## **Supplementary Information for**

The structure of a Tau fragment bound to tubulin prompts new hypotheses on Tau mechanism and oligomerization.



**Fig. S1.** Design of DARPin-Tau fusion proteins. (A) Design of D2-R3. The superposition of a  $\beta$  subunit of the microtubule decorated with Tau R2 (Left; pdb id 6CVN) to that of the tubulin:D2 DARPin complex (Right; pdb id 4F6R) leads to the model shown in the middle. (B) Schematic view and sequence of D2-R3. (C) Sequence alignment of R2 and R3 MTBRs (definition of the repeats as in Kellogg et al, Science, 360:1242–1246 (2018)). The Tau residues that are traced in the R2:microtubule cryo-EM model are in blue, those present in D2-R3 are highlighted in pink. The PHF6\* (in R2) and PHF6 (in R3) hexapeptides are in bold letters. Residue Cys322 (yellow) has been replaced by a Ser residue in D2-R3. (D) D2-R1, D2-R2 and D2-R4 were designed by replacing the R3 moiety in D2-R3 with the equivalent region of R1, R2 or R4 MTBR, respectively.



**Fig. S2.** Crystal packing of SLD1:tubulin:D2-R3. (A-C) The 2 complexes of the asymmetric unit. (A) View as in Fig. 1A. (B) View rotated 90°. (C) Close-up image of the part framed in panel B. (D-F) Same views but with two additional crystal-related asymmetric units. By virtue of crystal symmetry, the C-termini of the DARPins and of the SLD1 molecules (labeled in panel F) are in proximity to the N-termini of the Tau R3 stretches (schematized in panel G).



**Fig. S3.** Stereo view of the  $F_{obs}$ - $F_{calc}$  omit map contoured at the 3  $\sigma$  level (magenta). The final model for the Tau part is shown in cyan and green as in Fig. 1B.



**Fig. S4.** The NFVFGQ  $\beta$ -tubulin sequence, encompassing its S3  $\beta$ -strand, is also found in a bacterial phosphate-binding protein. (Top) Antiparallel interaction between the  $\beta$ -tubulin S3  $\beta$  strand and Tau R3. (Bottom) Antiparallel interaction of the NFVFGQ motif with a LVQI motif (magenta) in a *Stenotrophomonas maltophilia* phosphate-binding protein (pdb id 5J1D). The NFVFGQ motif is in cyan in both panels.

	D2-R3:tubulin:SLD1-R3
Data collection <sup>(a)</sup>	
Space group	P212121
Cell dimensions	
a, b, c (Å)	52.30, 222.25, 227.66
α, β, γ (°)	90, 90, 90
Resolution (Å)	159-2.21 (2.51-2.21)
Anisotropy resolution limits <sup>(b)</sup>	2.18, 2.63, 2.97
R <sub>meas</sub>	0.124 (3.96)
l / σ(l)	9.4 (1.8)
CC <sub>1/2</sub>	0.996 (0.607)
Completeness	
spherical (%)	60.6 (9.7)
ellipsoidal (%)	95.0 (68.4)
Multiplicity	27.4 (23.4)
Refinement	
Resolution (Å)	2.21
No. reflections	81,231
Rwork / Rfree	0.200 / 0.234
No. atoms	
Protein	16,810
Ligands	263
Solvent	328
B factors (Ų)	
Protein	45.1
Ligands	49.3
Solvent	30.0
Coordinate error (Å)	0.31
R.m.s.d.	
Bond lengths (Å)	0.008
Bond angles (°)	0.93
Ramachandran (%)	
Favored region	97.62
Allowed region	2.38
Outliers	0

 Table S1. Data collection and refinement statistics.

<sup>(a)</sup> Two dataset collected on the same crystal were merged. Values in parentheses are for the highest-resolution shell. <sup>(b)</sup> Determined by STARNISO using a local mean I /  $\sigma$ (I)=1.2.