intestinalis* Lineage A; (2) *L. alternans* (syn. *Digramma alternans*); (3) *L. intestinalis*
584 Lineage B; (4) *Ligula* sp. 1 from Australia and New Zealand (possibly two species lineages);
585 (5) *L. pavlovskii*; (6) *Ligula* sp. 2 from China; (7) *Ligula* sp. 3 from EAR; (8) *Ligula* sp. 4 from
586 CSA (9); *Ligula* sp. 5 from the Nearctic; and (10) *Ligula* sp. 6 from Ethiopia. In subsequent
587 analyses, we contrasted these evolutionarily significant lineages with their host and geographic
588 spectra and demographic histories.

589 **Dated phylogeography reveals historical dispersal and vicariance due to climatic** 590 **oscillations and host switching**

591 Climatic fluctuations have shaped the patterns of divergence in a multitude of species (Hewitt,
592 2001). In particular, quaternary glaciation cycles and climatic oscillations had a major impact
593 on both free-living and parasitic species, leading to the extinction of several parasites (Hoberg
594 et al., 2012; Li et al., 2018) and influencing their evolution and distribution (e.g. Galbreath
595 et al., 2020; Nazarizadeh et al., 2022b). Based on our mtDNA and ddRAD molecular dating,

596 the ancestor of *Ligula* is estimated to have appeared between the late Miocene and early
597 Pliocene (Fig. 7), and was found to parasitise one of the early diverging cyprinid species
598 (*Barbus*, of the subfamily Barbinae). The divergence time suggests that the parasite complex
599 did not follow the evolution of its intermediate host as the divergence of Cypriniformes had
600 already occurred during the late Jurassic (154 Mya) (Tao et al., 2019). Moreover, the historical
601 biogeography model revealed that the divergence of parasite populations from Afrotropical,
602 Nearctic and Palearctic realms was recovered as two vicariant events (Fig. 8). This allopatric
603 speciation could be associated with the isolation of migratory waterbirds due to the
604 biogeographical realms becoming isolated by major barriers. According to our results, the first
605 vicariance occurred between Afrotropical, Palearctic and Nearctic regions at approximately 5
606 Mya (Fig. 8) when desertification increased and the decline of global temperatures accelerated,
607 resulting in the development of the Northern Hemisphere ice sheets and the onset of the ice
608 ages. Moreover, the considerable expansion of the Sahara Desert had by then impeded trans-
609 Saharan migration and long-distance dispersal for several migratory waterbirds (Bruderer &
610 Salewski, 2008).

611 *Ligula* plerocercoids mature very quickly into adults in bird guts following ingestion of an
612 infected fish (Loot et al., 2001). The adult phase lasts up to five days (Dubinina, 1980), during
613 which the eggs can be dispersed. Consequently, a lack of suitable freshwater lake habitats
614 within a few days flight of the final host represents a strong barrier against gene flow of the
615 parasite, as documented by the lack of gene flow in Lineage A between Europe and a recently
616 introduced population in North Africa (Bouzig et al., 2013; Štefka et al., 2009). Thus,
617 aridification of the environment accompanied by reduced or ceased migration of the definitive
618 host likely caused the major vicariance mechanisms in the evolution of *Ligula*.

619 The second vicariant event took place between the Nearctic and Palearctic during the late-
620 Pliocene when the Nearctic lineage diverged from the other populations (Fig. 8). This lineage
621 was recovered from the Nearctic leuciscids of two subfamilies (*Semotilus* - Plagopterinae, and
622 *Rhinichthys* - Pogonichthyinae) which are thought to have originated in ancient Europe and
623 dispersed to North America during the Late Cretaceous to Paleocene (see Imoto et al., 2013).
624 In contrast to the intermediate hosts which had evolved a long time before *Ligula*'s speciation,
625 changes in the behaviour of the definitive hosts could have accelerated parasite's
626 diversification from Pliocene to Pleistocene (Hirase et al., 2016). It has been proposed that
627 glaciations possibly shifted the migratory routes and migratory behaviour of several North
628 American bird species (Zink & Gardner, 2017). In addition, ecological competition and
629 physical isolating barriers between the Palearctic and Nearctic during the late Pliocene or early
630 Pleistocene were determining factors (Naughton, 2003). On the other hand, the results of
631 TreeMix showed historical gene flow between the population from the Nearctic and the MRCA
632 of lineages of China and *L. alternans* (Fig. 9A). This introgression likely occurred after the
633 second vicariant event when both the Nearctic and Palearctic regions were connected through
634 the Bering Land Bridge, which allowed for a passage of many species between the two realms
635 (Beaudoin & Reintjes, 1950).

636 Founder event speciation is a form of allopatric speciation in which a small number of
637 individuals establishes a new population beyond the existing range of the main population (
638 Matzke, 2014). The results of the historical biogeographic model revealed that a jump dispersal
639 event took place from the Palearctic to the Afrotropical in the early Pleistocene, forming the
640 MRCA of EAR and CSA (Fig. 8). In line with the present study, several fossil records of
641 waterbirds have been discovered from the early Pleistocene in EAR, the Upper Pliocene in
642 Ethiopia as well as Miocene deposits in Kenya (Brodkorb & Mourer-Chauviré, 1982; Dyke &
643 Walker, 2008; Prassack et al., 2018). Our results do not support the theory of *Ligula*

644 colonization of Africa in recent decades (Gutiérrez & Hoole, 2021; Kihedu et al., 2001).
645 Instead, we suggest that the parasite had already colonized sub-Saharan Africa in the early
646 Pleistocene. Moreover, the dated tree based on both mtDNA and ddRAD data indicated that
647 parasite populations from EAR and CSA diverged from each other in the late Pleistocene (Fig.
648 7). EAR lineage showed host specificity to *Engraulicypris sardella* and *Rastrineobola*
649 *argentea*, whereas CSA lineage to barbels (*Enteromius anoplus*, *E. paludinosus*, *E.*
650 *trimaculatus* and *Barbus neefi*). Whilst barbels are a common host for *Ligula* in many regions
651 (Barson & Marshall, 2003; Emaminew et al., 2014) and represent one of its ancestral hosts, a
652 switch to the hosts endemic to the EAR region could have played a role in the differentiation
653 of the EAR lineage.

654 Our biogeographic analyses also suggested the occurrence of sympatric speciation in
655 *Ligula*. The historical biogeographic model revealed that three lineages (Lineage A, Lineage
656 B and *L. pavlovskii*) were formed as a result of two sympatric speciation events in the western
657 Palearctic. It also revealed that the MRCA of these lineages were historically distributed in the
658 Palearctic (Fig. 8A and B). Moreover, some samples from these lineages were found in the
659 same waterbodies. Lineages A and B were found in two different groups of cyprinid fish and
660 *L. pavlovskii* was discovered in gobies, i.e., members of the distantly related fish order
661 Gobiiformes. Similarly, Bouzid et al. (2008) revealed that the Euro-Mediterranean lineages
662 (lineages A and B) live in sympatry and infect the same definitive host. They also suggested
663 that reproductive isolation formed a significant genetic barrier between the two lineages.
664 Moreover, a local genetic population study on *Ligula* provides preliminary evidence of a host-
665 related ecological differentiation in Lineage A at an intra-lineage level (Nazarizadeh et al.,
666 2022a). Reproductive isolation in sympatry is mainly a consequence of the interaction between
667 various pre- and post-zygotic barriers (Nosil, 2012). The proximate mechanisms of
668 differentiation in sympatric lineages of *Ligula* are yet to be explored; however, for instance,

669 Henrich & Kalbe (2016) indicated that postzygotic ecological selection is more likely the cause
670 of restricted gene flow between *Schistocephalus solidus* and *S. pungitii*, sister species of
671 tapeworms related to *Ligula*.

672 The phenomena of geographical dispersal and host-switching are common in parasites at
673 different spatial and temporal scales (Hoberg & Brooks, 2008). Cyclical and episodic climatic
674 changes are predicted to influence ecological release and the response of populations to the
675 relaxation of isolation mechanisms that increase host-switching potential (e.g. Brooks et al.,
676 2006; Hoberg et al., 2002). Through host-switching, a small proportion of a species moves into
677 a new geographical area, which may be followed by speciation through peripheral isolation
678 and addition of a new species to the parasite's host range (Huysse et al., 2005). The results of
679 divergence times and historical biogeography confirmed that the latest "jumping speciation" in
680 *Ligula* occurred from the Palearctic to Indomalayan and Australian realms in the mid-
681 Pleistocene (approximately 2.0 to 1.2 mya). The host switch from Cypriniformes to native
682 freshwater fish (Osmeriformes in China, Galaxiiformes and Gobiiformes in Australia and New
683 Zealand) led to "peripheral isolate speciation" (Frey, 1993), a pattern previously reported in
684 several cestode species (Brooks, 1989; Zarlenga et al., 2006).

685 **Genome-wide hybridization and introgression**

686 Results of the TreeMix introgression and the f-branch tests were highly consistent, with the f-
687 branch statistics revealing a higher number of gene flow events. Introgression results suggested
688 the presence of several ancestral ghost lineages between Ethiopia and other lineages (Fig. 10B).
689 Ghost lineages, indicative of the existence of unsampled or extinct lineages in a species,
690 significantly influence gene flow detection (Tricou et al., 2022). TreeMix detects introgression
691 events based on the known lineages and branches in a tree and does not consider ghost lineages
692 (Tricou et al., 2022).

693 Historical and contemporary patterns of hybridization have been documented for many
694 tapeworms (Bello et al., 2021; Easton et al., 2020; Landeryou et al., 2022). Henrich and Kalbe
695 (2016) confirmed natural hybridization between *S. solidus* and *S. pungitii* from both allopatric
696 and sympatric populations. Our results detected historical hybridization in both sympatric
697 (Lineage A, Lineage B and *L. pavlovskii*) and allopatric lineages (Nearctic, China, Lineage A,
698 Australia and New Zealand). In addition, we discovered significant positive D-statistic values,
699 indicating an excess of derived allele sharing between China and Lineage B, in line with the
700 TreeMix migration edge (Fig. 10B). We suggest that historical introgressive hybridization
701 might have enabled *Ligula* to infect a broader range of host species (Osmeriformes,
702 Gobiiformes and Galaxiiformes), similar to adaptation of hybrids of nonparasitic taxa to novel
703 ecological conditions where their parents could not persist (Thaenkham et al., 2022). However,
704 future testing of specific questions related to adaptive introgression will require extended
705 sampling and a well-resolved and annotated genome assembly.

706 **Demographic history**

707 Demographic analyses based on mtDNA and ddRAD data showed mutually consistent results
708 for most lineages, with SNP based results providing a more recent picture (Fig. 11). EBSP
709 showed Lineage A and Lineage B experienced rapid range expansions (Fig. 11A) during the
710 Last Interglacial Period (LIG; Otto-Bliesner et al., 2006). Moreover, during the Medieval
711 Warm Period (MWP; Crowley & Lowery, 2000), these lineages exhibited rapid expansions
712 following a bottleneck event (Fig. 11B). Although these very recent events are not
713 straightforward to explain without more comprehensive data and analyses, they fit into the
714 general picture of a volatile population history expected for parasitic taxa, involving
715 bottlenecks and expansions following host-switches as well as expansions due to post-glacial
716 host range expansions. Periods of geographical and host range expansions allow parasites to
717 explore their capacity for generalism, while periods of isolation force parasites to specialise on

718 specific hosts. This specialisation will modify the “sloppy fitness space” (Agosta & Klemens,
719 2008; Araujo et al., 2015) of the parasite due to the parasite adapting to local environments.
720 Bottlenecks found in *Ligula* lineages, particularly those associated with lineage formation,
721 could be initiated by host switches followed by population expansion in case of a successful
722 specialisation. During a host switch, it is likely that the host immune system can decrease
723 overall parasite load, while at the same time it selects for more immune-resistant parasite’s
724 genotypes (Levin et al., 1999). *Ligula* interacts with the immune and metabolic systems of the
725 fish host intensively, leading to the castration of the host and redirecting energy resources
726 towards the parasite’s growth (Yoneva et al., 2015). Future analysis of coding genetic
727 differences and dating their origin, ideally using whole genome data, could allow testing the
728 hypotheses of host-switch associated bottlenecks.

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736 **References**

- 737 Aberer, A. J., Kobert, K., & Stamatakis, A. (2014). ExaBayes: massively parallel Bayesian
738 tree inference for the whole-genome era. *Molecular Biology and Evolution*, *31*(10),
739 2553–2556. doi:10.1093/molbev/msu236
- 740 Agosta, S. J., & Klemens, J. A. (2008). Ecological fitting by phenotypically flexible
741 genotypes: implications for species associations, community assembly and evolution.
742 *Ecology Letters*, *11*(11), 1123–1134. doi:10.1111/j.1461-0248.2008.01237.x
- 743 Alexander, D. H., Novembre, J., & Lange, K. (2009). Fast model-based estimation of
744 ancestry in unrelated individuals. *Genome Research*, *19*(9), 1655–1664.
745 doi:10.1101/gr.094052.109
- 746 Andrews, S. (2010). FastQC: a quality control tool for high throughput sequence data.
747 Babraham Bioinformatics, Babraham Institute, Cambridge, United Kingdom. Available
748 online at: <http://www.bioinformatics.babraham.ac.uk/projects/>

- 749 fastqc
- 750 Angst, P., Ebert, D., & Fields, P. D. (2022). Demographic history shapes genomic variation
751 in an intracellular parasite with a wide geographical distribution. *Molecular Ecology*,
752 *31*(9), 2528–2544. doi: 10.1111/mec.16419
- 753 Araujo, S. B. L., Braga, M. P., Brooks, D. R., Agosta, S. J., Hoberg, E. P., von Hartenthal, F.
754 W., & Boeger, W. A. (2015). Understanding host-switching by ecological fitting. *PLoS*
755 *One*, *10*(10), e0139225. doi: 10.1371/journal.pone.0139225
- 756 Barson, M., & Marshall, B. E. (2003). The occurrence of the tapeworm *Ligula intestinalis*
757 (L.), in *Barbus paludinosus* from a small dam in Zimbabwe. *African Journal of Aquatic*
758 *Science*, *28*(2), 175–178. doi: 10.2989/16085910309503782
- 759 Beaudoin, A. B., & Reintjes, F. D. (1950). Occasional Paper: Late Quaternary Studies in
760 Beringia and Beyond, 1950-1993: *An Annotated Bibliography*.
- 761 Bello, E., Palomba, M., Webb, S. C., Paoletti, M., Cipriani, P., Nascetti, G., & Mattiucci, S.
762 (2021). Investigating the genetic structure of the parasites *Anisakis pegreffii* and *A.*
763 *berlandi* (Nematoda: Anisakidae) in a sympatric area of the southern Pacific Ocean
764 waters using a multilocus genotyping approach: first evidence of their interspecific
765 hybridization. *Infection, Genetics and Evolution*, *92*, 104887. doi:
766 10.1016/j.meegid.2021.104887
- 767 Bianchini, G., & Sánchez-Baracaldo, P. (2021). sMap: Evolution of independent, dependent
768 and conditioned discrete characters in a Bayesian framework. *Methods in Ecology and*
769 *Evolution*, *12*(3), 479–486. doi: 10.1111/2041-210X.13540
- 770 Bisconti, R., Porretta, D., Arduino, P., Nascetti, G., & Canestrelli, D. (2018). Hybridization
771 and extensive mitochondrial introgression among fire salamanders in peninsular Italy
772 OPEN. *Scientific Reports*, *8*, 13187. doi: 10.1038/s41598-018-31535-x
- 773 Bothma, J. C., Matthee, S., & Matthee, C. A. (2020). The evolutionary history of parasitic
774 sucking lice and their rodent hosts: a case of evolutionary co-divergences. *Zoologica*
775 *Scripta*, *49*(1), 72–85. doi: 10.1111/zsc.12389
- 776 Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., Suchard, M. A.,
777 Rambaut, A., & Drummond, A. J. (2014). BEAST 2: a software platform for Bayesian
778 evolutionary analysis. *PLoS Computational Biology*, *10*(4), e1003537. doi:
779 10.1371/journal.pcbi.1003537
- 780 Bouzid, W., Lek, S., Mace, M., Ben Hassine, O., Etienne, R., Legal, L., & Loot, G. (2008).
781 Genetic diversity of *Ligula intestinalis* (Cestoda: Diphylobothriidea) based on analysis
782 of inter-simple sequence repeat markers. *Journal of Zoological Systematics and*
783 *Evolutionary Research*, *46*(4), 289–296. doi: 10.1111/j.1439-0469.2008.00471.x.
- 784 Bouzid, W., Štefka, J., Bahri-Sfar, L., Beerli, P., Loot, G., Lek, S., Haddaoui, N., Hypša, V.,
785 Scholz, T., & Dkhil-Abbes, T. (2013). Pathways of cryptic invasion in a fish parasite
786 traced using coalescent analysis and epidemiological survey. *Biological Invasions*,
787 *15*(9), 1907–1923. doi: 10.1007/s10530-013-0418-y.

- 788 Bouzid, Wafa, Štefka, J., Hypša, V., Lek, S., Scholz, T., Legal, L., Hassine, O. K. Ben, &
789 Loot, G. (2008). Geography and host specificity: Two forces behind the genetic
790 structure of the freshwater fish parasite *Ligula intestinalis* (Cestoda:
791 Diphyllbothriidae). *International Journal for Parasitology*, 38(12), 1465–1479. doi:
792 10.1016/j.ijpara.2008.03.008.
- 793 Brodkorb, P., & Mourer-Chauviré, C. (1982). Fossil Anhingas (Aves: Anhingidae) from Early
794 Man Sites of Hadar and Omo (Ethiopia) and Olduvai Gorge (Tanzania). *Geobios*, 15(4),
795 505–515. doi:10.1016/S0016-6995(82)80071-5.
- 796 Brooks, D. R. (1989). A summary of the database pertaining to the phylogeny of the major
797 groups of parasitic platyhelminths, with a revised classification. *Canadian Journal of*
798 *Zoology*, 67(3), 714–720. doi: 10.1139/z89-103.
- 799 Brooks, D. R., León-Règagnon, V., McLennan, D. A., & Zelmer, D. (2006). Ecological
800 fitting as a determinant of the community structure of platyhelminth parasites of
801 anurans. *Ecology*, 87(sp7), S76–S85. doi: 10.1890/0012-
802 9658(2006)87[76:EFAADO]2.0.CO;2.
- 803 Bruderer, B., & Salewski, V. (2008). Evolution of bird migration in a biogeographical
804 context. In *Journal of biogeography* (Vol. 35, Issue 11, pp. 1951–1959). Wiley Online
805 Library. doi: 10.1111/j.1365-2699.2008.01992.x.
- 806 Bryant, D., Bouckaert, R., Felsenstein, J., Rosenberg, N. A., & RoyChoudhury, A. (2012).
807 Inferring Species Trees Directly from Biallelic Genetic Markers: Bypassing Gene Trees
808 in a Full Coalescent Analysis. *Molecular Biology and Evolution*, 29(8), 1917–1932. doi:
809 10.1093/molbev/mss086.
- 810 Capella-Gutiérrez, S., Silla-Martínez, J. M., & Gabaldón, T. (2009). trimAl: a tool for
811 automated alignment trimming in large-scale phylogenetic analyses. *Bioinformatics*,
812 25(15), 1972–1973. doi: 10.1093/bioinformatics/btp348.
- 813 Carstens, B. C., Pelletier, T. A., Reid, N. M., & Satler, J. D. (2013). How to fail at species
814 delimitation. *Molecular Ecology*, 22(17), 4369–4383. doi: 10.1111/mec.12413
- 815 Caye, K., Jay, F., Michel, O., & François, O. (2018). Fast inference of individual admixture
816 coefficients using geographic data. *The Annals of Applied Statistics*, 12(1), 586–608.
817 doi: 10.1214/17-AOAS1106
- 818 Chang, C. C., Chow, C. C., Tellier, L. C. A. M., Vattikuti, S., Purcell, S. M., & Lee, J. J.
819 (2015). Second-generation PLINK: rising to the challenge of larger and richer datasets.
820 *GigaScience*, 4(1), s13742-015-0047–0048. doi: 10.1186/s13742-015-0047-8
- 821 Chapman, A., Hobbs, R. P., Morgan, D. L., & Gill, H. S. (2006). Helminth parasitism of
822 *Galaxias maculatus* (Jenyns 1842) in southwestern Australia. *Ecology of Freshwater*
823 *Fish*, 15(4), 559–564. doi: 10.1111/j.1600-0633.2006.00198.x.
- 824 Chifman, J., & Kubatko, L. (2014). Quartet Inference from SNP Data Under the Coalescent
825 Model. *Bioinformatics*, 30(23), 3317–3324. doi: 10.1093/bioinformatics/btu530.

- 826 Crowley, T. J., & Lowery, T. S. (2000). How warm was the medieval warm period? *AMBIO:*
827 *A Journal of the Human Environment*, 29(1), 51–54. doi: 10.1639/0044-
828 7447(2000)029[0051:HWWTMW]2.0.CO;2
- 829 de Jong, M. J., de Jong, J. F., Hoelzel, A. R., & Janke, A. (2021). SambaR: An R package for
830 fast, easy and reproducible population-genetic analyses of biallelic SNP data sets.
831 *Molecular Ecology Resources*, 21(4), 1369–1379. doi: doi: 10.1111/1755-0998.13339
- 832 Degnan, J. H., & Rosenberg, N. A. (2009). Gene tree discordance, phylogenetic inference and
833 the multispecies coalescent. *Trends in Ecology & Evolution*, 24(6), 332–340. doi:
834 10.1016/J.TREE.2009.01.009
- 835 Ding, F., Gu, S., Yi, M.-R., Yan, Y.-R., Wang, W.-K., & Tung, K.-C. (2022). Demographic
836 history and population genetic structure of *Anisakis pegreffii* in the cutlassfish
837 *Trichiurus japonicus* along the coast of mainland China and Taiwan. *Parasitology*
838 *Research*, 121(10), 2803–2816. doi: 10.1007/s00436-022-07611-7.
- 839 Dobson, A., Lafferty, K. D., Kuris, A. M., Hechinger, R. F., & Jetz, W. (2008). Homage to
840 Linnaeus: How many parasites? How many hosts? *Proceedings of the National*
841 *Academy of Sciences of the United States of America*, 105(SUPPL. 1), 11482–11489.
842 doi: 10.1073/pnas.0803232105
- 843 Doyle, S. R., Søre, M. J., Nejsun, P., Betson, M., Cooper, P. J., Peng, L., Zhu, X.-Q.,
844 Sanchez, A., Matamoros, G., & Sandoval, G. A. F. (2022). Population genomics of
845 ancient and modern *Trichuris trichiura*. *Nature Communications*, 13(1), 1–12. doi:
846 s41467-022-31487-x
- 847 Drummond, A. J., Suchard, M. A., Xie, D., & Rambaut, A. (2012). Bayesian Phylogenetics
848 with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution*, 29(8), 1969–1973.
849 doi: 10.1093/molbev/mss075
- 850 Dubinina, M. N. (1980). Tapeworms (Cestoda, Ligulidae) of the fauna of the USSR. Amerind
851 Publishing Company, Delhi, 320 pp.
- 852 Dyke, G. J., & Walker, C. A. (2008). New records of fossil ‘waterbirds’ from the Miocene of
853 Kenya. *American Museum Novitates*, 2008(3610), 1–10. doi: 10.1206/0003-
- 854 Easton, A., Gao, S., Lawton, S. P., Bennuru, S., Khan, A., Dahlstrom, E., Oliveira, R. G.,
855 Kepha, S., Porcella, S. F., & Webster, J. (2020). Molecular evidence of hybridization
856 between pig and human *Ascaris* indicates an interbred species complex infecting
857 humans. *Elife*, 9, e61562. doi: 10.7554/eLife.61562.
- 858 Emamineh, T., Dereje, B., & Abdu, M. (2014). Prevalence of *Ligula intestinalis* larvae in
859 *Barbus* fish genera at Lake Tana, Ethiopia. *World Journal of Fish and Marine Sciences*,
860 6(5), 408–416.
- 861 Engelstädter, J., & Fortuna, N. Z. (2019). The dynamics of preferential host switching: Host
862 phylogeny as a key predictor of parasite distribution. *Evolution*, 73(7), 1330–1340. doi:
863 10.1111/evo.13765.

- 864 Evanno, G., Regnaut, S., & Goudet, J. (2005). Detecting the number of clusters of individuals
865 using the software STRUCTURE: a simulation study. *Molecular Ecology*, *14*(8), 2611–
866 2620. doi: 10.1111/j.1365-294X.2005.02553.x.
- 867 Fitak, R. R. (2021). OptM: estimating the optimal number of migration edges on population
868 trees using Treemix. *Biology Methods and Protocols*, *6*(1), bpab017. doi:
869 10.1093/biomethods/bpab017.
- 870 Frey, J. K. (1993). Modes of peripheral isolate formation and speciation. *Systematic Biology*,
871 *42*(3), 373–381.
- 872 Gabagambi, N. P., Salvanes, A.-G. V, Midtøy, F., & Skorping, A. (2019). The tapeworm
873 *Ligula intestinalis* alters the behavior of the fish intermediate host *Engraulicypris*
874 *sardella*, but only after it has become infective to the final host. *Behavioural Processes*,
875 *158*, 47–52. doi: 10.1016/j.beproc.2018.11.002.
- 876 Galbreath, K. E., Toman, H. M., Li, C., & Hoberg, E. P. (2020). When parasites persist:
877 tapeworms survive host extinction and reveal waves of dispersal across Beringia.
878 *Proceedings of the Royal Society B*, *287*(1941), 20201825. doi: 10.1098/rspb.2020.1825.
- 879 Gutiérrez, J. S., & Hoole, D. (2021). *Ligula intestinalis*. *Trends in Parasitology*, S1471-4922.
880 doi:10.1016/j.pt.2021.09.005
- 881 Hafner, M., & Nadler, S. A. (2017). Society of Systematic Biologists Cospeciation in Host-
882 Parasite Assemblages : Comparative Analysis of Rates of Evolution and Timing of
883 Cospeciation Events Author (s): Mark S . Hafner and Steven A . Nadler Published by :
884 Taylor & Francis , Ltd . for the S. *Systematic Zoology*, *39*(3), 192–204. doi:
885 10.2307/2992181.
- 886 Haukisalmi, V., Hardman, L. M., Fedorov, V. B., Hoberg, E. P., & Henttonen, H. (2016).
887 Molecular systematics and Holarctic phylogeography of cestodes of the genus
888 *Anoplocephaloides* Baer, 1923 ss (Cyclophyllidae, Anoplocephalidae) in lemmings (*Lemmus*,
889 *Synaptomys*). *Zoologica Scripta*, *45*(1), 88–102. doi:10.1111/zsc.12136.
- 890 Heled, J., & Drummond, A. J. (2008). Bayesian inference of population size history from
891 multiple loci. *BMC Evolutionary Biology*, *8*(1), 289. doi: 10.1186/1471-2148-8-289
- 892 Heled, Joseph. (2015). Extended Bayesian skyline plot tutorial for BEAST 2. WWW
893 document] URL [http://evomics.org. wpengine. netdna-cdn. com/wp-content](http://evomics.org/wpengine.netdna-cdn.com/wp-content)
- 894 Henrich, T., & Kalbe, M. (2016). The role of prezygotic isolation mechanisms in the
895 divergence of two parasite species. *BMC Evolutionary Biology*, *16*(1), 1–10.
896 doi:10.1186/s12862-016-0799-5.
- 897 Hewitt, G. M. (2001). Speciation, hybrid zones and phylogeography—or seeing genes in
898 space and time. *Molecular Ecology*, *10*(3), 537–549. doi:10.1046/j.1365-
899 294x.2001.01202.x.
- 900 Hirase, S., Yokoyama, Y., Lee, C.-T., & Iwasaki, W. (2016). The Pliocene-Pleistocene
901 transition had dual effects on North American migratory bird speciation.

- 902 *Palaeogeography, Palaeoclimatology, Palaeoecology*, 462, 85–91.
903 doi:10.1016/j.palaeo.2016.09.006
- 904 Hoberg, E. P., & Brooks, D. R. (2008). A macroevolutionary mosaic: episodic host-
905 switching, geographical colonization and diversification in complex host–parasite
906 systems. *Journal of Biogeography*, 35(9), 1533–1550. doi:10.1111/j.1365-
907 2699.2008.01951.x.
- 908 Hoberg, E. P., & Brooks, D. R. (2013). Episodic processes, invasion, and faunal mosaics
909 in evolutionary and ecological time. In: *The Balance of Nature and Human*
910 *Impact* (ed. Rohde K), pp. 199–213. *Cambridge University Press*. doi:
911 10.1017/CBO9781139095075.021
- 912 Hoberg, E. P., Galbreath, K. E., Cook, J. A., Kutz, S. J., & Polley, L. (2012). Northern Host–
913 Parasite Assemblages: History and Biogeography on the Borderlands of Episodic
914 Climate and Environmental Transition. In D. Rollinson & S. I. B. T.-A. in P. Hay (Eds.),
915 *Advances in Parasitology* (Vol. 79, pp. 1–97). Academic Press. doi: doi: 10.1016/B978-
916 0-12-398457-9.00001-9
- 917 Hoberg, E. P., Kutz, S. J., Nagy, J., Jenkins, E., Elkin, B., Branigan, M., & Cooley, D.
918 (2002). *Protostrongylus stilesi* (Nematoda: Protostrongylidae): Ecological isolation and
919 putative host-switching between Dall’s sheep and muskoxen in a contact zone.
920 *Comparative Parasitology*, 69(1), 1–9. doi: 10.1654/1525-
921 2647(2002)069[0001:PSNPEI]2.0.CO;2.
- 922 Hoole, D., Carter, V., & Dufour, S. (2010). *Ligula intestinalis* (Cestoda: Pseudophyllidea): an
923 ideal fish-metazoan parasite model? *Parasitology*, 137(3), 425–438. doi:
924 10.1017/S0031182010000107.
- 925 Huson, D. H., & Bryant, D. (2006). Application of phylogenetic networks in evolutionary
926 studies. *Molecular biology and evolution*, 23(2), 254–267. doi: 10.1093/molbev/msj030.
- 927 Huyse, T., Poulin, R., Théron, A., & Theron, A. (2005). Speciation in parasites: a population
928 genetics approach. *Trends in Parasitology*, 21(10), 469–475. doi:
929 10.1016/j.pt.2005.08.009.
- 930 Imoto, J. M., Saitoh, K., Sasaki, T., Yonezawa, T., Adachi, J., Kartavtsev, Y. P., Miya, M.,
931 Nishida, M., & Hanzawa, N. (2013). Phylogeny and biogeography of highly diverged
932 freshwater fish species (Leuciscinae, Cyprinidae, Teleostei) inferred from mitochondrial
933 genome analysis. *Gene*, 514(2), 112–124. doi: 10.1016/j.gene.2012.10.019.
- 934 Johnson, K. P., Adams, R. J., & Clayton, D. H. (2002). The phylogeny of the louse genus
935 *Brueelia* does not reflect host phylogeny. *Biological Journal of the Linnean Society*,
936 77(2), 233–247. doi: 10.1046/j.1095-8312.2002.00107.x.
- 937 Jombart, T., & Collins, C. (2015). *Analysing genome-wide SNP data using adegenet 2.0. 0*.
- 938 Kalbe, M., Eizaguirre, C., Scharsack, J. P., & Jakobsen, P. J. (2016). Reciprocal cross
939 infection of sticklebacks with the diphyllbothriidean cestode *Schistocephalus solidus*
940 reveals consistent population differences in parasite growth and host resistance.

- 941 *Parasites & Vectors*, 9(1), 1–12. doi: 10.1186/s13071-016-1419-3.
- 942 Katoh, K., & Standley, D. M. (2013). MAFFT multiple sequence alignment software version
943 7: improvements in performance and usability. *Molecular Biology and Evolution*, 30(4),
944 772–780. doi: 10.1093/molbev/mst010.
- 945 Kennedy, C. R., Shears, P. C., & Shears, J. A. (2001). Long-term dynamics of *Ligula*
946 *intestinalis* and roach *Rutilus rutilus*: a study of three epizootic cycles over thirty-one
947 years. *Parasitology*, 123(3), 257–269. doi: 10.1017/S0031182001008538.
- 948 Kihedu, K. J., Mlay, M. K. L., Mwambungu, J. A., & Ngatunga, B. P. (2001). Drifting long
949 line, a potential fishing method for the northern part of Lake Nyasa/Malawi/Niassa. *Lake*
950 *Malawi Fisheries Management Symposium*, 121.
- 951 Kozlov, A. M., Darriba, D., Flouri, T., Morel, B., & Stamatakis, A. (2019). RAxML-NG: a
952 fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference.
953 *Bioinformatics*, 35(21), 4453–4455. doi: 10.1093/bioinformatics/btz305
- 954 Kuchta, R., & Scholz, T. (2017). Diphyllobothriidea Kuchta, Scholz, Brabec & Bray, 2008.
955 In *In: Planetary Biodiversity Inventory (2008-2017): Tapeworms from Vertebrate*
956 *Bowels of the Earth*.
957 <https://www.marinespecies.org/aphia.php?p=sourcedetails&id=283093>
- 958 Lagrue, C., Presswell, B., Dunckley, N., & Poulin, R. (2018). The invasive cestode parasite
959 *Ligula* from salmonids and bullies on the South Island, New Zealand. *Parasitology*
960 *Research*, 117(1), 151–156. doi: 10.1007/s00436-017-5684-7.
- 961 Landeryou, T., Rabone, M., Allan, F., Maddren, R., Rollinson, D., Webster, B. L., Tchuem-
962 Tchuente, L.-A., Anderson, R. M., & Emery, A. M. (2022). Genome-wide insights into
963 adaptive hybridisation across the *Schistosoma haematobium* group in West and Central
964 Africa. *PLoS Neglected Tropical Diseases*, 16(1), e0010088. doi:
965 10.1371/journal.pntd.0010088.
- 966 Lanfear, R., Frandsen, P. B., Wright, A. M., Senfeld, T., & Calcott, B. (2017). Partitionfinder
967 2: New methods for selecting partitioned models of evolution for molecular and
968 morphological phylogenetic analyses. *Molecular Biology and Evolution*, 34(3), 772–
969 773. doi: 10.1093/molbev/msw260
- 970 Langmead, B., & Salzberg, S. L. (2012). Fast gapped-read alignment with Bowtie 2. *Nature*
971 *Methods*, 9(4), 357–359. doi: 10.1038/nmeth.1923.
- 972 Leaché, A. D., Fujita, M. K., Minin, V. N., & Bouckaert, R. R. (2014). Species delimitation
973 using genome-wide SNP data. *Systematic Biology*, 63(4), 534–542. doi:
974 10.1093/sysbio/syu018.
- 975 Leigh, J. W., & Bryant, D. (2015). POPART: Full-feature software for haplotype network
976 construction. *Methods in Ecology and Evolution*, 6(9), 1110–1116. doi: 10.1111/2041-
977 210X.12410.
- 978 Levin, B. R., Lipsitch, M., & Bonhoeffer, S. (1999). Population biology, evolution, and

- 979 infectious disease: convergence and synthesis. *Science*, 283(5403), 806–809. doi:
980 10.1126/science.283.5403.806.
- 981 Li, L., Lü, L., Nadler, S. A., Gibson, D. I., Zhang, L.-P., Chen, H.-X., Zhao, W.-T., & Guo,
982 Y.-N. (2018). Molecular phylogeny and dating reveal a terrestrial origin in the early
983 carboniferous for ascaridoid nematodes. *Systematic Biology*, 67(5), 888–900. doi:
984 10.1093/sysbio/syy018.
- 985 Li, W. X., Fu, P. P., Zhang, D., Boyce, K., Xi, B. W., Zou, H., Li, M., Wu, S. G., & Wang,
986 G. T. (2018). Comparative mitogenomics supports synonymy of the genera *Ligula* and
987 *Digramma* (Cestoda: Diphyllbothriidae). *Parasites & Vectors*, 11(1), 1–11. doi:
988 10.1186/s13071-018-2910-9.
- 989 Librado, P., & Rozas, J. (2009). DnaSP v5: a software for comprehensive analysis of DNA
990 polymorphism data. *Bioinformatics*, 25(11), 1451–1452. doi:
991 10.1093/bioinformatics/btp187
- 992 Liu, X., & Fu, Y.-X. (2020). Stairway Plot 2: demographic history inference with folded SNP
993 frequency spectra. *Genome Biology*, 21(1), 1–9. doi: 10.1186/s13059-020-02196-9.
- 994 Loot, G., Aulagnier, S., Lek, S., Thomas, F., & Guégan, J.-F. (2002). Experimental
995 demonstration of a behavioural modification in a cyprinid fish, *Rutilus rutilus* (L.),
996 induced by a parasite, *Ligula intestinalis* (L.). *Canadian Journal of Zoology*, 80(4), 738–
997 744. doi: 10.1139/z02-043.
- 998 Loot, G., Lek, S., Brown, S. P., & Guégan, J.-F. (2001). Phenotypic modification of roach
999 (*Rutilus rutilus* L.) infected with *Ligula intestinalis* L.(Cestoda: Pseudophyllidea).
1000 *Journal of Parasitology*, 87(5), 1002–1010. doi: 10.1645/0022-
1001 3395(2001)087[1002:PMORRR]2.0.CO;2.
- 1002 Luo, H. Y., Nie, P., Yao, W. J., Wang, G. T., & Gao, Q. (2003). Is the genus *Digramma*
1003 synonymous to the genus *Ligula* (Cestoda: Pseudophyllidea)? *Parasitology Research*,
1004 89(5), 419–421. doi: 10.1007/s00436-002-0802-5.
- 1005 Mácová, A., Hoblíková, A., Hypša, V., Stanko, M., Martinů, J., & Kvičerová, J. (2018).
1006 Mysteries of host switching: Diversification and host specificity in rodent-coccidia
1007 associations. *Molecular Phylogenetics and Evolution*, 127, 179–189. doi:
1008 10.1016/j.ympev.2018.05.009.
- 1009 Maddison, W. P., & Maddison, D. R. (2021). *Mesquite: a modular system for evolutionary*
1010 *analysis. Version 3.70. <http://www.mesquiteproject.org>. 62, 1103–1118.*
- 1011 Malinsky, M., Matschiner, M., & Svardal, H. (2021). Dsuite-Fast D-statistics and related
1012 admixture evidence from VCF files. *Molecular Ecology Resources*, 21(2), 584–595. doi:
1013 10.1111/1755-0998.13265.
- 1014 Malinsky, M., Trucchi, E., Lawson, D. J., & Falush, D. (2018). RADpainter and
1015 fineRADstructure: population inference from RADseq data. *Molecular Biology and*
1016 *Evolution*, 35(5), 1284–1290. doi: 10.1093/molbev/msy023.

- 1017 Martinu, J., Hypsa, V., & Stefka, J. (2018). Host specificity driving genetic structure and
1018 diversity in ectoparasite populations: Coevolutionary patterns in Apodemus mice and
1019 their lice. *Ecology and Evolution*, 8(20), 10008–10022. <https://doi.org/10.1002/ece3.4424>
- 1020 Matthews, A. E., Barnett, C. J., & Boves, T. J. (2022). Differential survival and dispersal of
1021 avian feather mites with contrasting host specificities. *Ecological Entomology*, 47(5),
1022 864–871. doi: 10.1111/een.13176.
- 1023 Matzke, N. J. (2014). Model selection in historical biogeography reveals that founder-event
1024 speciation is a crucial process in island clades. *Systematic Biology*, 63(6), 951–970. doi:
1025 10.1093/sysbio/syu056.
- 1026 Matzke, N. J. (2013). BioGeoBEARS: BioGeography with Bayesian (and likelihood)
1027 evolutionary analysis in R Scripts. *R Package, Version 0.2, 1*, 2013.
- 1028 Minh, B. Q., Schmidt, H. A., Chernomor, O., Schrempf, D., Woodhams, M. D., von
1029 Haeseler, A., & Lanfear, R. (2020). IQ-TREE 2: New Models and Efficient Methods for
1030 Phylogenetic Inference in the Genomic Era. *Molecular Biology and Evolution*, 37(5),
1031 1530–1534. doi: 10.1093/molbev/msaa015
- 1032 Mussmann, S. M., Douglas, M. R., Chafin, T. K., & Douglas, M. E. (2020). A dmixPipe:
1033 population analyses in A dmixture for non-model organisms. *BMC Bioinformatics*,
1034 21(1), 1–9. doi: 10.1186/s12859-020-03701-4.
- 1035 Naughton, D. (2003). Annotated bibliography of Quaternary vertebrates of northern North
1036 America: with radiocarbon dates. *University of Toronto Press*.
- 1037 Nazarizadeh, M., Martinů, J., Nováková, M., Stanko, M., & Štefka, J. (2022).
1038 Phylogeography of the parasitic mite *Laelaps agilis* in Western Palearctic shows
1039 lineages lacking host specificity but possessing different demographic histories. *BMC*
1040 *Zoology*, 7(1), 1–16. doi: 10.1186/s40850-022-00115-y.
- 1041 Nazarizadeh, M., Peterka, J., Kubečka, J., Vašek, M., Jůza, T., de Moraes, K. R., Čech, M.,
1042 Holubová, M., Souza, A. T., & Blabolil, P. (2022). Different hosts in different lakes:
1043 prevalence and population genetic structure of plerocercoids of *Ligula intestinalis*
1044 (Cestoda) in Czech water bodies. *Folia Parasitologica*, 69, 18. doi:
1045 10.14411/fp.2022.018
- 1046 Nosil, P. (2012). Ecological Speciation. *Oxford University Press*, 1–304. doi:
1047 10.1093/acprof:osobl/9780199587100.001.0001.
- 1048 Otto-Bliesner, B. L., Marshall, S. J., Overpeck, J. T., Miller, G. H., Hu, A., & members, C. L.
1049 I. P. (2006). Simulating Arctic climate warmth and icefield retreat in the last
1050 interglaciation. *Science*, 311(5768), 1751–1753. doi: 10.1126/science.1120808.
- 1051 Patterson, N., Moorjani, P., Luo, Y., Mallick, S., Rohland, N., Zhan, Y., Genschoreck, T.,
1052 Webster, T., & Reich, D. (2012). Ancient admixture in human history. *Genetics*, 192(3),
1053 1065–1093. doi: 10.1534/genetics.112.145037.
- 1054 Perrot-Minnot, M., Špakulová, M., Wattier, R., Kotlik, P., Düßen, S., Aydoğdu, A., &

- 1055 Tougard, C. (2018). Contrasting phylogeography of two Western Palaearctic fish
1056 parasites despite similar life cycles. *Journal of Biogeography*, 45(1), 101–115. doi:
1057 10.1111/jbi.13118.
- 1058 Peterson, B. K., Weber, J. N., Kay, E. H., Fisher, H. S., & Hoekstra, H. E. (2012). Double
1059 digest RADseq: an inexpensive method for de novo SNP discovery and genotyping in
1060 model and non-model species. *PloS One*, 7(5), e37135. doi:
1061 10.1371/journal.pone.0037135.
- 1062 Petkevičiūtė, R. (1992). Comparative cytogenetics of *Diphyllbothrium ditremum* (Creplin,
1063 1925) and *Ligula intestinalis* (Linnaeus, 1758)(Cestoda: Pseudophyllidea). *Systematic*
1064 *Parasitology*, 23(3), 167–173. doi: 10.1007/BF00010869.
- 1065 Pickrell, J., & Pritchard, J. (2012). Inference of population splits and mixtures from genome-
1066 wide allele frequency data. *Nature Precedings*, 1. doi: 10.1371/journal.pgen.1002967.
- 1067 Pons, J., Barraclough, T. G., Gomez-Zurita, J., Cardoso, A., Duran, D. P., Hazell, S.,
1068 Kamoun, S., Sumlin, W. D., & Vogler, A. P. (2006). Sequence-Based Species
1069 Delimitation for the DNA Taxonomy of Undescribed Insects. *Systematic Biology*, 55(4),
1070 595–609. doi: 10.1080/10635150600852011
- 1071 Poulin, R. (2011). Evolutionary ecology of parasites. In *Evolutionary ecology of parasites*.
1072 Princeton university press.
- 1073 Prassack, K. A., Pante, M. C., Njau, J. K., & de la Torre, I. (2018). The paleoecology of
1074 Pleistocene birds from Middle Bed II, at Olduvai Gorge, Tanzania, and the
1075 environmental context of the Oldowan-Acheulean transition. *Journal of Human*
1076 *Evolution*, 120, 32–47. doi: j.jhevol.2017.11.003.
- 1077 Puillandre, N., Brouillet, S., & Achaz, G. (2021). ASAP: assemble species by automatic
1078 partitioning. *Molecular Ecology Resources*, 21(2), 609–620. doi: 10.1111/1755-
1079 0998.13281.
- 1080 Rahmouni, C., Vanhove, M. P. M., Koblmüller, S., & Šimková, A. (2022). Molecular
1081 phylogeny and speciation patterns in host-specific monogeneans (*Cichlidogyrus*,
1082 *Dactylogyridae*) parasitizing cichlid fishes (Cichliformes, Cichlidae) in Lake
1083 Tanganyika. *International Journal for Parasitology*, 52(6), 359–375. doi:
1084 10.1016/j.ijpara.2021.12.004.
- 1085 Rochette, N. C., Rivera-Colón, A. G., & Catchen, J. M. (2019). Stacks 2: Analytical methods
1086 for paired-end sequencing improve RADseq-based population genomics. *Molecular*
1087 *Ecology*, 28(21), 4737–4754. doi: 10.1111/mec.15253.
- 1088 Ronquist, F., & Huelsenbeck, J. P. (2003). MrBayes 3: Bayesian phylogenetic inference
1089 under mixed models. *Bioinformatics*, 19(12), 1572–1574. doi:
1090 10.1093/bioinformatics/btg180
- 1091 Shaw, K. L. (2002). Conflict between nuclear and mitochondrial DNA phylogenies of a
1092 recent species radiation: what mtDNA reveals and conceals about modes of speciation in
1093 Hawaiian crickets. *Proceedings of the National Academy of Sciences*, 99(25), 16122-

- 1094 16127. doi: doi: 10.1073/pnas.24258589.
- 1095 Šimková, A., Morand, S., Jobet, E., Gelnar, M., & Verneau, O. (2004). Molecular phylogeny
1096 of congeneric monogenean parasites (Dactylogyruis): a case of intrahost speciation.
1097 *Evolution*, 58(5), 1001–1018. doi: 10.1111/j.0014-3820.2004.tb00434.x.
- 1098 Šimková, A., Řehulková, E., Choudhury, A., & Seifertová, M. (2022). Host-Specific
1099 Parasites Reveal the History and Biogeographical Contacts of Their Hosts: The
1100 Monogenea of Nearctic Cyprinoid Fishes. *Biology*, 11(2), 229. doi:
1101 10.3390/biology11020229.
- 1102 Štefka, J., Hypša, V., & Scholz, T. (2009). Interplay of host specificity and biogeography in
1103 the population structure of a cosmopolitan endoparasite: Microsatellite study of *Ligula*
1104 *intestinalis* (Cestoda). *Molecular Ecology*, 18(6), 1187–1206. doi: 10.1111/j.1365-
1105 294X.2008.04074.x.
- 1106 Štefka, J., Hoeck, P. E. A., Keller, L. F., & Smith, V. S. (2011). A hitchhikers guide to the
1107 Galápagos: co-phylogeography of Galápagos mockingbirds and their parasites. *BMC*
1108 *Evolutionary Biology*, 11(1), 284. doi: 10.1186/1471-2148-11-284.
- 1109 Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., & Kumar, S. (2011). MEGA5:
1110 molecular evolutionary genetics analysis using maximum likelihood, evolutionary
1111 distance, and maximum parsimony methods. *Molecular Biology and Evolution*, 28(10),
1112 2731–2739. doi: 10.1093/molbev/msr121.
- 1113 Tao, W., Yang, L., Mayden, R. L., & He, S. (2019). Phylogenetic relationships of
1114 Cypriniformes and plasticity of pharyngeal teeth in the adaptive radiation of cyprinids.
1115 *Science China Life Sciences*, 62(4), 553–565. doi: 10.1007/s11427-019-9480-3.
- 1116 Techer, M. A., Roberts, J. M. K., Cartwright, R. A., & Mikheyev, A. S. (2022). The first
1117 steps toward a global pandemic: Reconstructing the demographic history of parasite host
1118 switches in its native range. *Molecular Ecology*, 31(5), 1358–1374. doi:
1119 10.1111/mec.16322.
- 1120 Thaenkham, U., Chaisiri, K., & Hui En Chan, A. (2022). Overview of Parasitic Helminth
1121 Diversity: How Molecular Systematics Is Involved BT - *Molecular Systematics of*
1122 *Parasitic Helminths* (U. Thaenkham, K. Chaisiri, & A. Hui En Chan (eds.); pp. 61–86).
1123 Springer Nature Singapore. doi: 10.1007/978-981-19-1786-8_3
- 1124 Tricou, T., Tannier, E., & de Vienne, D. M. (2022). Ghost lineages can invalidate or even
1125 reverse findings regarding gene flow. *PLoS Biology*, 20(9), e3001776.
1126 doi:10.1371/journal.pbio.3001776.
- 1127 Vitál, Z., Boross, N., Czeglédi, I., Preiszner, B., Erős, T., Molnár, K., Cech, G., Székely, C.,
1128 Sándor, D., & Takács, P. (2021). First genetically verified occurrence of *Ligula*
1129 *pavlovskii* outside its native range and characteristics of its infection in *Neogobius*
1130 *fluvialilis*. *Journal of Great Lakes Research*, 47(1), 236–241. doi:
1131 10.1016/j.jglr.2020.10.008.
- 1132 Wacker, S., Larsen, B. M., Karlsson, S., & Hindar, K. (2019). Host specificity drives genetic

1133 structure in a freshwater mussel. *Scientific Reports*, 9(1), 1–7. doi: 10.1038/s41598-019-
1134 46802-8.

1135 Wang, S., Wang, S., Luo, Y., Xiao, L., Luo, X., Gao, S., Dou, Y., Zhang, H., Guo, A., &
1136 Meng, Q. (2016). Comparative genomics reveals adaptive evolution of Asian tapeworm
1137 in switching to a new intermediate host. *Nature Communications*, 7(1), 1–12. doi:
1138 10.1038/ncomms12845

1139 Weigand, H., Weiss, M., Cai, H., Li, Y., Yu, L., Zhang, C., & Leese, F. (2017). Deciphering
1140 the origin of mito-nuclear discordance in two sibling caddisfly species. *Molecular*
1141 *Ecology*, 26(20), 5705–5715. doi: 10.1111/mec.14292

1142 Yoneva, A., Scholz, T., Młocicki, D., & Kuchta, R. (2015). Ultrastructural study of
1143 vitellogenesis of *Ligula intestinalis* (Diphylobothriidea) reveals the presence of
1144 cytoplasmic-like cell death in cestodes. *Frontiers in Zoology*, 12(1), 1–9. doi:
1145 10.1186/s12983-015-0128-7.

1146 Zarlenga, D. S., Rosenthal, B. M., La Rosa, G., Pozio, E., & Hoberg, E. P. (2006). Post-
1147 Miocene expansion, colonization, and host switching drove speciation among extant
1148 nematodes of the archaic genus *Trichinella*. *Proceedings of the National Academy of*
1149 *Sciences*, 103(19), 7354–7359. doi: 10.1073/pnas.0602466103.

1150 Zhang, J., Kapli, P., Pavlidis, P., & Stamatakis, A. (2013). A general species delimitation
1151 method with applications to phylogenetic placements. *Bioinformatics*, 29(22), 2869–
1152 2876. doi: 10.1093/bioinformatics/btt499.

1153 Zink, R. M., & Gardner, A. S. (2017). Glaciation as a migratory switch. *Science*
1154 *Advances*, 3(9), e1603133. doi: 10.1126/sciadv.160313.

1155 **Data Accessibility Statement***

1156 Raw sequence reads are deposited in the SRA (BioProject XXX). Individual genotype
1157 data are available on DataDryad (XXXX). MtDNA data are deposited to NCBI
1158 Nucleotide Database (XXXX). *Data will be made available prior to publication of the
1159 manuscript in a peer-reviewed journal.

1160 **Benefit-Sharing Statement**

1161 All authors took part in field-collecting samples for the research via a collaborative effort.

1162 **Author Contributions**

1163 MNaz. and JŠ defined the research objective and drafted the manuscript. MNov., PK and
1164 MNaz. performed laboratory analyses. MNaz. analysed the data under the supervision of JŠ
1165 and ET. All authors took part in sample collecting and read and approved the final version of
1166 the manuscript.

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1169 **Tables and Figures (with captions)**

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Table 1. Genetic characteristics of different parasite populations based on the mtDNA and ddRAD data sets. hap: Number of mitochondrial haplotypes, hd: Mitochondrial haplotype diversity, Pi: Mitochondrial nucleotide diversity, He: Expected heterozygosity, MLH: Multi-locus heterozygosity, Ho: Observed heterozygosity, Fis: Inbreeding coefficient.

Parasite lineages	Host order / genera	Geographic distribution	N (mtDNA/ddRAD)	mtDNA (Cytb + COI)			ddRAD					
				hap	hd	Pi	He	MLH	Pi (SD)	Ho	Fis (SD)	Private alleles
Lineage A	<i>Cypriniformes</i> / <i>Abramis, Blicca, Scardinus, Exos, Squalius, Rutilus, Phoxinus, Alburnus, Carassius</i>	Czech Republic, France, Germany, Ireland, Italy, Russia, United Kingdom, Ukraine, and Tunisia (introduced)	(69/65)	63	0.99	0.0095	0.079	0.045 (0.02)	0.057 (0.03)	0.050	0.15 (0.20)	8.11
Lineage B	<i>Cypriniformes</i> / <i>Rhodeus, Gobio, Pseudophoxinus, Barbus</i>	Algeria, Czech Republic, Ireland, Poland, Tunisia, Turkey	(18/7)	16	0.98	0.015	0.092	0.059 (0.02)	0.071 (0.03)	0.060	0.09 (0.15)	5.86
<i>L. alternans</i>	<i>Cypriniformes</i> / <i>Hemiculter, Squalius</i>	China, Iran, and Russia	(6/3)	6	1	0.015	0.0064	0.0046 (0.02)	0.0057 (0.02)	0.0005	-0.25 (0.14)	5.81
China	<i>Osmeriformes</i> / <i>Neosalanx</i>	China	(5/7)	4	0.90	0.0025	0.038	0.027 (0.004)	0.028 (0.001)	0.027	-0.01 (0.14)	6.73
Australia and New Zealand	<i>Galaxiiformes</i> / <i>Galaxias</i> <i>Gobiiformes</i> / <i>Gobiomorphus</i>	Australia and New Zealand	(11/6)	5	0.61	0.0196	0.0023	0.002 (0.02)	0.002 (0.04)	0.002	-0.08 (0.22)	5.80
<i>L. pavlovskii</i>	<i>Gobiiformes</i> / <i>Neogobius, Pomatoschistus</i>	Hungary, Germany, Poland, Ukraine	(10/17)	9	0.97	0.0029	0.104	0.043 (0.01)	0.089 (0.03)	0.044 (0.01)	0.49 (0.24)	9.8
East African Rift (EAR)	<i>Cypriniformes</i> / <i>Engraulicypris, Rastrineobola</i>	Kenya and Tanzania	(15/7)	9	0.91	0.0047	0.019	0.0145 (0.02)	0.015 (0.03)	0.014 (0.02)	- 0.005(0.24)	6
Nearctic	<i>Cypriniformes</i> / <i>Semotilus, Rhinichthys/ Couesius</i>	Canada, USA (Oregon)	(11/10)	8	0.92	0.0042	0.020	0.014 (0.003)	0.017 (0.001)	0.014	0.11 (0.25)	8.9
Central and South Africa (CSA)	<i>Cypriniformes</i> / <i>Barbus, Enteromius</i>	Democratic Republic of the Congo, Namibia and South Africa	(9/5)	6	0.83	0.0055	0.011	0.007(0.019)	0.008 (0.02)	0.007 (0.001)	0.02 (0.16)	5.80
Ethiopia	<i>Cypriniformes</i> / <i>Labeobarbus, Enteromius</i>	Ethiopia (Lake Tana)	(6/12)	6	1	0.0185	0.030	0.015 (0.02)	0.022 (0.03)	0.013 (0.004)	0.27(0.16)	12.8

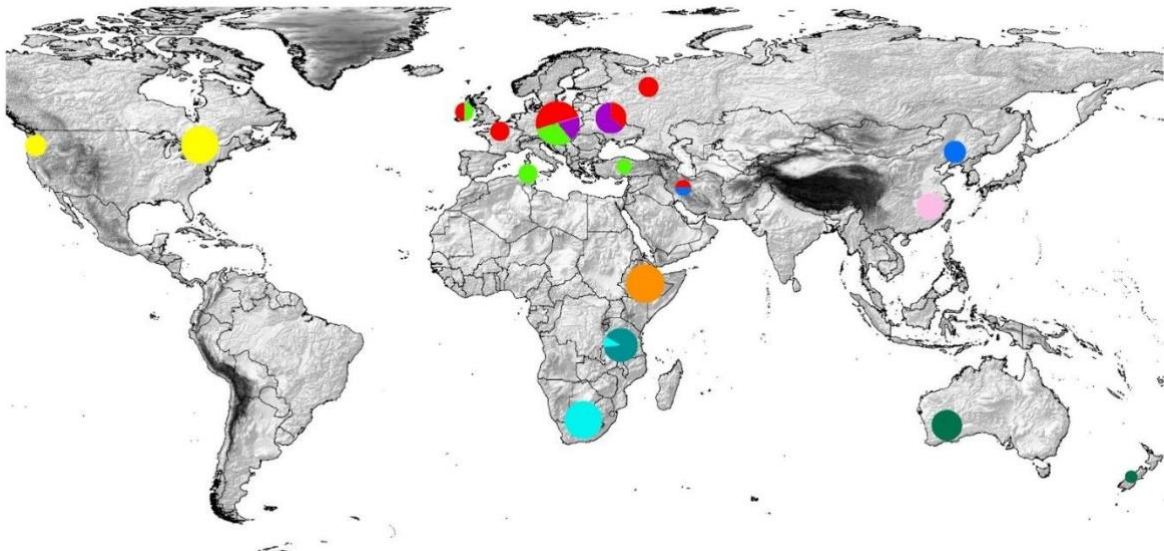


Fig. 1. *Ligula* sampling localities and patterns of genetic structure. Pie charts reflect the proportion of individuals assigned to each phylogenetic lineage (shown in Fig. 2); Lineage A (green), Lineage B (red), *L. alternans* (blue), China (light pink), Australia and New Zealand (dark green), *L. pavlovskii* (magenta), East African Rift (EAR; dark turquoise), Central and South Africa (CSA; light turquoise), Ethiopia (orange), and Nearctic (yellow).

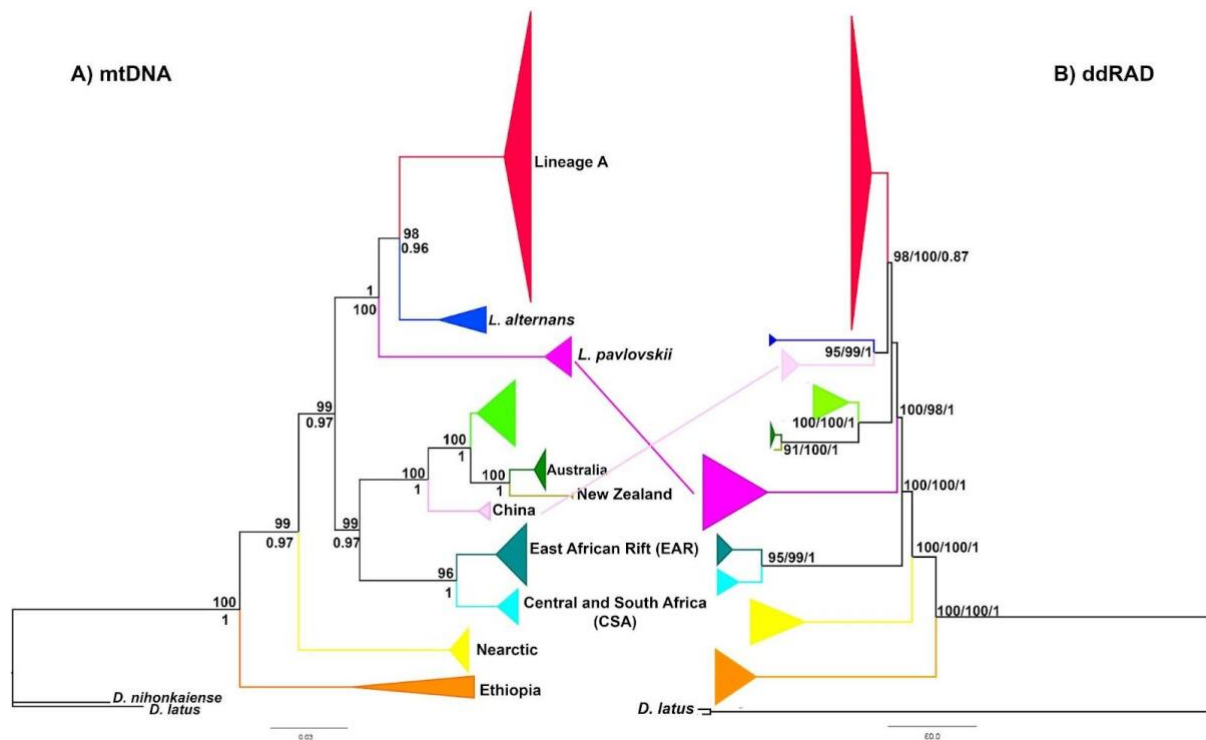


Fig. 2. Phylogenetic relationships of the *Ligula* species complex. A) Bayesian tree reconstructed using the concatenated mitochondrial genes (Cyt *b*, COI and ND1) using two outgroups (*D. latus* and *D. nihonkaiensis*). Lineage positions and branching patterns are concordant with the ML tree. Nodal supports at each node represent support values of BI (above branches) and ML (below branches). B) Reconstructed phylogenetic tree based on ddRAD. Bootstrap support values and Bayesian posterior probabilities are given at the nodes (RAxML/svdquartets/ExaBayes). Diagonal lines represent incongruences between the mtDNA and ddRAD topologies. Topology congruence tests demonstrated significant differences between the mtDNA and ddRAD trees ($P < 0.05$).

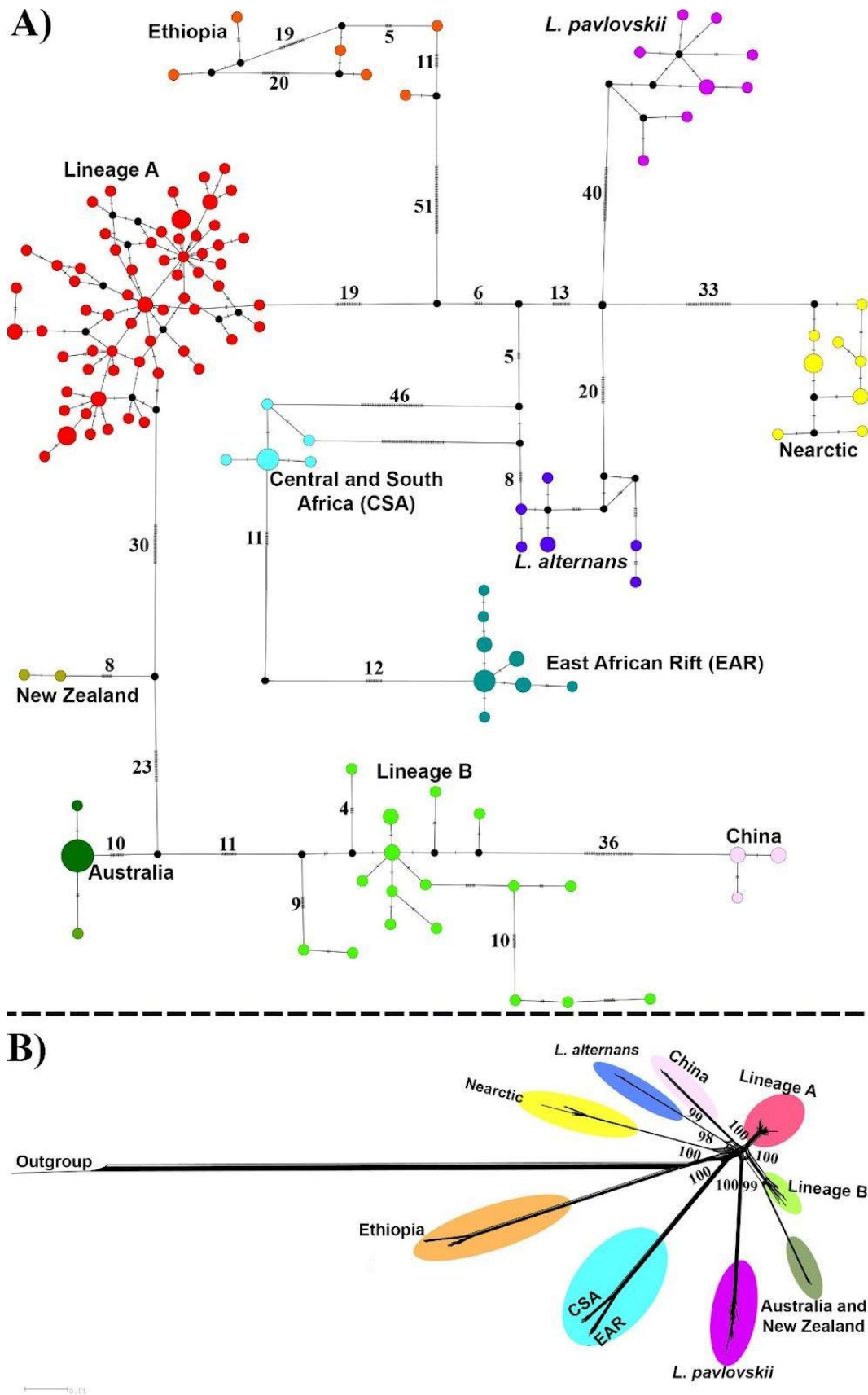


Fig. 3. Population relationships. A) Median-joining network showing relationships among the *Ligula* species complex, reconstructed based on the concatenated *Cyt b* and COI sequences (801 bp). Circle sizes are proportional to haplotype frequencies. Black circles indicate unsampled or extinct haplotypes. Mutational steps are displayed

by numbers and dashed symbols along each line. B) Phylogenetic network with bootstrap support values for the ddRAD data showing 10 distinct clusters in the *Ligula* species complex. *D. latus* was used as an outgroup.

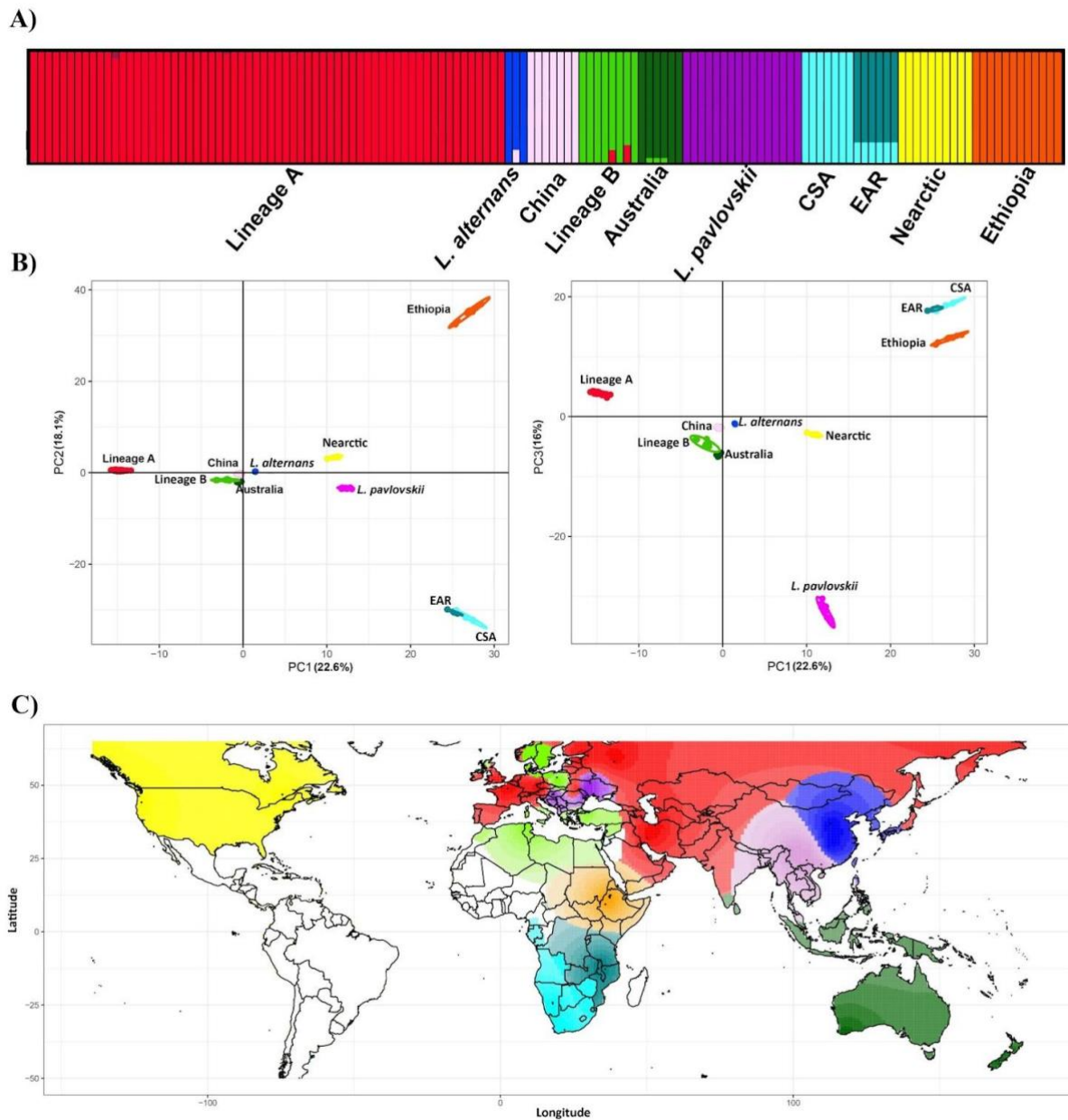


Fig. 4. Genetic structure of the populations. A) Population structure based on the ADMIXTURE analysis. Results are shown at $K = 10$. Each vertical (100%) stacked column indicates an individual representing the proportions of ancestry in K -constructed ancestral populations. B) PCA of *Ligula* sp. samples with 10937 SNPs, where the first three components explain 22.6%, 18.6% and 16% of the variance, respectively. C) Interpolated spatial genetic structure according to the individual ancestry coefficients of TESS3R. The colour gradient shows the degree of difference among individuals.

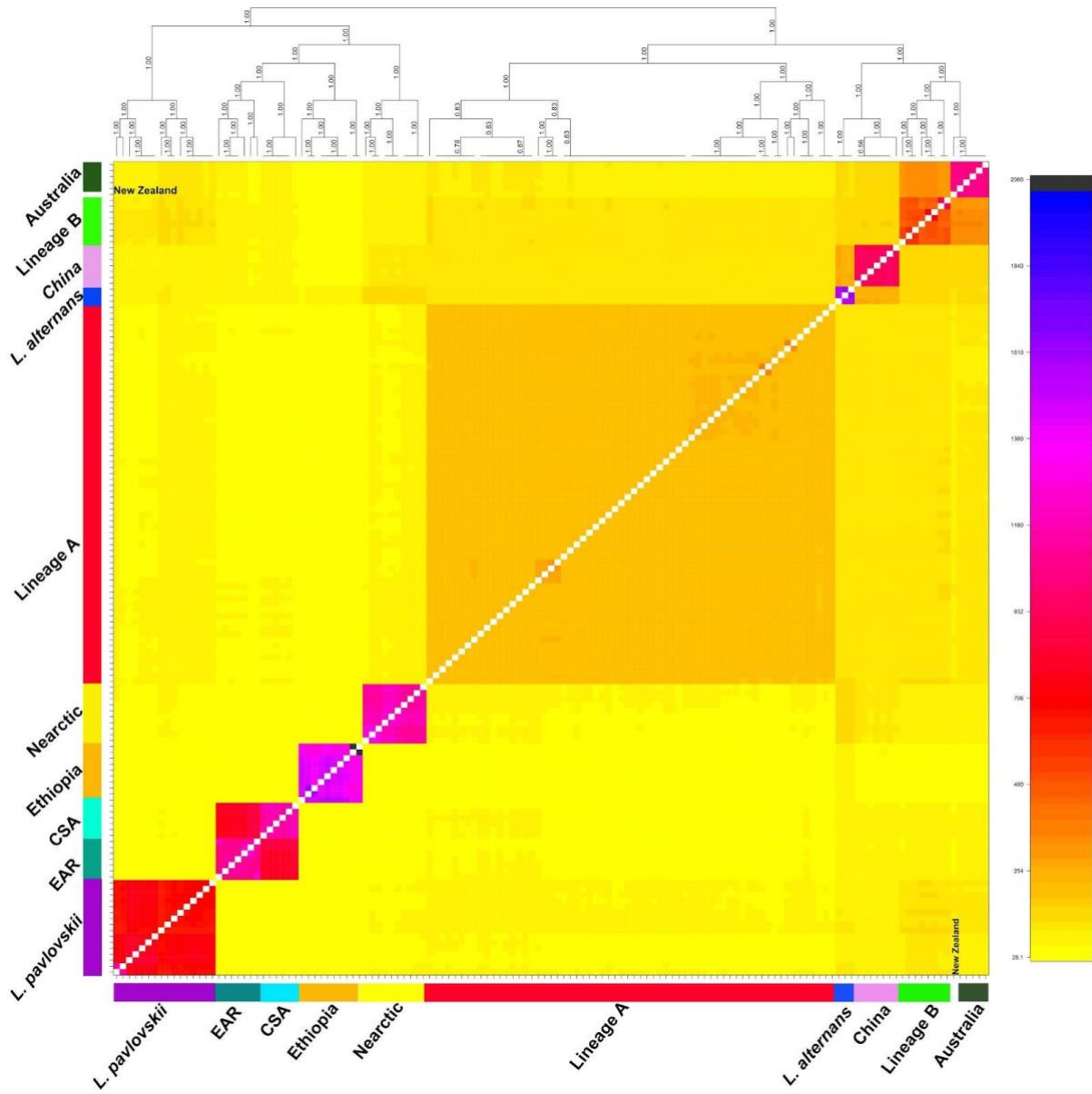


Fig. 5. Pairwise genetic similarity among individuals indicated by FineRADStructure co-ancestry matrix. Inferred populations are clustered together in their accompanying dendrogram.

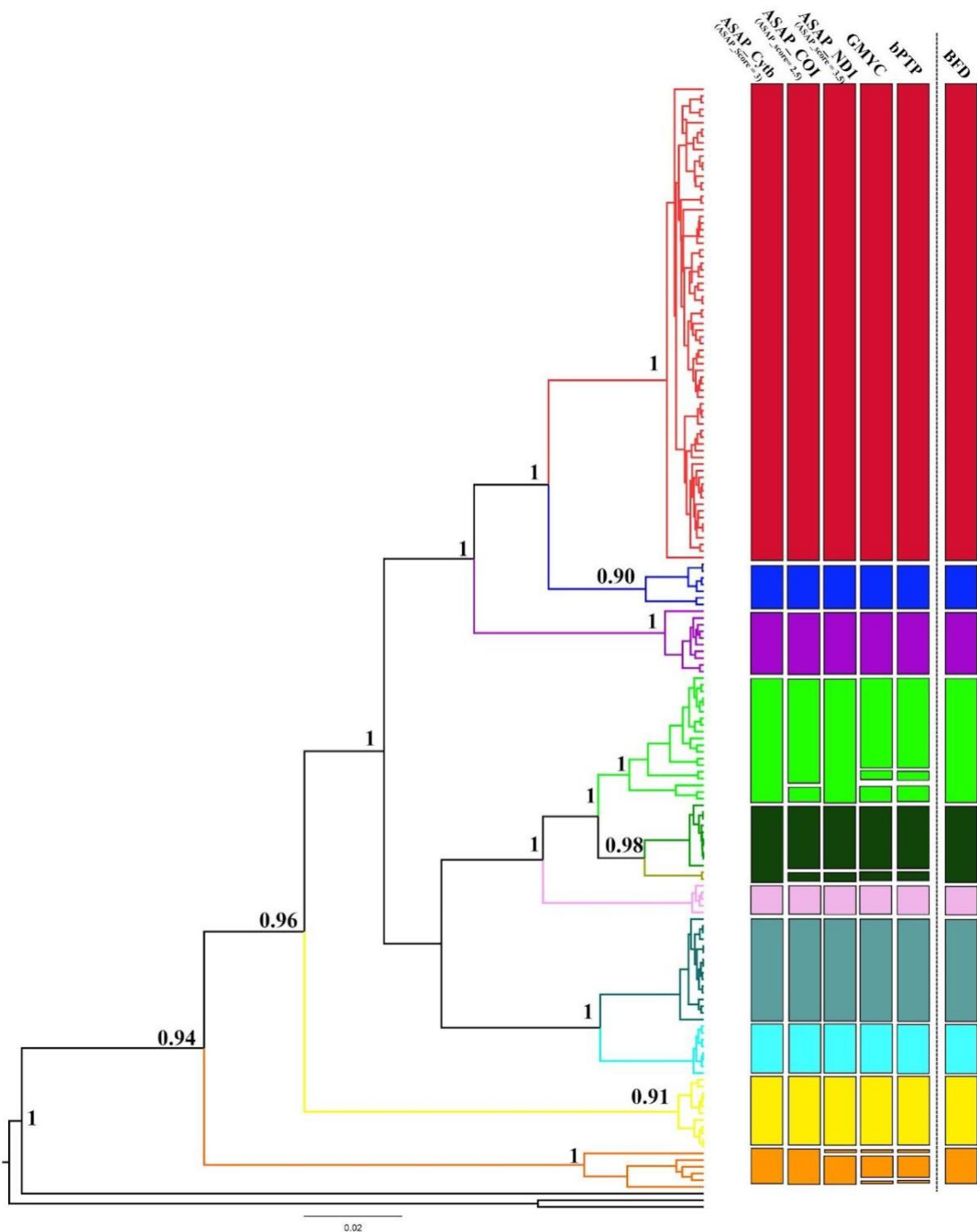


Fig. 6. Multi-approach species delimitation of *Ligula* species complex. Results of the mtDNA method (ASAP, GMYC, bPTP) are compared to the results of genome-wide SNP methods (BFD). Coloured bars depict results from different methods, indicating congruence among them.

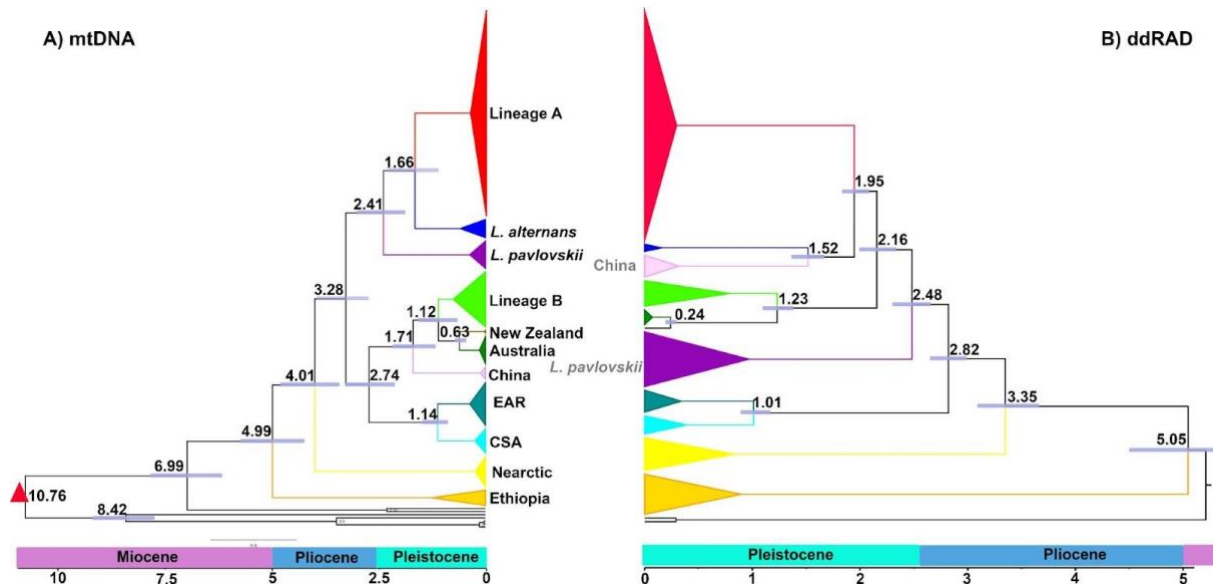
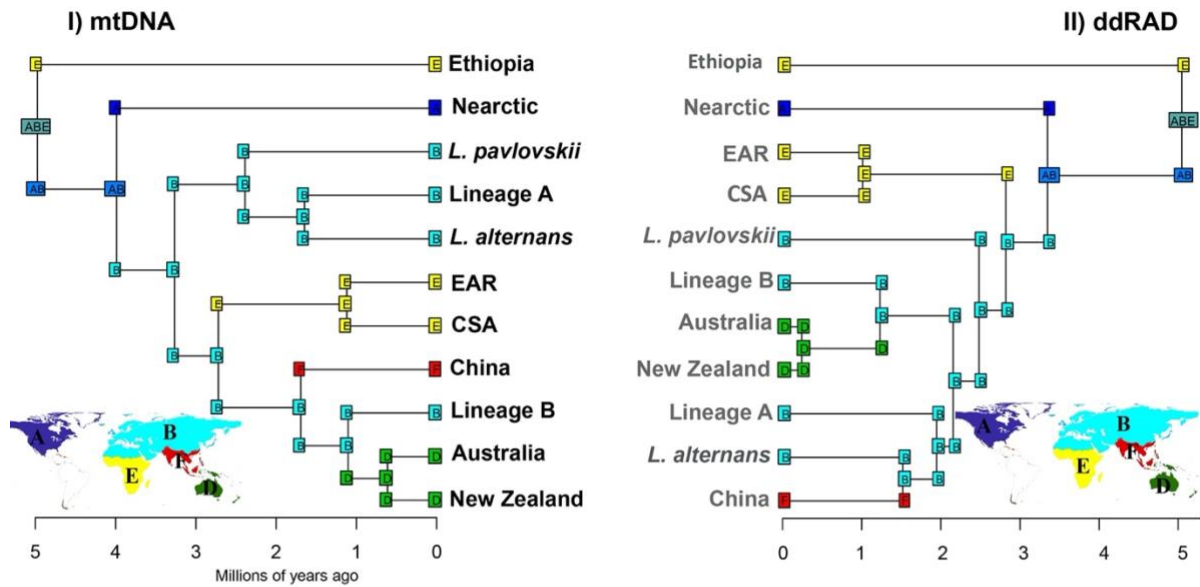


Fig. 7. Chronograms obtained from the dating analyses. A) Concatenated data set of three mitochondrial genes (*Cyt b*, *COI*, and *ND1*) containing 165 *Ligula* sequences. Numbers on each branch indicate divergence times. Calibration point is displayed by a red triangle. B) Divergence time estimated from the ddRAD data set (4471 unlinked SNPs). The bars on the nodes represent 95% HPD for divergence times.

A) Ancestral range estimation



B) Ancestral host evolution

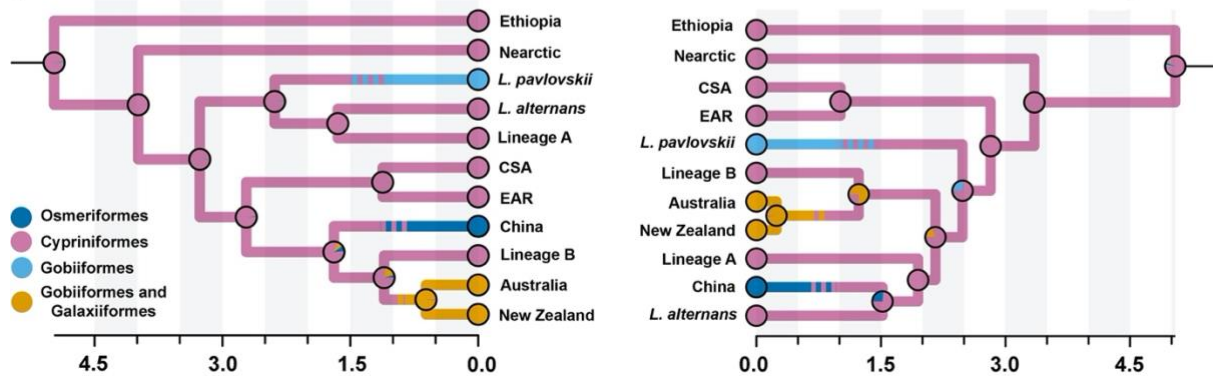


Fig. 8 Ancestral range and host estimations. A) Ancestral range estimation for the tapeworm chronograms in BioGeoBEARS under the DIVALIKE+J model. Squares at nodes indicate the highest ML probability in the area prior to the sudden speciation event, while squares on branches refer to the states of descendant lineages immediately after the speciation event. Squares showing more than one letter represent ancestral areas with more than one biogeographical area. B) Bayesian stochastic character map displaying the ancestral host evolution based on mtDNA (I) and ddRAD (II). Posterior probabilities for each node in different states are depicted by pie charts.

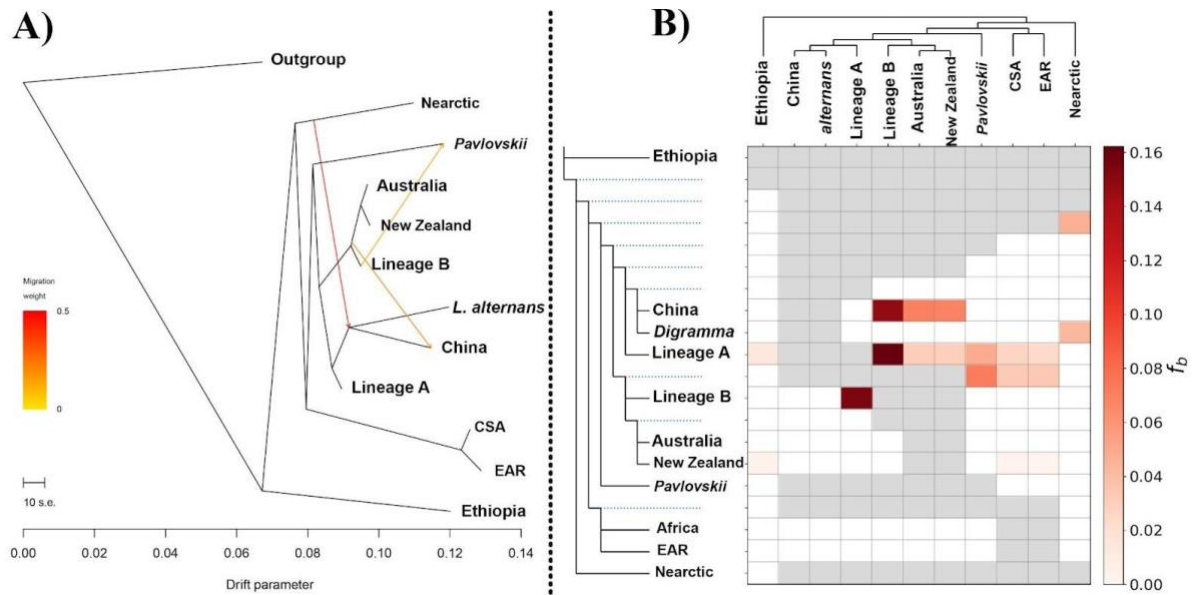


Fig. 9. Patterns of historical gene flow A) ML tree from Treemix, representing the relationships between the 11 *Ligula* lineages. The scale bar displays 10 times the average standard error for entries estimated in the covariance matrix. Migration arrows (events) are coloured based on their weight. The fraction of ancestry from the migration edge is referred to as migration weight. B) Results of genome-wide f_b -branch among the 11 *Ligula* lineages. Columns represent tips in the tree topology and rows indicate tree nodes. Cells display f_b statistics between nodes (rows) and tips (columns). When comparisons cannot be made, the cells are left empty and grey. F-branch (f_b (C)) statistics in our data suggest excess allele sharing among tips of the phylogenetic tree (representing lineages which are arranged horizontally at the top) and between each tip and node in the tree (arranged vertically on the left).

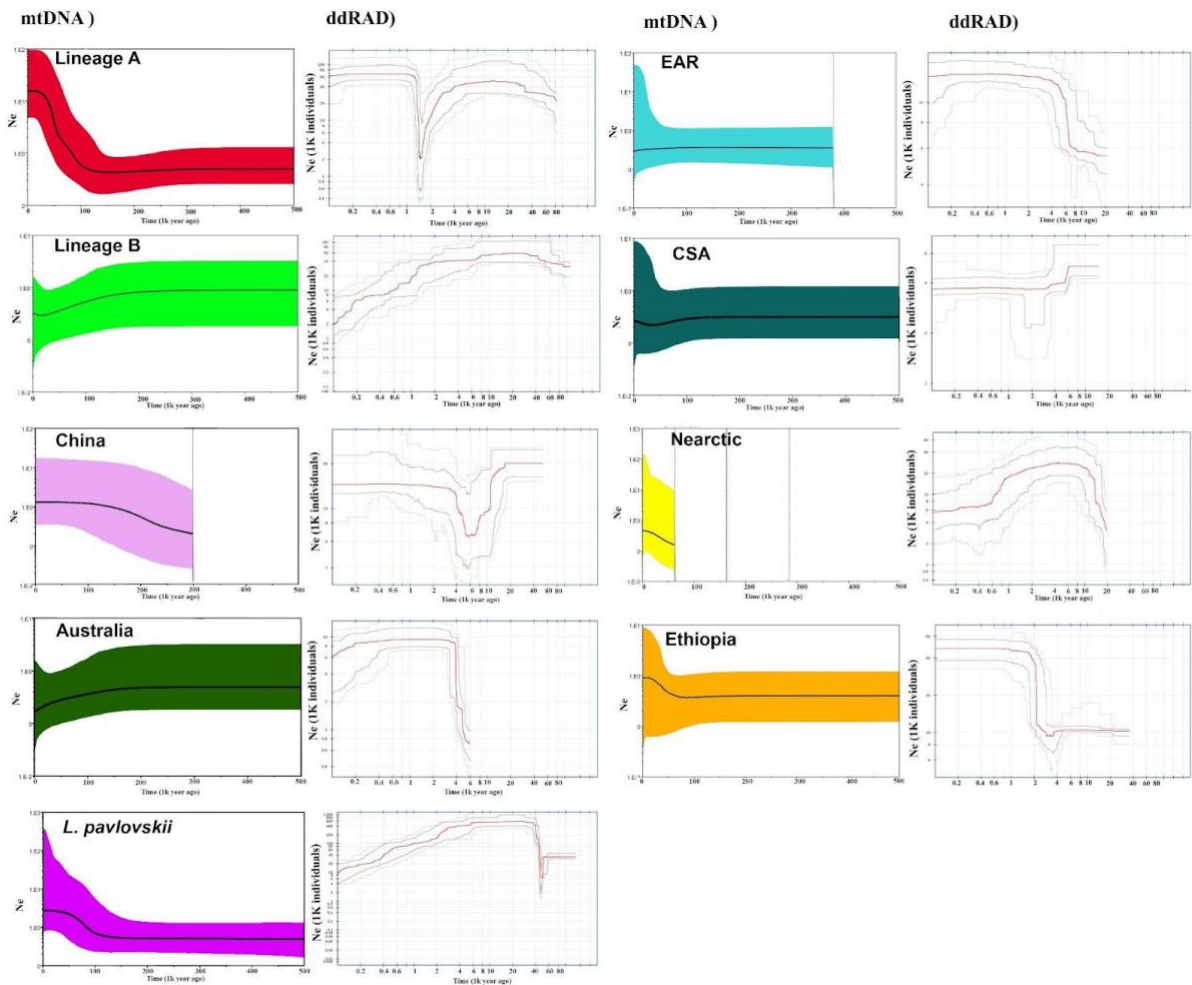


Fig. 10. Demographic history analyses. The plot on the left is EBSP based on two mtDNA genes (*Cyt b* and *COI*) for each main lineage. Bold line represents the median and the coloured area indicates 95% confidence interval. On the right, the stairway plot shows calculated changes in effective population size (N_e) for each lineage over time estimated from site frequency spectrum data. The grey lines above and below the red line (mean estimation) indicate 75% (dark) and 95% (light) confidence intervals.